

VITAMIN K₂

VITAL FOR HEALTH AND WELLBEING

CHAPTER 3 Vitamin K₂ Rich Food Products
by Muhammad Yasin, Masood Sadiq Butt
and Aurang Zeb

CHAPTER PUBLISHED BY

INTECH
open science | open minds

WORLD'S LARGEST
Science, Technology & Medicine Open Access book publisher.

KaQPa
BIOSCIENCE

CHAPTER PUBLISHED BY

INTECH

open science | open minds

World's largest Science,
Technology & Medicine
Open Access book publisher



2,900+

OPEN ACCESS BOOKS



100,000+

INTERNATIONAL
AUTHORS AND EDITORS



94+ MILLION

DOWNLOADS



BOOKS

DELIVERED TO
151 COUNTRIES

AUTHORS AMONG

TOP 1%

MOST CITED SCIENTISTS



12.2%

AUTHORS AND EDITORS
FROM TOP 500 UNIVERSITIES

This chapter was first published in March 2017 by **InTech** in the book
Vitamin K2 - Vital for Health and Wellbeing, Edited by Jan O. Gordeladze

Read, download and share for FREE

<http://www.intechopen.com/books/vitamin-k2-vital-for-health-and-wellbeing>

Abstract

Naturally, vitamin K exists in two bioactive forms mainly phyloquinone (vitamin K₁) and menaquinones (vitamin K₂). Phylloquinone is mostly found in green leafy vegetables such as kale, spinach, broccoli, and vegetable oils. However, menaquinones abundantly occurs in fermented vegetable products as menaquinones-7 (MK-7) and in animal-based products as menaquinone-4 (MK-4). Diverse concentrations of menaquinones are present in various dietary sources such as fermented pulses and milk-based products, cheese, meat, and animal organs. Presently, MK-7 and MK-4 contribute about 24 and 7%, respectively, of the total vitamin K dietary intake in the population consuming fermented products regularly. However, about 10% of menaquinones are pooled in the liver out of total intake of vitamin K. Conclusively, fermented soybean products and fermented milk-based products such as cheese and soured milk contain ample amount of MK-7, whereas animal organs, meat, fish, and egg contain appreciable amount of MK-4.

Keywords: vitamin K₂, menaquinones, fermented soybean, fermented milk, meat, cheese, poultry products

1. Introduction

Vitamin K is an indispensable anti-hemorrhagic fat soluble nutrient important for posttranslational modification of the proteins. Generally, these proteins are called vitamin K-dependent or Gla proteins. These coagulation proteins are produced in liver and played an active role in blood-clotting cascade. Moreover, during γ -carboxylation process, vitamin K hydroquinone is oxidized to its epoxide. When vitamin K is insufficient in blood-circulating system, carboxylation of vitamin K-dependent proteins is hampered and synthesis of undercarboxylated proteins is high that affects the Coagulation *cascade* parameters such as bleed and clotting times, and

prothrombin and partial prothrombin times. Additionally, inadequacy of vitamin K also affects the bone mineral density (BMD) that leads toward higher bone fracture rate among population especially postmenopausal women and increased the risk of coronary artery diseases [1, 2].

Vitamin K deficiency is one of the alarming dilemmas among newborns, teenagers, and postmenopausal women. Neonates and their nursing mothers are 33.3% and 65% deficit in vitamin K, respectively [3]. Moreover, gastrointestinal disorders, unnecessary use of antibiotics, and alcoholics are some potential causes of vitamin K deficiency that hinder the fat incorporation in the body, and thus absorption of this vitamin is reduced [4]. Similarly, continuous utilization of broad-spectrum antibiotics suppresses the synthesis of vitamin K in the intestinal gut, whereas antagonist drugs badly effect to vitamin K functioning that delays the blood coagulation and necessary modifies certain protein which are indispensable for bone health [5]. Likewise, inflammatory bowel disease individual also showed vitamin K deficiency prevalence that declined the bone mineral density possibly due to its malabsorption [6].

The ingestion of vitamin rich dietary sources is rational approach to manage the vitamin K deficiency population [7]. Additionally, supplementation of vitamin K in the diet is also improved the serum vitamin K concentration [8, 9]. In this context, pharmacological dose of vitamin K (45 mg/day) is effective to ameliorate problem of bone fractures [10]. Therefore, more osteocalcin carboxylation is attained via consumption of well above recommended dietary requirement amount of vitamin K [11].

Vitamin K predominantly, naturally, presents in the form of phyloquinone (vitamin K₁) and menaquinone (vitamin K₂). Primarily, phyloquinone is present in green leafy vegetables such as spinach, kale, broccoli, and certain vegetable oils [12]. However, menaquinones are present in the fermented soybean and animal products [8].

Fermented soybean (natto) is one of the richest sources of menaquinones principally menaquinone-7 (MK-7) (882–1034 µg/100 g) among other fermented plant and animal-based food products [8, 13]. Menaquinone-7 containing natto is prepared by the fermentation process with *Bacillus subtilis*. Raw soybean is converted into slimy and sticky food predominantly owing to glutamic residues [14]. This menaquinone is also available in other food products such as cheese, meat and meat-based products, and fermented milk. Menaquinones are also synthesized by intestinal microflora such as *Escherichia coli*, *Bacteroides vulgatus*, and *Bacteroides fragilis* [15].

From total vitamin K dietary intake, menaquinone-4 (MK-4), menaquinone-7 (MK-7), and phyloquinone contribute around 7, 24, and 60%, respectively [16]. Humans and laboratory animals have ability to transform phyloquinone into MK-4 and MK-7 by rearrangement of integral side chain that has the ability to strengthen the bone [17]. In both phyloquinone and menaquinones, total vitamin K is about 1.16 µg/kg body weight is required for normal functioning. Menaquinone-7 normally stored around 10% in the liver. However, only 5–25% of ingested vitamin K is catabolized to MK-4 followed by the conversion of menaquinones in the liver via prenylation [10].

2. Fermented soybean food products

Traditionally, *Bacillus subtilis* fermented soybean food products are consumed in different parts of world. Among the fermented soybean products, natto is attained special attention which is consumed in Japan, whereas similar products such as kinema is produced in India, Satlyangser and Bari in Bhutan, Thua-Nao in Thailand, Tao-si in Philippines, Douchi in China and Dawadawa in Nigeria, Pepok in Myanmar. Kinema is similar natto fermented soybean which is originated from south China and expanded in Nepal, India, Burma, Thailand, and Japan [18–20].

Soybean conquers special attention in the public owing to its protein and fat contents. Numerous cultivars are currently available for household consumption. Among, yellow, green, and black soybeans are commonly used after cooking or fermentation. *Bacillus* fermented products such as natto and cheonggukjang are also consumed [21, 22]. *B. subtilis* fermented soybean has slimy appearance, softer texture, and white goosy substance with distinctive rotten flavor. Moreover, fermentation also removes the beany odor of raw soybean completely and improves its hedonic response [19]. However, large number of population does not like its slimy appearance and powerful rotten odor of fermented soybean such as kinema, while other has enjoyable experience. Freshly prepared fermented soybean is consumed as a fried curry with boiled rice or mixed with vegetables or consumed with boiled rice, soup, and pickle [20].

Researchers are trying to identify the new strains that may have potential similar *B. subtilis* and have ability to produce ample amount of MK-7 in the soybean. In this context, *B. amyloliquefaciens* KCTC11712BP under its optimum conditions remarkably enhanced MK-7 concentration in cheonggukjang (fermented soybean) and has shorter production time than that of natto [23]. Menaquinones are produced by microorganisms during electron transport chain respiration process [24].

Moreover, natto is rich in menaquinones especially MK-7 followed by other menaquinones such as MK-5, MK-6, MK-8 which are ranged from 882–1034 $\mu\text{g}/100\text{ g}$, 7.1–7.8 $\mu\text{g}/100\text{ g}$, 12.7–14.8 $\mu\text{g}/100\text{ g}$, 78.3–89.8 $\mu\text{g}/100\text{ g}$, respectively [13]. Similarly, Berenjian et al. [25] also reported that MK-7 content in fermented soybean is varied from 800–900 $\mu\text{g}/100\text{ g}$. Moreover, Booth [26] also reported that natto contains 998 $\mu\text{g}/100\text{ g}$ MK-7. Previously, Dajanta et al. explicated that low fat (15.6%), high protein (45.8%), and small seed size of soybean are ideal characteristics of soybean for nato preparation [22]. Moreover, fermentation with *B. subtilis* exerts better nutritional attributes and sensory acceptability of the developed natto. Earlier, *B. subtilis* fermented Pakistani soybean also improved the concentration of MK-7 ranged from 681.35 to 803.82 $\mu\text{g}/100\text{ g}$. Contrary, MK-7 was not present in freshly cooked or reconstituted spinach [2]. The details of menaquinones concentration in various fermented soybean products are summarized in **Table 1**.

Food	Type	Country	MK-4	MK-5	MK-6	MK-7	References
Natto	Fermented soybean	Netherlands	7.5	13.8	998	84.1	[13]
Natto	Fermented soybean	Japan	-	-	-	939	[12]
Hikiwari natto	Chopped natto	Japan	ND	-	-	827	[12]
Natto	Fermented black soybean	Japan	-	-	-	796	[12]
Natto	Fermented soybean	Japan	ND	-	-	87–102	[23]
Natto	Fermented soybean	Pakistan	-	-	-	668–881	[2]
Cheonggukjang	Fermented soybean	Korea	ND	-	-	112–461	[23]
Cheonggukjang	Fermented soybean	Korea	74–76	-	-	271–1171	[23]
Cheonggukjang	Fermented soybean extract	Korea	-	-	-	1674–3438	[23]
Sauerkraut	Fermented vegetables	Netherlands	0.4	0.8	1.5	0.2	[13]
Cotton tofu	Hard type	Japan	0.04	-	-	-	[12]

ND = not detected; (-) = unknown or not reported.

Table 1. Menaquinones content of various fermented soybean ($\mu\text{g}/100\text{ g}$ or mL).

However, the level of MK-7 in natto is still less the recommended daily amount of $180\ \mu\text{g}/\text{day}$ that needs the utilization of $20\text{--}22\ \text{g}$ natto/day. Many consumers find natto unpalatable; therefore, the ingestion of sufficient MK-7 is impractical. In some countries such as Japan, China, Thailand, and India fermented soybean products are commonly consumed, whereas other consumers do not like the fermented soybean products due to unpalatable, slimy, and sticky in nature, so the ingestion of MK-7 is insufficient among the population. Furthermore, the digestion and utilization of MK-7 from fermented soybean products are less efficient in humans with aging. Hence, there is a need for production of concentrated, supplementary MK-7 in the diet [25]. The extracted MK-7 with oil is consumed in various functional foods to fulfill the recommended requirement of vitamin K.

2.1. Natto

Natto is a Japanese traditional food prepared by fermentation of soybean with *B. subtilis natto*. The sterilized soybeans are fermented at 40°C for $14\text{--}18\ \text{h}$ till it was fully covered with a viscous, sticky, and string-like material on surface. The resultant fermented soybean has characteristic odor and musty flavor [28].

2.2. Thua nao

For thua nao preparation, washed soybeans are soaked in water at ambient temperature for $16\ \text{h}$. Subsequently, soaked soybeans are autoclaved at 121°C for $40\ \text{min}$ and cooled to 55°C after removing water. Powder culture of *B. subtilis* is mixed and incubated at 42°C about $24\ \text{h}$ for fermentation. Afterwards, fermented soybeans are placed at 4°C to hamper the bacterial activity.

2.3. Kinema

Cleaned and pre-soaked soybeans are autoclaved at 121°C for 35 min and cooled to 30–35°C. The soybean is inoculated with active culture of *B. subtilis*. The inoculated soybeans are placed for fermentation at 37°C for 48 h at 85% RH. The activity of bacteria is reduced by using the refrigeration temperature for 12 h. Fresh kinema is immediately consumed because its shelf life is about 2–3 days in summer and 5–7 days in winter without refrigeration. After sun drying, it can be stored for several months [18].

2.4. Hawaii jar

Generally, small-sized soybean seeds are used for the production of hawaii jar. Soybean seeds are boiled without soaking and packed loosely in bamboo basket lined with fig or banana leaves. Natural fermentation process is completed within 3–5 days. Mixed microflora including *B. subtilis*, *B. cereus*, *B. licheniformis*, *Staphylococcus sciuri*, *Staphylococcus aureus*, *Providencia rettgeri*, *Alcaligenes* sp. are fermented organism present in the final product. Production of mucilage and ammonia flavor is the indicator of quality hawaii jar [18].

2.5. Cheonggukjang

Washed soybean seeds are soaked in tap water for 12 h. They are autoclaved at 121°C for 30 min and cooled up to 40°C prior to inoculation with pre-culture cell suspension of *Bacillus amyloliquefaciens strain KCTC 11712BP*. General for the preparation of cheonggukjang inoculated soybean seeds were placed in fermented for 48 h [28].

3. Menaquinones synthesizing organisms

Menaquinones are commonly synthesized by the action of intestinal microflora *B. vulgatus*, *B. fragilis*, and *E. coli* in human large intestine during electron transport chain reaction of respiration. Commercially, MK-7 was produced by action of bacteria such as *E. coli*, *Flavobacterium*, and *B. subtilis natto*. Moreover, among the fermented microflora, *B. subtilis* is considered one of the best options for the production of MK-7 due to its safety aspects as well as potential to synthesize the range of menaquinones such as MK-4 to MK-8 [15, 24, 29]. These hydrolytic bacteria are significantly consumed and reduced the indigestible oligo- and polysaccharides. The *Bacillus* species also lowered the activity of anti-nutrients of the soybean that hamper the availability of nutrients such as proteins and bioactive molecules [30].

B. subtilis is rod shaped, aerobic, heat resistant, spore forming Gram-positive bacteria. It has potential to synthesize number of enzymes such as proteolytic, amylolytic, and lipolytic enzymes during fermentation. The colonies of *B. subtilis* are irregular with hair-like structure having somewhat slimy appearance. Food Drug Administration granted the status of generally recognized as safe (GRAS) to the *Bacillus subtilis* and its close relative strains due to the lack of pathogenicity [20].

Administration of high single dose of MK-7 (2000 mg/kg) did not impart any toxic effect in animal modeling study in both genders. Moreover, prolonged treatment of MK-7 is considered also safe for human consumption due to its non-toxic effect on biochemical, hematological, urinary, and histopathological parameters [31].

Recently, Indian researcher Puri et al. identified a *B. subtilis* strain (*Bacillus subtilis* MTCC 2756) for the biosynthesis of menaquinone-7 for commercial purposes [32]. They found that addition of Tween 80 in fermentation medium significantly boost the MK-7 yield. Likewise, for the production of MK-7, groundnut meal is also used as a substrate owing to its protein and oil quality with strong minerals and vitamins profile using *Flavobacterium* sp. *SP-L-01* [33].

4. Stability of menaquinones

The vitamin K₂ was quite stable at room temperature for a period of up to 3 years. However, its concentration in final product was little affected by storage timed. Therefore, it is recommended that the menaquinone-7 rich product should be stored not more than 15°C temperature in a dry, cool, and dark place away from humidity, high heat, and sunlight.

5. Nutritional profile of fermented soybeans products

Fermented soybean is demonstrated higher amount of protein than raw soybean. The higher content of protein in fermented soybean might be elevated due to the microbial synthesis of single cell protein or rearrangements of molecules or enzymes followed by mortification of other moieties [19]. Premarani et al. explicated the impact of fermentation on the proximate composition of hawaijar [34]. They inferred that natural fermented soybean had 62.1% moisture, 26.02% soluble protein 24.36% crude fat, 8.2% crude fiber, 1.42% ash, and 3.8% free amino acids content nonetheless, inoculation with bacteria increases these parameters. However, hawaijar prepared from different soybean varieties showed non-momentous differences in lipid, fatty acid, and amino acid contents [35]. The nutritional profile of some fermented soybean was listed in **Table 2**.

Food	Moisture	Crude protein	Crude fat	Crude fiber	Ash	Carbohydrate	References
Kinema (traditional) dry basis	7.2	44.0	21.8	4.2	4.7	29.7	[79]
Kinema (pure culture) dry basis	6.8	45.1	23.0	4.8	4.5	27.4	[79]
Natto fresh weight basis	62.72	13.19	7.61	1.17	1.37	13.89	[2]
Fermented soybean	62.1	26.02	24.68	8.2	1.42		[34]
B. natto Itobiki	59–61	36–42	19–23	–	4.38–4.97	31–34	[37]
B. Natto NRRL B-3393	61	39–44	25–27	-	4.72–4.86	25–29	[37]

Table 2. Nutritional profile of menaquinones rich fermented soybean (%).

Likewise, natto has 40% protein and 24.68% fat contents. Controlled fermentation process improves the nutritional composition of natto than that of natural fermentation process [36]. Earlier, Wei and Chang delineated that the natto was prepared from four different soybean cultivars using *B. natto* "Itobiki" for moisture, protein, lipid, ash and carbohydrates as 59–61, 37–43, 20–23, 4–5, and 31–34%, respectively [37]. However, the natto synthesis through *B. natto* NRRL B-3393 exhibited the values ranged from 61–62, 39–44, 25–27, 4.72–4.86, and 25–29%, for these attributes.

Similarly, free amino acid contents were enhanced 60-fold in fermented products [38]. Free fatty acids impart significant role in the development of unique flavor in fermented soybean. Earlier, Kiuchi et al. reported that carbon length of fatty acids was momentarily increased during fermentation [39]. Moreover, the amount of vitamin B complex such thiamine, riboflavin, and niacin is momentarily improved during *B. subtilis* fermentation process of cooked soybeans. Additionally, total folate concentration was enhanced during fermentation process of Tempeh (fermented soybean). Fermented soybeans such as kinema and natto have appreciable amount of mineral contents such as iron, magnesium, copper, zinc, sodium, and calcium [40–42].

6. Antioxidant potential of fermented soybean

Fermented soybean had higher amount of total phenolic, antioxidant capacities, and flavonoid contents in thua nao produce by black soybeans by pure culture of *B. subtilis* TN51 compared with yellow soybeans and cooked non-fermented soybeans. Fermented black soybean and yellow soybean with *B. subtilis* TN51 exhibited higher amount of total phenolics 27.05 and 35.88 mgGAE/g extract than that of non-fermented products. Whereas total flavonoids contents of fermented black and yellow soybean were 3.50 and 1.38 mg catechin eq/g methanolic extract. However, contents of total phenolics and flavonoids were significant less in non-fermented black and yellow cooked soybeans. Likewise, 2,2-diphenylpicrylhydrazyl (DPPH)-antiradical activity (IC_{50} , mg/mL), and lipid peroxidation inhibition potential were higher in fermented black and yellow soybean compared with non-fermented cooked soybean [22].

Total phenol content of kinema was 144% higher than the raw soybean. Moreover, kinema has better free radical scavenger ability and metal chelating power and improved reducing power than that of non-fermented soybean. Therefore, it is suggested that kinema may has potential used as designer foods to alleviate oxidative stress [43]. Likewise, *B. subtilis* fermentation inclines to improve total polyphenolic and anthocyanins content about 10 and 250% during natto preparation from black soybeans, respectively. Moreover, DPPH radical scavenging activity has positive association with fermentation time and concentration of black soybean [19].

Fermentation increases about 58% content of phytosterol in kinema [43]. Later, Moktan et al. [44] reported that the kinema had 144%, 44%, 147%, and 92% higher total phenolics content, antioxidant activity, DPPH scavenging activity, and Fe^{2+} -chelating activity, respectively, compared to non-fermented cooked soybean. Similarly, better total phenolic contents and

DPPH activity were observed in *Bacillus* fermented soybean than that of soaked or cooked soybean [19].

During fermentation process of soybean, peptides are released by the hydrolysis of soybean proteins. Specific bioactive peptides such as glycinin and β -conglycinin are synthesized through the hydrolysis of soybean proteins. These bioactive peptides may act as regulatory compounds and have potential to minimize the physiological dysfunctions such anti-diabetic and anticancer activities [45].

Generally, fermented products are still widely synthesized by traditional methods. Consequently, it was recommended to develop standard operating producers and adhere with the good manufacturing practice (GMP) for individuals directly involved in its production for ensuring its safety [43].

7. Sensory response of fermented products

Sensory response including color, flavor, odor, texture, and overall acceptability is the main contributing parameters for the acceptance of products. Sensory evaluation of fermented soybean is mostly carried out using seven-point hedonic scale. The prepared natto obtained higher scores for color, appearance, taste, and viscosity than non-fermented cooked soybean [46]. Similarly, chungkukjang flavor, taste, and overall acceptability are evaluated using nine-point hedonic scale and reported that fermented soybean has high savory flavor and lower bitterness than traditional natto [47].

Moreover, soybean fermented with *B. subtilis* TN51 has superior aroma than that of conventionally fermented soybean [27]. In another study, sensory traits of natto, *that are*, color, aroma, stickiness, bitterness, sweetness, sourness, and chewiness were determined through continuous linear intensity-scale of 10 cm with multiple demographic panelists such as Chinese, Japanese, and American. There were non-significant variations observed in sweetness or sourness and flavor in commercial available natto and laboratory prepared similar product [37].

8. Animal-based vitamin K₂ food products

8.1. Meat

Concerning the vitamin K, only plants and fermented food commodities are considered as a major natural source, but limited attention has been paid to meat for its menaquinones contents. Recently, Rødbotten et al. reported that cattle meat such as Jersey and Norwegian Red have better amount of menaquinones predominately MK-4 [48]. Jersey meat has higher amount of MK-4 in *M. longissimus dorsi* (LD), *M. biceps femoris* (BF), and *M. psoasmajor* muscles compared to Norwegian Red Cattle. Some traces of MK-6 and MK-7 were also detected in both types of cattle meats. Moreover, it is suggested that vitamin K₂ content has no association with

intramuscular fat and tenderness of meat. Contrary, Fujiwara et al. reported that poultry birds rely on natto did not improve the menaquinones content of meat [49]. Previously, Elder et al. [50] quantified the MK-4 in beef, chicken, fish, and liver of calf available in retail outlets or fast food restaurants of various cities of USA and reported that MK-4 was present in chicken in substantial amount ranged from 6.3 to 22.1 $\mu\text{g}/100\text{ g}$ as compared to other tested animal meat and meat products (Table 3).

Meat	Type	Country	MK-4	MK-6	MK-7	Reference
Chicken	Breast meat	Netherlands	8.9	-	-	[13]
Chicken	Leg meat	Netherlands	8.5	-	-	[13]
Chicken	Thigh raw	Japan	ND	-	27 \pm 15	[12]
Chicken	Leg and thigh ng/g	Japan	600	ND	ND	[12]
Chicken	Fresh ng/g	Japan	89.9	ND	ND	[12]
Beef	Chuck, raw	Japan	0.6	-	15	[12]
	Beef raw	Netherlands	1.1	-	-	[13]
Beef	Fresh raw ground beef	USA	4.9	-	-	[50]
Beef	Fresh raw ground beef (medium fat)	USA	8.1	-	-	[50]
Beef	Fresh raw ground beef (high fat)	USA	7.4	-	-	[50]
Bovine beef	Fresh beef (ng/g)	Japan	34.3	0.3	0.3	[12]
Cattle (Jersey)	M. Biceps Femoris	Norway	4.85	0.004	0.006	[48]
	<i>M. psoas major</i>	Norway	2.46	-	-	[48]
	M. longissimus dorsi	Norway	3.39	-	-	[48]
Cattle (Norwegian Reg)	M. Biceps Femoris	Norway	3.02	0.006	0.082	[48]
	<i>M. psoas major</i>	Norway	1.82	-	-	[48]
	M. longissimus dorsi	Norway	2.43	-	-	[48]
Beef	Minced meat	Netherlands	6.7	-	-	[13]
Beef Product	Salami	Netherlands	9.0	-	-	[13]
Pork	Thigh, raw	Japan	ND	-	6 \pm 2	[12]
Pork	Fresh pork meat (ng/g)	Japan	9.4	0.3	0.3	[12]
Pork	Pork meat chop (ng/g)	Japan	31	ND	1.2	[12]
Horse	Fresh house meat ng/g	Japan	2.0	0.2	2.3	[12]
Luncheon	Meat	Netherlands	7.7	-	-	[13]
Hare	Leg meat	Netherlands	0.1	-	-	[13]
Deer	Back meat	Netherlands	0.7	-	-	[13]
Goose	Leg meat	Netherlands	31	-	-	[13]
Goose	Liver paste	Netherlands	369	-	-	[13]
Duck	Breast meat	Netherlands	3.6	-	-	[13]

Table 3. Menaquinones contents of meat of various fresh meat ($\mu\text{g}/100\text{ g}$).

Some studies indicated that thigh raw chicken meat contained the menaquinone-7 about 27 ng/g [21], whereas other only reported menaquinone-4 in both leg and breast meat of chicken ranged from 89.9 ng/g to 8.9 µg/100 g [13, 21]. Similarly, beef meat including raw, raw ground low, medium, and high fat meats from Japan, Netherlands, and USA contained menaquinone-4 ranged from 0.6 to 8.1 µg/100 g. However, Japanese origin beef meat contained menaquinone-7 as 15 µg/100 g. Likewise, cattle meat including Jersey and Norwegian *Biceps Femoris* meat merely contained MK-4, MK-6, and MK-7, whereas other parts did not contained MK-7, but their amount is very low [13, 21, 48, 50]. Additionally, pork and horse fresh meat contained the limited amount of MK-7 and all other tested meat such as luncheon, hare, deer goose, and duck contained only MK-4 (Table 3).

In various countries, people used the organs of animal as a source of meat. Therefore, concentration of vitamin K₂ is also very important to know in the commonly consumed organs such as liver, kidney, and heart. Hirauchi and coworkers reported that the organs meat of horse, chicken, and pork had significant amount of MK-4 compared with long-chain menaquinones (MK-7 to MK-13) and phylloquinone [51]. However, bovine liver was rich in MK-13 (215 ng/g) followed by MK-12 (215.6 ng/g), whereas the lowest concentration was noticed of MK-9 (15.3 ng/g). Other livers of various tested animal contained traces of higher menaquinones. The higher amount of long-chain menaquinones are possibly synthesized by gut microflora and stored in liver [52].

However, roasted beef contained 2–4 µg/100 g of MK-4, while other menaquinones such as MK-5, MK-7 and MK-8 were also present with low concentration [53]. Few publications are available regarding the vitamin K₂ content of meat (Tables 4 and 5). Previously, it was reported that beef meat has limited amount of vitamin K₂ without specifying the type of muscle and breeds [50, 53].

Meat	Type	Country	MK-4	MK-5	MK-6	MK-7	MK-8	Reference
Beef liver	Raw (µg/100 g)	USA	0.4	–	–	–	–	[50]
Beef liver	Pan-fried (µg/100 g)	USA	0.4	–	–	–	–	[50]
Beef liver	Braised (µg/100 g)	USA	1.9	–	–	–	–	[50]
Beef liver	Raw (ng/g)	USA	8.2	–	24.5	181.8	48.4	[51]
Beef liver	Raw (ng/g)	Finland	6.8	ND	9.44	25.6	13.8	[53]
Beef liver	Fresh heart (ng/g)	Japan	21.7	–	2.8	0.9	ND	[51]
Calf liver	Raw (µg/100 g)	USA	5.0	–	–	–	–	[50]
Calf liver	Pan-fried (µg/100 g)	USA	6.0	–	–	–	–	[50]
Calf liver	Braised (µg/100 g)	USA	1.1	–	–	–	–	[50]
Chicken Liver	Raw (ng/g)	Japan	39.6	–	0.3	ND	0.9	[51]
Chicken Liver	Raw (µg/100 g)	USA	14.1	–	–	–	–	[50]
Chicken Liver	Pan-fried (µg/100 g)	USA	12.6	–	–	–	–	[50]

Meat	Type	Country	MK-4	MK-5	MK-6	MK-7	MK-8	Reference
Chicken Liver	Braised ($\mu\text{g}/100\text{ g}$)	USA	6.7	-	-	-	-	[50]
Chicken heart	Fresh (ng/g)	Japan	142.6	-	0.1	ND	ND	[51]
Horse liver	Raw (ng/g)	Japan	2.1	-	1.0	2.3	1.2	[51]
Horse heart	Fresh heart (ng/g)	Japan	0.4	-	0.2	ND	ND	[51]
Pork heart	Raw (ng/g)	Finland	10.8	ND	ND	16	25	[53]
Pork liver	Fresh	Japan	1.2		0.2	1.1	ND	[51]
Pork	Raw	Netherlands	0.3	-	-	0.3	-	[13]
Pork	Raw Liver (ng/g)	Japan	5.9	-	0.4	6.1	5.6	[51]

ND = not detected; (-) = unknown or not reported.

Table 4. Vitamin K-2 content of different organ meats.

Food	Type	Country	MK-4	MK-5	MK-6	MK-7	Reference
Beef product	Hot dogs, regular fat	USA	5.7	-	-	-	[50]
Beef product	Ham roasted and pan broiled	USA	5.1	-	-	-	[50]
Beef product	Bacon (raw, pan-fried, microwaved, cooked and baked)	USA	5.6	-	-	-	[50]
Beef product	Beef meat roasted (ng/g)	Finland	28	1.2	ND	1.17	[53]
Beef product	Beef products	USA	1.7-8.1	-	-	-	[50]
Beef product	Roasted beef		2-4	-	-	-	[53]
Beef product	Broiled ground beef (low-fat steak)	USA	1.7	-	-	-	[50]
Beef product	Broiled ground beef (medium fat)	USA	7.2	-	-	-	[50]
Beef product	Broiled ground beef (high fat)	USA	5.1	-	-	-	[50]
Pork product	Loin (raw, broiled, pan-broiled, braised)	USA	0.9	-	-	-	[50]
Pork product	Meat franks, regular fat	USA	9.8	-	-	-	[50]
Pork product	Pork steak	Netherlands	2.1	-	-	0.5	[13]

ND = not detected; (-) = unknown or not reported.

Table 5. Vitamin K-2 content of different animal meat-based products ($\mu\text{g}/100\text{ g}$).

8.2. Animal-based sea foods

Fishes such as rainbow trout contained MK-4 (31 ng/g), MK-5 (0.9 ng/g), and MK-7 (2.0 ng/g), whereas MK-6 and MK-8 were not present (**Table 6**). Similarly, pike-perch also contained these menaquinones along with MK-6. Baltic herring and salmon only contained MK-4. Moreover, plaice and eel contained MK-4 as 0.2 and 1.7, MK-6 as 0.3 and 0.1, and MK-7 as 1.6 and 0.0, respectively. Horse mackerel from Netherlands and Japan only contained MK-4 content (0.4 and 0.6 µg/100 g). Furthermore, shrimp also had MK-4 (0.2 µg/100 g). Canned crab and tilapia fillets did not contain any form of menaquinones [13, 21, 50].

Food	Type	Country	MK-4	MK-5	MK-6	MK-7	MK-8	Reference
Fishes	Rainbow trout (ng/g)	Finland	31	0.9	ND	2.0	ND	[53]
	Pike-perch (ng/g)	Finland	1.9	0.49	0.52	4.9	ND	[53]
	Baltic herring (ng/g)	Finland	2.07	-	ND	ND	ND	[53]
Fish	Mackerel	Netherlands	0.4	-	-	-	-	[13]
	Plaice	Netherlands	0.2	-	0.3	0.1	1.6	[13]
	Eel	Netherlands	1.7	-	0.1	0.4	-	[13]
	Salmon	Netherlands	0.5	-	-	-	-	[13]
Fish	Horse mackerel, raw	Japan	0.6	-	-	ND	-	[12]
	Mackerel, raw	Japan	1.0	-	-	ND	-	[12]
Crab	Canned	USA	ND	-	-	-	-	[50]
Shrimp	Cooked and canned	USA	0.2	-	-	-	-	[50]
Salmon	Raw, Alaska wild	USA	0.3	-	-	-	-	[50]
Tilapia fillets	Raw and baked	USA	ND	-	-	-	-	[50]

ND = not detected; (-) = unknown or not reported.

Table 6. Menaquinones contents of various sea foods (µg/100 g).

8.3. Milk

Fresh milk having varied amount of fat also contained some amount of menaquinones especially MK-4. Sheep and cow whole milk contained about 17.4 and 8.60 ng/g of MK-4, respectively, while menaquinones were not detected in goat and donkey milk [54]. Moreover, milk having 1% fat had 0.4 µg/100 g of MK-4, whereas milk with higher amount of fat (2%) and whole milk showed more MK-4 contents as 0.5 and 1.0 µg/100 g which is available in retail outlets of USA [50].

There are various microorganisms such as *L. lactis* subsp. *cremoris*, *L. lactis* subsp. *Lactis*, and *Leuconostoc lactis* which have potential to produce long-chain menaquinones like MK-7 to

MK-10 about 230 nmol/g of dry cells. Moreover, these strains also have capacity to synthesis the ample amount of long-chain menaquinones in reconstituted non-fat dry milk and soy milk. Therefore, the milk-based fermented foods have significant amount of menaquinones and considered as an important dietary sources of vitamin K₂ [55]. Earlier, it was documented that the human milk only contained phyloquinone, but menaquinones were not detected [56].

Some of the fresh milk only contained MK-4 and other menaquinones are not detected or contained in the whole milk with varied concentration of fat from different animals (Table 7). Likewise, various creams and dressing also had MK-4 as source of vitamin K₂. Interestingly, fermented milk and sourced milk contained higher amount of long-chain menaquinones such as MK-6, MK-7, MK-8, and MK-9 while MK-4 and MK-5 were not present. Similarly, butter milk also contained MK-4 to MK-8 but in limited quantity, butter contained only MK-4 (15 µg/100 g).

Food	Type	Country	MK-4	MK-5	MK-6	MK-7	MK-8	MK-9	Reference
Fresh Milk	1% fat	USA	0.4	-	-	-	-	-	[50]
Fresh Milk	2% fat (Regular and chocolate)	USA	0.5	-	-	-	-	-	[50]
Fresh Milk	Whole milk	USA	1.0	-	-	-	-	-	[50]
Fresh Milk	Cow 3.5% fat (µg/L)	Italy	8.60	-	-	-	-	-	[54]
Fresh Milk	Buffalo 5.0% fat (µg/L)	Italy	ND	-	-	-	-	-	[54]
Fresh Milk	Sheep 5.5% fat (µg/L)	Italy	17.4	-	-	-	-	-	[54]
Fresh Milk	Goat 5.0% fat (µg/L)	Italy	ND	-	-	-	-	-	[54]
Fresh Milk	Donkey 1.0% fat (µg/L)	Italy	ND	-	-	-	-	-	[54]
Fresh Milk	Whole milk	Japan	2.03	-	-	ND	-	-	[12]
Fresh Milk	Whole milk	Netherlands	0.8	0.1	-	-	-	-	[13]
Yoghurt	Yogurt plain (ng/g)	Finland	3.6	1.01	ND	ND	ND	ND	[53]
Yoghurt	Whole milk	Japan	0.6	0.1	0	0.2	-	-	[51]
Yoghurt	Skimmed milk	Japan	0	0	0	0.1	-	-	[51]
Yoghurt	Yogurt, plain (whole milk)	Japan	1.0	-	-	0.1	-	-	[12]
Yoghurt	Whole yoghurt	Netherlands	0.6	0.1	-	-	0.1	-	[13]
Yoghurt	Fortified MK-7	Netherlands	-	-	-	11.65	-	-	[57]
Cream	Ice cream Regular fat (vanilla and chocolate)	USA	2.6	-	-	-	-	-	[50]

Food	Type	Country	MK-4	MK-5	MK-6	MK-7	MK-8	MK-9	Reference
Cream	Cream	Japan	8	-	-	ND	-	-	[12]
Cream	Whipping cream	Netherlands	5.4	-	-	-	-	-	[13]
Dressing	Mayonnaise (whole egg type)	Japan	17	-	-	ND	-	-	[12]
Dressing	Mayonnaise (egg yolk type)	Japan	38	-	-	ND	-	-	[12]
Chocolate	Market (brand or type is no specified)	Netherlands	1.5	-	-	-	-	-	[13]
Buttermilk	Market (brand or type is no specified)	Netherlands	0.2	0.1	0.1	0.1	0.6	-	[13]
Butter	Market (brand or type is no specified)	Netherlands	15.0	-	-	-	-	-	[13]
Soured milk (ng/g)		Finland	5.7	2.93	1.7	4.1	20.1	47	[53]
Fermented milk	Mesophilic fermented milk (MFM) ng/g	France	-	-	1.3-4.9	1.2-6.1	7.237.9	29-145	[58]
Fermented milk	MFM (ng/g)	Germany	-	-	2.1-6	4.1-6.3	31-42	88.4-198.5	[58]
Fermented milk	MFM (ng/g)	Poland	-	-	0.5-11.93	2-10.97	1-89.317	414.2	[58]

ND = not detected; (-) = unknown or not reported.

Table 7. Menaquinones contents of various dairy products (µg/100 g).

Recently, Knapen et al. delineated that vitamin K fortified foods are healthy choice to increase the nutritional intake of MK-7 [57]. The fortified yoghurt drink containing MK-7 about 28 µg/ml has similar absorption pattern as the soft gel containing same amount of pure menaquinone-7. It is therefore suggested that to fortify food products that are ideal choice among the public to enhance the nutritional intake of menaquinones in the body.

8.4. Yoghurt

Thermophilic bacteria such as *Lactobacillus delbrueckii*, *Streptococcus thermophilus*, and *Bifidobacterium* which are mostly used a lactic acid bacteria starter culture not have ability to produce the menaquinones. Therefore, yogurt type milk-based products prepared with pure culture of thermophilic bacteria have limited or no menaquinones. However, among other milk-based fermented products, 60% contained some amount of vitamin K₂ (Table 7). The mesophilic lactic acid bacteria species which as used as starter culture for the fermentation have a capability to produce ample amount of vitamin K₂[53, 58]. However, yoghurt including plain, whole and

skimmed milk contained higher amount of MK-4, whereas very low concentration of other menaquinones such as MK-5, MK-6 and MK-7 was present (Table 7).

Fortified yoghurt drink with MK-7 significantly improved serum concentration from 0.38 ng/ml to 2.00 ng/ml, whereas yoghurt supplemented with MK-7 along with other vitamins increased better serum MK-7 level as 2.17 ng/ml. Fortified MK-7 yoghurt and soft gel containing MK-7 showed statistically non-significantly variations [57].

8.5. Cheeses

Soft cheese as well as blue cheese have tremendous amount of vitamin K₂ as 1100 and 700 ng/g [58]. Earlier, menaquinones content of these cheeses has never been reported because soft and blue cheeses were not evaluated in respect of their vitamin K₂ content. These cheeses have higher amount of menaquinones possibly due to the activity of lactic acid bacteria particularly *Leuconostoc* species and yeasts or molds which are involved in ripening of cheeses. Most of the soft (from table name) and blue cheeses characterized had high contents of vitamin K₂ [59]. Generally, MK-9 was present in soft and blue cheese almost fourfold than that of MK-8. It is suggested that *lactococci* is responsible for the production of MK-9 in the dairy products. However, these bacteria also produced MK-8 comparatively constant ratio. Likewise, *propionibacteria* also synthesized the MK-9 as a key menaquinone in cheese. Menaquinone-9 concentration was higher in Norwegian Jarlsberg cheese trailed by Swiss Emmental cheese. However, Appenzeller or Gruyère cheeses had extremely low concentrations of MK-9. Additionally, Comte and Raclette cheeses contained lesser amount of MK-9 than both Jarlsberg and Emmental cheeses (Table 8).

Food	Type	Country	MK-4	MK-5	MK-6	MK-7	MK-8	MK-9	Reference
Cheeses	Hard cheeses (µg/100g)	Nether lands	4.7	1.5	0.8	1.3	16.9	-	[13]
	Soft cheese (µg/100g)	Nether lands	3.7	0.3	0.5	0.5	1.0	-	[13]
	Curd cheese (µg/100 g)	Nether lands	0.4	0.1	0.2	0.3	5.1	-	[13]
Semi-hard cheese	Semi-hard cheese (ng/g)	Nether lands	-	-	14.5–34.5	0–14.1	33.9–73.1	100–321	[58]
	Semi-hard cheese (ng/g)	Denmark	-	-	16.1–19.8	7.1–13.5	25–35.8	115.3–185.1	[58]
	Semi-hard cheese (ng/g)	Poland	-	-	9.8–15.8	ND	27.8–56.4	124.5–166.3	[58]
Soft cheese	Soft cheese (ng/g)	France	-	-	13.7–25.9	0–17.1	89.2–139.9	176.1–939.7	[58]
Cheese	Edam type (ng/g)	Finland	33	10.2	5.6	12.6	105	300	[53]
	Emmental type (ng/g)	Finland	52.3	ND	Traces	Traces	ND	Nd	[53]
Blue cheese	Blue cheese (ng/g)	France	-	-	14.4–35.4	24.6	59.8	189–230	[58]
	Blue cheese (ng/g)	England	-	-	96.7	223	103	301	[58]
Cheddar	Hard cheddar (ng/g)	England	-	-	8.7–29.9	0–23.1	10.5–61.8	0–66.9	[58]

Food	Type	Country	MK-4	MK-5	MK-6	MK-7	MK-8	MK-9	Reference
	Cheshire hard cheese (ng/g)	England	-	-	15.7	Nd	57.9	241	[58]
Leicester	Leicester hard cheese (ng/g)	France	-	-	20	21.5	47.6	162.4	[58]
Cheese	Appenzeller (ng/g)	Swiss	43-52	-	-	-	-	20	[59]
	Comte (ng/g)	France	55-84	-	-	-	-	52-60	[59]
	Emmental (ng/g)	Swiss	81-86	-	-	-	-	222-314	[59]
	Gruyere (ng/g)	Swiss	81-96	-	-	-	-	ND	[59]
	Jarlsberg (ng/g)	Norway	84	-	-	-	-	652	[59]
	Raclette (ng/g)	Swiss	50	-	-	-	-	47	[59]
	Cheese	Cheddar cheese (µg/100 g)	USA	10.2	-	-	-	-	-
Swiss cheese (µg/100 g)		USA	7.8	-	-	-	-	-	[50]
Mozzarella cheese (µg/100 g)			-	-	-	-	-	-	[50]
Processed cheese ((µg/100 g)		Japan	5	-	-	0.3	-	-	[12]

Table 8. Menaquinones contents of various cheeses

Positive correlation was found in propionate concentration and viable propionibacterial cell count which is contributed toward the production of MK-9 in cheeses [59]. Earlier, different reports showed that components of menaquinones are varied among the types of cheeses. In this context, menaquinones concentration is better in Edam-type cheeses than Emmental-type cheeses-specific bacterial activity [53]. Starter culture has prime importance during the preparation of cheese with higher amount of menaquinones. Commonly, Swiss-type cheeses are prepared with *propionibacteria* and lactic acid bacteria, while Edam-type cheese are produced with the action of lactic acid bacteria only such as *Lc. lactis* ssp. *Cremoris* and *Lactococcus lactis* ssp. *Lactis* which are mainly responsible for the synthesis of MK-8 and MK-9 [60]. Moreover, *Propionibacterium freudenreichii* isolated from Swiss-type cheese has a potential equivalent to the *Bacillus subtilis* to produce menaquinone-9 in milk whey [61].

Long-chain menaquinones such as MK-6 to MK10 was not present in Comte hard cheese produced in France. Likewise, these menaquinones were not present in the Emmental hard cheese while some amount of MK-10 and MK-11 was detected. Interestingly, mozzarella cheese did not have any type of menaquinones because during its production process no fermentation is involved [58]. Accordingly, further research is required to evaluate the stability of menaquinones in cheese that are stored for a long time [59].

8.6. Egg

Similar to the other animal-based products, hen egg also contained considerable amount of MK-4, whereas MK-7 was not detected or not quantified by the researchers. Egg yolk contained greater concentration of the MK-4 (31.4-64 µg/100 g) than that of egg white (0.9-7 µg/100 g). Additionally, cooking also increased the MK-4 content in the egg might be due to loss of moisture content compared with whole fresh egg (Table 9). In this context, whole fried egg

contained 9.0 µg/100 g and hard cooked whole egg contained 7.0 µg/100 g compared to fresh whole egg 5.6 µg/100 g [13, 21, 50].

Food	Type	Country	MK-4	MK-6	MK-7	Reference
Egg	yolk	Netherlands	31.4	0.7	–	[13]
Egg	albumen	Netherlands	0.9	–	–	[13]
Egg	Whole and raw	Japan	7	–	ND	[12]
Egg	Raw yolk	Japan	64	–	ND	[12]
Egg	White fresh raw	USA	0.4	–	–	[50]
Egg	Yolk fresh raw	USA	15.5	–	–	[50]
Egg	Whole and fresh	USA	5.6	–	–	[50]
Egg	Whole and fried	USA	9.0	–	–	[50]
Egg	Whole and hard cooked	USA	7.0	–	–	[50]

Other menaquinones were not detected or quantified.

MK-7 was not detected in fresh whole egg and raw egg yolk.

Table 9. Menaquinones contents of hen egg (µg/100 g).

8.7. Fast foods

Elder et al. reported that various fast food products including hamburgers, sandwiches, burrito, taco, pepperoni, and shakes contained MK-4 [50]. Regular hamburger contained lower amount of MK-4 which was subsequently increased by the addition of cheese or sauces or both in the hamburgers. Likewise, Chicken sandwich contained relatively higher amount of MK-4 as 2.7–10.6 µg/100 g than that of hamburger due to higher amount of chicken that possibly contained higher amount of MK-4. Burrito prepared with beans, beef, or chicken contained MK-4 ranged from 0.6 to 2.7 µg/100 g. Pepperoni contained almost similar amount MK-4 as present in the burrito. Shakes available in USA market including chocolate and vanilla also have some amount of MK-4 (**Table 10**).

Food	Type	Country	MK-4	Reference
Hamburger	Regular, with cheese, sauces, and both	USA	1.4–2.9	[50]
Sandwich	Prepared the various meat-based products chicken sandwich	USA	2.7–10.6	[50]
Sandwich	Fish sandwich	USA	0.3	[50]
Burrito	Burrito with bean, beef, and chicken	USA	0.6–2.7	[50]
Taco	Taco regular, with beef, chicken, or cheese	USA	1.0–4.5	[50]
Pepperoni	Pepperoni (regular, thin, and thick crust) or meat and vegetables	USA	1.9–2.1	[50]
Shakes	Shakes, chocolate, and vanilla	USA	3.4	[50]

Other menaquinones were not quantified in this study.

Table 10. Menaquinones contents of various fast foods (µg/100 g).

9. Bioavailability

The Food and Nutrition Board established the RDA level for vitamin K as 65 and 80 μg for adult women and men, respectively [62]. The adequate intake of vitamin K from food sources is relative higher about 120 $\mu\text{g}/\text{day}$ and 90 $\mu\text{g}/\text{day}$ for men and women, respectively [63, 64]. Neonates need approximately 2–2.5 $\mu\text{g}/\text{day}$ of vitamin K that progressively increases up to 30–55 $\mu\text{g}/\text{day}$ in children.

Both active forms of vitamin K, *that is*, phylloquinone and menaquinones, have similar absorption and assimilation pattern. However, relative few studies were carried out to estimate the absorption efficiency, including transportation, distribution, and cellular uptake of various menaquinones [10, 65]. Absorbed vitamin K is transported mainly through lymphatic system in chylomicrons to the hepatic tissue which is main storage organ for long-chain dietary forms of vitamin K [2].

Gut absorption of all dietary forms of vitamin K appears to occur through the common pathway like most of dietary lipids. Bile acids and pancreatic enzymes accelerate the solubility, emulsification, and assimilation of vitamin K into mixed micelles in digestive system. In enterocytes, vitamin K is attached with chylomicrons and enters in lymphatic circulation system. The bioavailability of vitamin K dietary forms is positively associated with dietary lipids and integrity of food matrix [66, 67].

It was reported that isoprenoid side chain length was changed during cellular uptake, transportation, and storage of long-chain menaquinones. Variations were observed in absorption and transportation of vitamin K dietary forms such as phylloquinone, menaquinones (MK-4 and MK-9) after equivalent amount administration of respective form. Postprandial plasma concentration and absorption of MKs are relatively less than phylloquinone, and its uptakes are more in tissues.

In contract to phylloquinone, which is principally fund in triglyceride-rich lipoproteins during postprandial as well as fasting condition, menaquinones are reallocated from triglycerides-rich lipoproteins to low-density lipoproteins (LDL) in and after postprandial consumption of vitamin K. Whereas shorter-chain menaquinones, *that is*, MK-4 were redistributed earlier and incorporated with high-density lipoproteins (HDLs) than MK-9. Moreover, only MK-4 is transported via high-density lipoproteins (HDLs) [10, 68].

The clearance of shorter-chain menaquinones was quick, while other was detected after days in plasma. Likewise, MK-7 has same plasma kinetics with higher half-life of 72 h than that of MK-4 and phylloquinone [9, 13, 69–71]. Nevertheless, no information of plasma kinetics is available of other long-chain menaquinones. High concentration of MK-4 was found in non-hepatic tissues of the body after the ingestion of phylloquinone. This might be due to the conversion of phylloquinone to MK-4. The exact phenomenon is still unclear; however, some researchers suggested that it was converted to other menaquinones via prenylation. In this context, deuterium labeled MK-4 was administrated to mice which are converted to MK-4 via integral side chain removal through prenylation. Likewise, ingestion of MK-7 also increases the serum MK-4 level considerably [2, 12, 17, 69, 71]. Contrary, in germ-free rats, MK-4 and

phylloquinone content were increased in extrahepatic tissues through administration of their respective supplements after deficient condition. Moreover, MK-4 concentration was enhanced by phylloquinone administration. They also inferred that the conversion of phylloquinone in to MK-4 in extrahepatic tissues did not require the intestinal bacterial population. This conversion is purely biochemical and remain unclear thus far [72]. Optimal daily vitamin K₂ especially MK-4 intake as well as sufficient serum concentration is required to activate Vitamin K-dependent proteins [27, 73].

10. Demographic study

Progressive administration of MK-7 momentarily increased the level of plasma MK-7; nonetheless, MK-4 supplementation did not enhance the MK-4 concentration in healthy individuals. Therefore, lower dose of MK-7 (45–90 µg/day) is considered to be effective for ameliorating the physiological dysfunctions [69]. Prime circulating form of vitamin K is phylloquinone, whereas menaquinones (MK-9 to MK-13) are abundantly present in liver. Stored vitamin K is rapidly depleted from the body, and almost 60–70% of absorbed vitamin K is finally lost from body through urine (20%) and feces (40–50%) [74, 75].

Various demographic studies were carried out to estimate the level of circulating MK-7 level in normal and unhealthy subjects. In 1990, study was conducted in London to estimate the level of MK-7 in young and elderly normal subjects. They found that serum concentrations ranged from 0.293 to 0.328 ng/ml in healthy individuals. Jamal et al. also assessed the circulating concentration in patients with hip and vertebral fractures subjects and noticed less amount (0.039 and 0.148 ng/ml) compared to normal subjects [76]. Additionally, French young and elderly women had non-significant varied amount of MK-7 (0.221–0.241 ng/ml), whereas hip fracture old women had 0.120 ng/ml of MK-7.

Japanese healthy adults and vertebral fracture older women had 3.820 and 3.290 ng/ml of MK-7, whereas elderly normal women had significantly higher amount of MK-7 (6.260 ng/ml) might be due to higher consumption of natto which are rich with MK-7. However, postmenopausal women contained less amount of serum MK-7 (0.75–1.10 ng/ml) as compared to normal adults (1.214 ng/ml). Moreover, postmenopausal women from Osaka Japan having lower bone mineral density (BMD) showed lesser amount of circulating MK-7 than that of normal BMD women [77].

Likewise, Kaneki et al. reported the serum levels of MK-7 in postmenopausal women from United Kingdom, Hiroshima, and Tokyo [78]. They inferred that Tokyo women had higher amount of MK-7 as 5.26 ng/ml followed by women lived in Hiroshima 1.221 ng/ml and the lowest concentration as 0.371 ng/ml was noticed in United Kingdom population. They also reported that natto intake has positive association with serum concentration of MK-7 in elderly women living in Tokyo, Japan. Serum MK-7 level was maximum (7.91 ng/ml) in women that consumed natto twice or more in a week, whereas MK-7 level was decreased as 2.81 and 0.873 ng/ml in women when the intake of natto was reduced once or less than once in a week, respectively.

Recently, Knapen et al. demonstrated that the intake of MK-7 fortified yoghurt momentarily increased the plasma concentration of MK-7 from 0.28 to 1.66 ng/ml after 14 weeks of intervention in postmenopausal women and healthy men of aged 45–65 years from the Limburg, Netherlands [57]. The regular intake of MK-4 was momentarily lowered (29%) cardiovascular problems in hemodialysis patients of Poland. Although lower serum value of MK-4 in hemodialysis subjects might be due to the less intake of vitamin K₂ or probably slow conversion rate of phylloquinone to menaquinones, MK-4 intake is positively related with the amount of protein and fat consumed [73].

In Japanese young women, average consumption of vitamin K was adequate 230.2/day and nearly 94% individuals consume adequate intake level of vitamin K. Their mean daily intake of MK-4 and MK-7 was 16.9 and 57.4 µg/day, respectively. Both menaquinones, *that is*, MK-4 and MK-7 contribute almost 30% of the daily requirement of vitamin K in the body [12].

11. Conclusion

Vitamin K₂ is present in numerous in varied concentration of long-chain menaquinones and their types. Fermented soybean of all region of the world contained abundant concentration of MK-7 compared with other menaquinones. Fermentation process is facilitated by action of bacteria which attained the status of generally recognized as safe (GRAS) due to their non-pathogenicity. However, animal-based products such as fresh meat of cow, buffalo, other animals, milk, fish, and egg contained greater amount of MK-4 contents. Additionally, fermented milk-based products such as cheese, source milk, butter milk, and menophilic fermented milk contained ample amount of long-chain menaquinones and MK-4 content were limited in these products due to the bacterial action. Some non-fermented cheese did not contain any form of vitamin K₂. During the physiological functioning, MK-4 is converted in to MK-7 which is more effective to curtail the vitamin K deficiency-associated dilemma.

Acknowledgements

Authors are highly thankful to the Higher Education Commission (HEC), Pakistan for providing funds for the research project entitled “Nutritional and biochemical evaluation of vitamin K enriched dietary sources.” We are also thankful to the Dr. Leon J Schurger (*Department of Biochemistry, Maastricht University the Netherlands*) for valuable suggestions during my studies regarding the vitamin K. Moreover, special thanks for the generous support of Pakistan Atomic Energy Commission for providing the facilities.

Author details

Muhammad Yasin^{1*}, Masood Sadiq Butt² and Aurang Zeb¹

*Address all correspondence to: yasinft_uaf@yahoo.com

1 Food Science Division, Nuclear Institute for Food and Agriculture (NIFA) Peshawar, Pakistan Atomic Energy Commission, Pakistan

2 National Institute of Food Science and Technology, University of Agriculture, Faisalabad, Pakistan

References

- [1] Yasin, M., Butt, M.S., Yasmin, A., Bashir, S., 2014. Chemical, antioxidant and sensory profiling of vitamin K-rich dietary sources. *J. Korean Soc. Appl. Biol. Chem.*, 57(2), 153–160.
- [2] Yasin, M., Butt, M.S., Anjum, F.M., Shahid, M., 2013. Nutritional and antioxidant profiling of vitamin K dietary sources. *Pak. J. Nutr.*, 12(11), 996–1002.
- [3] Data, S., Mwanga, J., Shearer, M., Harrington, D., Voong, K., Parlett, T., Wariyar, U., 2012. Prevalence and associated risk factors for vitamin K deficiency in mothers and their newborn babies in an East African setting. *Arch. Dis. Child.*, 97, 90.
- [4] McCann, J.C., Ames, B.N., 2009. Vitamin K, an example of triage theory: is micronutrient inadequacy linked to diseases of aging? *Am. J. Clin. Nutr.*, 90, 889–907.
- [5] Booth, S.L., Al-Rajabi, A., 2008. Determinants of vitamin K status in humans. *Vitam. Horm.*, 78, 1–22.
- [6] Kuwabara, A., Tanaka, K., Tsugawa, N., Nakase, H., Tsuji, H., Shide, K., Kamao, M., Chiba, T., Inagaki, N., Okano, T., Kido, S., 2009. High prevalence of vitamin K and D deficiency and decreased BMD in inflammatory bowel disease. *Osteoporos Int.*, 20, 935–942.
- [7] Booth, S.L., Martini, L., Peterson, J.W., Saltzman, E., Dallal, G.E., Wood, R.J., 2003. Dietary phyloquinone depletion and repletion in older women. *J. Nutr.*, 133, 2565–2569.
- [8] Schurgers, L.J., Teunissen, K.J., Hamulyák K, Knapen, M.H., Vik, H., Vermeer, C., 2007. Vitamin K containing dietary supplements: comparison of synthetic vitamin K₁ and natto-derived menaquinone-7. *Blood*, 109, 3279–3283.

- [9] Novotny, J.A., Kurilich, A.C., Britz, S.J., Baer, D.J., Beverly, A., 2010. Vitamin K absorption and kinetics in human subjects after consumption of ¹³C-labelled phylloquinone from kale. *Br. J. Nutr.*, 104(6), 858–862.
- [10] Shearer, M.J., Newman, P., 2008. Metabolism and cell biology of vitamin K. *Thromb. Haemost.*, 100, 530–547.
- [11] van Summeren, M.J., Braam, L.A., Lilien, M.R., Schurgers, L.J., Kuis, W., Vermeer, C., 2009. The effect of menaquinone-7 (Vitamin K₂) supplementation on osteocalcin carboxylation in healthy prepubertal children. *Br. J. Nutr.*, 102(8), 1171–1178.
- [12] Kamao, M., Suhara, Y., Tsugawa, N., Uwano, M., Yamaguchi, N., Uenishi, K., Ishida, H., Sasaki, S., Okano, T., 2007. Vitamin K content of foods and dietary vitamin k intake in Japanese young women. *J. Nutr. Sci. Vitaminol.*, 53, 464–470.
- [13] Schurgers, L.J., Vermeer, C., 2000. Determination of phylloquinone and menaquinones in food; effect of food matrix on circulating vitamin K concentrations. *Haemostasis*, 30, 298–307.
- [14] Kwak, C.S., Lee, M.S., Park, S.C., 2007. Higher antioxidant properties of chungkookjang, a fermented soybean paste, may be due to increased aglycone and malonylglycoside isoflavone during fermentation. *Nutr. Res.*, 27(11), 719–727.
- [15] Narumi, S., Sasaki, M., Okudera, D., 1998. Postoperative abnormal prothrombinemia in patients with cefoperazone: report of two cases. *Surg. Today*, 28, 227–230.
- [16] Iwamotoa, J., Satob, Y., Takedaa, T., Matsumotoa, H., 2009. High dose vitamin K supplementation reduces fracture incidence in postmenopausal women: a review of the literature. *Nutr. Res.* 29(4), 221–228.
- [17] Okano, T., Shimomura, Y., Yamane, M., Suhara, Y., Kamao, M., Sugiura, M., 2008. Conversion of phylloquinone (Vitamin K₁) into menaquinone-4 (Vitamin K₂) in mice: two possible routes for menaquinone-4 accumulation in cerebra of mice. *J. Biol. Chem.*, 283, 11270–11279.
- [18] Tamang, J.R., Chettri, Rajen, Sharma, Rudra Mani, 2009. Indigenous knowledge of Northeast women on production of ethnic fermented soybean foods. *Indian J. Tradit. Knowl.*, 8(1), 122–126.
- [19] Hu, Y., Ge, C., Yuan, W., Zhu, R., Zhang, W., Due, L., Xue, J., 2010. Characterization of fermented black soybeans Natto inoculated with *Bacillus Natto* during fermentation. *J. Sci. Food Agric.*, 90, 1194–1202.
- [20] Shrestha, A.K., Dahal, N.R., Ndungutse, V., 2010. *Bacillus* fermentation of soybean: a review. *J. Food Sci. Technol. Nepal*, 6, 1–9.
- [21] Juan, M.Y., Chou, C.C., 2010. Enhancement of antioxidant activity, total phenolic and flavonoid content of black soybeans by solid state fermentation with *Bacillus subtilis* BCRC 14715. *Food Microbiol.*, 27, 586–591.

- [22] Dajanta, K., Janpum, P., Leksing, W., 2013. Antioxidant capacities, total phenolics and flavonoids in black and yellow soybeans fermented by *Bacillus subtilis*: a comparative study of Thai fermented soybeans (thua nao). *Int. Food Res. J.* 20(6), 3125–3132.
- [23] Wu, W.J., Ahn, B.Y., 2011. Improved menaquinone (Vitamin K2) production in cheong-gukjang by optimization of the fermentation conditions. *Food Sci. Biotechnol.*, 20(6), 1585–1591.
- [24] Berenjian, A., Mahanama, R., Talbot, A., Regtop, H., Kavanagh, J., Dehghani, F., 2014. Designing of an intensification process for biosynthesis and recovery of menaquinone-7. *Appl. Biochem. Biotechnol.*, 172, 1347–1357.
- [25] Berenjian, A., Raja, M., Peter, V., Andrea, T., Ray, B., Hubert, R., John, K., Fariba, D., 2011. Extraction of menaquinone-7 for supplementation of food. In: *Chemeca 2011: Engineering a Better World: Sydney Hilton Hotel, NSW, Australia, 18–21 September 2011*. Barton, A.C.T.: Engineers Australia, pp. 1616–1625. Availability: <<http://search.informit.com.au>>.
- [26] Booth, S.L., 2012. Vitamin K: food composition and dietary intakes. *Food Nutr. Res.*, 56, 5505.
- [27] Dajanta, K., Apichartsrangkoon, A., Chukeatirote, E., 2011. Antioxidant properties and total phenolics of Thua Nao (a Thai fermented soybean) as affected by *Bacillus*-fermentation. *J. Microbial Biochem. Technol.*, 3, 56–59.
- [28] Obeid A.E.F.E., Alawad, Aisha Mudawi, Ibrahim, Hanan Moawia, 2015. Isolation and characterization of *Bacillus subtilis* with potential production of nattokinase. *Int. J. Adv. Res.*, 3(3), 94–101.
- [29] Sato, T., Yamada, Y., Ohtani, Y., Mitsui, N., Murasawa, H., Araki, S., 2001. Production of menaquinone (Vitamin K2)-7 by *Bacillus subtilis*. *J. Biosci. Bioeng.*, 91(1), 16–20.
- [30] Esteves, E.A., Martino, H.S.D., Olivieira, F.C.E., Bressan, J., Costa, N.M.B., 2010. Chemical composition of soybeans cultivar lacking lipoxygenases. *Food Chem.*, 122, 238–242.
- [31] Ravishankar, B., Dound, Y.A., Mehta, D.S., Ashok, B.K., de Souza, A., Pan, M-H., Ho, C-T., Badmaev, V., Vaidya, A.D.B., 2014. Safety assessment of menaquinone-7 for use in human nutrition. *J. Food Drug Anal.* doi:10.1016/j.jfda.2014.03.001.
- [32] Puri, A., Iqbal, M., Zafar, R., Panda, B.P., 2015. Influence of physical, chemical and inducer treatments on menaquinone-7 biosynthesis by *Bacillus subtilis* MTCC 2756. *Songklanakarinn J. Sci. Technol.*, 37(3), 283–289.
- [33] Tan, M., Liu, H., Li, Z., Sun, X., Zheng, Z., Zhao, G., 2016. Optimization medium composition for vitamin K2 by *Flavobacterium* sp. using response surface methodology and addition of *Arachis hypogaea*. *Braz. Arch. Biol. Technol.*, 59, e16150343.
- [34] Premarani, T., Chhetry, G.K.N., 2011. Nutritional analysis of fermented soybean (Hawaijar). *Assam. Uni. J. Sci. Technol.* 7(1), 96–100.

- [35] Li, H., Feng, F.Q., Shen, L.R., Xie, Y., Li, D., 2007. Nutritional evaluation of different bacterial douche. *Asia Pac. J. Clin. Nutr.*, 16(S1), 215–221.
- [36] Jeff-Agboola, Y.A., Oguntuase, O.S., 2006. Effect of *Bacillus sphaericus* on proximate composition of soybean (*Glycine max*) for the production of soy iru. *Pak. J. Nutr.*, 5(6), 606–607.
- [37] Wei, Q., Chang, S.K.C., 2004. Characteristics of fermented natto products as affected by soybean cultivars. *J. Food Process. Preserv.*, 28(4), 251–273.
- [38] Ang C.Y.W., Liu K., Huang Y., 1999. *Asian Foods Science and Technology*. Technomic Publishing Company Inc., pp. 139–195.
- [39] Kiuchi K., Ohta O., Itoh H., Takahayahi T., Ebine H., 1976. Studies on lipids of Natto. *J. Agric. Food Chem.*, 55, 271–272.
- [40] Ginting E., Arcot J., 2004. High-performance liquid chromatographic determination of naturally occurring folates during tempeh preparation. *J. Agric. Food Chem.*, 52, 7752–7758.
- [41] Sarkar P.K., Morrison E., Tinggi U., Somerset S.M., Craven G.S., 1998. B-group vitamin and mineral contents of soybeans during Kinema production. *J. Sci. Food Agric.*, 78(4), 498–502.
- [42] Nikkuni, S., Karki, T.B., Vilkuh, K.S., Suzukl, T., Shindoh, K., Suzuk, C., Okada, N., 1995. Mineral and amino acid contents of kinema, a fermented soybean food prepared in Nepal. *Food Sci. Technol. Int.*, 1(2), 107–111.
- [43] Iwuoha, C.I., Eke, O.S., 1996. Nigerian indigenous fermented foods, their traditional process operation, inherent problems, improvements and current status. *Food Res. Int.*, 29(5–6), 527–540.
- [44] Moktan, B., Saha, J., Sparker, P.K., 2008. Antioxidant activities of soybean as affected by *Bacillus*-fermentation to cinema. *Food Res. Int.* 41(6), 586–593.
- [45] Sanjukta, S., Rai, A.K., 2016. Production of bioactive peptides during soybean fermentation and their potential health benefits. *Trends Food Sci. Technol.*, 50, 1–10.
- [46] Kim, N.Y., Song, E.J., Kwon, D.Y., Kim, H.P., Heo, M.Y., 2008. Antioxidant and antige-notoxic activities of Korean fermented soybean. *Food Chem. Toxicol.*, 46(3), 1184–1189.
- [47] Lee, N.K., Sowa, H., Hinoi, E., Ferron, M., Ahn, J.D., 2007. Endocrine regulation of energy metabolism by the skeleton. *Cell*, 130, 456–469.
- [48] Rødbotten, R., Gundersen, Thomas, Vermeer, Cees, Kirkhus, Bente, 2014. Vitamin K2 in different bovine muscles and breeds. *Meat Sci.*, 97, 49–53.
- [49] Fujiwara, K., Miyaguchi, Y., Feng, X.H., Toyoda, A., Nakamura, Y., Yamazaki, M., Nakashima, K., Abe, H., 2008. Effect of fermented soybean, “Natto” on the production and qualities of chicken meat. *Asian-Australasian J. Anim. Sci.*, 21(12), 1766–1772.

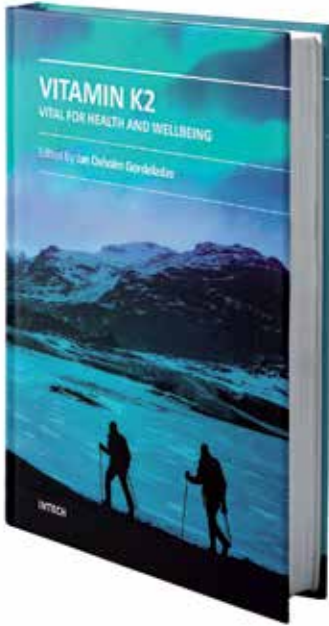
- [50] Elder, S.J., Haytowitz, D.B., Howe, J.R., Peterson, J.W., Booth, S.L., 2006. Vitamin K contents of meat, dairy, and fast food in the US diet. *J. Agric. Food Chem.*, 54(2), 463–467.
- [51] Hirauchi, K., Sakano, T., Notsumoto, S., Nagaoka, T., Morimoto, A., Fujimoto, K., Masuda, S., Suzuki, Y., 1989. Measurement of K vitamins in animal tissues by high-performance liquid chromatography with fluorimetric detection. *J Chromatogr.*, 497, 131–137.
- [52] Matschiner, J.T., Amelotti, J.M., 1968. Characterization of vitamin K from bovine liver. *J. Lipid Res.*, 9(2), 176–179.
- [53] Koivu-Tikkanen, T.J., Ollilainen, V., Piironen, V.I., 2000. Determination of phyloquinone and menaquinones in animal products with fluorescence detection after post-column reduction with metallic zinc. *J. Agric. Food Chem.*, 48, 6325–6331.
- [54] Gentili, A., Caretti, Fulvia, Bellante, Simona, Ventura, Salvatore, Canepari, Silvia, Curini, Roberta, 2013. Comprehensive profiling of carotenoids and fat-soluble vitamins in milk from different animal species by LC-DAD-MS/MS hyphenation. *J. Agric. Food Chem.*, 61, 1628–1639.
- [55] Morishita, T., Tamura, N., Makino, T., Kudo, S., 1999. Production of menaquinones by lactic acid bacteria. *J. Dairy Sci.* 82, 1897–1903.
- [56] Canfield, L.M., Hopkinson, Judy M., Limal, Anne F., Martin, Gall S., Sugimoto, Kyoto, Burr, Jeanne, Clark, Larry, McGee, Daniel L., 1990. Quantitation of vitamin K in human milk. *Lipids* 25, 406–411.
- [57] Knapen, M.H.J., Braam, L.A.J.L.M., Teunissen, K.J., van't Hoofd, C.M., Zwijsen, R.M.L., van den Heuvel, E.G.H.M., Vermeer, C, 2016. Steady-state vitamin K2 (menaquinone-7) plasma concentrations after intake of dairy products and soft gel capsules. *Eur. J. Clin. Nutr.*, 1–6.
- [58] Manoury, E., Jourdon, K., Boyaval, P., Fourcassié, P., 2013. Quantitative measurement of vitamin K2 (menaquinones) in various fermented dairy products using a reliable high-performance liquid chromatography method. *J. Dairy Sci.*, 96(3), 1335–1346.
- [59] Hojo, K., Watanabe, R., Mori, T., Taketomo, N., 2007. Quantitative measurement of tetrahydromenaquinone-9 in Cheese. Fermented by propionibacteria. *J. Dairy Sci.* 90, 4078–4083.
- [60] Morishita, T., Tamura, N., Makino, T., Kudo, S., 1999. Production of menaquinones by lactic acid bacteria. *J. Dairy Sci.*, 82, 1897–903.
- [61] Furuichi, K., Hojo, K., Katakura, Y., Ninomiya, K., Shioya, S., 2006. Aerobic culture of *Propionibacterium freudenreichii* ET-3 can increase production ratio of 1,4-dihydroxy-2-naphthoic acid to menaquinone. *J. Biosci. Bioeng.* 101, 464–470.
- [62] Institute of Medicine, Food and Nutrition Board, 2001. Dietary Reference Intake for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese,

- Molybdenum, Nickel, Silicon, Vanadium and Zinc: A Report of the Panel on Micro-Nutrients. National Academies Press, Washington, DC.
- [63] Food and Nutrition Board, Institute of Medicine, 2001. Vitamin K. In: Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium, and Zinc. National Academy Press, Washington, DC, pp. 162–196.
- [64] Schurgers, L.J., Shearer, M.J., Hamulya, K., Stocklin, E., Vermeer, C., 2004. Effect of vitamin K intake on the stability of oral anticoagulant treatment: dose response relationships in healthy subjects. *Blood*, 104(9), 2682–2689.
- [65] Shearer, M.J., Fu, X., Booth, S.L., 2012. Vitamin K nutrition, metabolism, and requirements: current concepts and future research. *Adv. Nutr.*, 3, 182–195.
- [66] Booth, S.L., Lichtenstein, A.H., Dallal, G.E., 2002. Phylloquinone absorption from phylloquinone-fortified oil is greater than from a vegetable in younger and older men and women. *J. Nutr.*, 132, 2609–2612.
- [67] Jones, K.S., Bluck, L.J., Wang, L.Y., Stephen, A.M., Prynn, C.J., Coward, W.A., 2009. The effect of different meals on the absorption of stable isotopelabelled phylloquinone. *Br. J. Nutr.*, 102, 1195–1202.
- [68] Schurgers, L.J., Vermeer, C., 2002. Differential lipoprotein transport pathways of K-vitamins in healthy subjects. *Biochim. Biophys. Acta*, 1570, 27–32.
- [69] Sato, T., Schurgers, L.J., Uenishi, K., 2012. Comparison of menaquinone-4 and menaquinone-7 bioavailability in healthy women. *Nutr. J.*, 11, 93.
- [70] Schurgers, L.J., Teunissen, K.J., Hamulyak, K., Knapen, M.H., Vik, H., Vermeer, C., 2007. Vitamin K-containing dietary supplements: comparison of synthetic vitamin K1 and natto-derived menaquinone-7. *Blood*, 109, 3279–3283.
- [71] Walther, B., Karl, J., Philip, Booth, Sarah L., Boyaval, Patrick, 2013. Menaquinones, bacteria, and the food supply: the relevance of dairy and fermented food products to vitamin K requirements. *Adv. Nutr.* 4, 463–473.
- [72] Ronden, J.E., Vermeer, C., Thijssen, H.H.W., 1998. Intestinal flora is not involved in the phylloquinone–menaquinone-4 conversion in the rat. *Biochem. Biophys. Acta*, 1379, 69–75.
- [73] Katarzyna, W., Żak-Gołąb, A., Łabuzek, K., Suchy, D., Ficek, R., Pośpiech, K., Olszanecka-Glinianowicz, M., Okopień Bogusław, M., Więcek, A., Chudek, J., 2015. Daily intake and serum concentration of menaquinone-4 (MK-4) in hemodialysis patients with chronic kidney disease, *Clin. Biochem.* doi:10.1016/j.clinbiochem.2015.08.011.
- [74] Shearer, M.J., McBurney, A., Barkhan, P., 1974. Studies on the absorption and metabolism of phylloquinone (vitamin K1) in man. *Vitam. Horm.*, 32, 513–542.

- [75] Usui, Y., Tanimura, H., Nishimura, N., Kobayashi, N., Okanoue, T., Ozawa, K., 1990. Vitamin K concentrations in the plasma and liver of surgical patients. *Am. J. Clin. Nutr.*, 51, 846–852.
- [76] Jamal, S.A., Browner, W.S., Bauer, D.C., et al., 1998. Warfarin use and risk for osteoporosis in elderly women. Study of Osteoporotic Fractures Research Group. *Ann. Intern. Med.*, 128, 829.
- [77] Kanai, T., Takagi, T., Masuhiro, K., et al., 1997. Serum vitamin K level and bone mineral density in post-menopausal women. *Int. J. Gynaecol. Obstet.*, 56, 25.
- [78] Kaneki, M., Hodges, S.J., Hosoi, T., Fujiwara, S., Lyons, A., Crean, S.J., Ishida, N., Nakagawa, M., Takechi, M., Sano, Y., Mizuno, Y., Hoshino, S., Miyao, M., Inoue, S., Horiki, K., Shiraki, M., Ouchi, Y., Orimo, H., 2001. Japanese fermented soybean food as the major determinant of the large geographic difference in circulating levels of vitamin K2: possible implications for hip-fracture risk. *Nutrition*, 17(4), 315–21.
- [79] Shrestha A.K., Noomhorm A., 2001. Composition and functional properties of fermented soybeans flour (Kinema). *J. Food Sci. Technol., Mysore*, 38(5), 467–470.

CHAPTER TITLE

VITAMIN K₂ RICH FOOD PRODUCTS



READ, SHARE
& DOWNLOAD FOR FREE

CHAPTER OFFPRINT FROM

VITAMIN K2 - VITAL FOR HEALTH AND WELLBEING

Edited by Jan Oxholm Gordeladze

ISBN 978-953-51-3020-8

Print ISBN 978-953-51-3019-2

Publisher: InTech

Published March 22, 2017

under CC BY 3.0 license

Book DOI: 10.5772/61430

Chapter DOI: 10.5772/63902

Edited Volume

OPEN ACCESS

CHAPTER PUBLISHED BY

INTECH
open science | open minds

WORLD'S LARGEST
Science, Technology & Medicine Open Access book publisher.

kaqpa
BIOSCIENCE