

Effect of Heat on Egg White Proteins

Zoubida Akkouche*, Lyes Aissat, Khodir Madani

Faculty of Nature and Life Sciences, 3BS Laboratory A. Mira University, Bejaia, Algeria

*Corresponding author, Tel.: +213 0790285388; fax: +213 30 40 62 71

Email: akkzou@yahoo.fr

Abstract

Thermal treatments applied to the egg white cause undesirable modifications of their physicochemical and functional properties. The goal of this study was to acquire some knowledge on the effect of heat on some physico-chemical properties of the albumen while applying different scales (temperature / time). The results show: a sigmoïdale evolution with negative slope of the transmittance, an irreversible loss of solubility. Native-Page of protein's albumen shows the apparition of several bands what denotes diversity of proteins in egg white. The comparison of these profiles dealt to different temperatures /20 min with those unprocessed, shows apparition of new bands and different relative mobility due to the formation of aggregates. Coagulation and/or precipitation of these proteins have been observed around 74°C/20 min.

Keywords: Proteins, egg white, heat denaturation.

1. Introduction

Man has used bird eggs as food since prehistoric times [1, 2]. This reserve of proteins, lipids, vitamins and minerals is remarkable for varieties of nutrients it contains and their high nutritional value, due to a perfect balance of its constituents. Egg is always recognized as a food of high nutritional quality for humans [3, 4, 5, 6, 7]. Beyond their nutritional value, their primary function, namely the formation of a new individual, suggests that they contain many compounds essential to life, and therefore they constitute a major potential source of biological active molecules which interest the pharmaceutical, cosmetic and food [8, 9, 10]. The food industry currently uses largely hen egg (especially egg white) as an ingredient, despite competition from increasingly strong dairy products. The origin of this interest is many functional properties of this latter (foaming and gelling properties in particular) [11]. Egg white protein's are rich in essential amino acids, and possess excellent nutritional value, thus they have long been considered as reference proteins [12, 13, 14]. Nevertheless, the white gotten from breaking eggs is sensitive to contaminations of the environment [15, 16]. To fight against these infections and protect the consumer, the heat treatment of food egg is often required to ensure microbial safety or to obtain desirable organoleptic attributes [17]. The egg white is a hydrated medium, viscous, rich on protein, heat treatment would cause a distortion of its components, the non-enzymatic browning and coagulation of proteins [18, 19]. Polypeptide chains are unfolded and can form a gel or aggregates by the association of these molecules [20]. This aggregation leads to significant changes in the physico-chemical and functional properties of egg proteins [20, 21, 22, 23, 24]. It is therefore very



important that manufacturers in the industry provide an egg product that combines functionality and safety [25]. To limit the damaging effects of heat treatment, some researches propose the addition of additives to increase protein stability to heat. Plant's extracts rich in polyphenols as caffeic acid, helps to increase the thermal stability of milk proteins to the heat [26]. Chemical structure of these substances makes them a highly developed ability to bind to all sorts of molecules, primarily proteins [27].

The purpose of this study was to gain knowledge on the effect of heat on some properties of egg white proteins. This new knowledge will be useful in improving methods of pasteurization and sterilization to reduce the negative impact of heat on technological properties of egg white.

2. Materials and methods

2.1. Preparation of egg white

After washing and cleaning of eggs bought in trade, they are carefully broken and the white and yolk separated, taking the precaution of removing chalazae. The pH of the albumen is measured immediately using a pH meter (Hanna Instruments, Microprocessor pH211 pH Meter); it is equal to 9.45 ± 0.02 .

The volume of egg white is measured using a test-tube and then adjusted with the buffer solution Tris-HCl (20 mM pH 8.2) to obtain a dilution solution of 1:200.

A gentle stirring for 10 minutes is done on the samples, followed by filtration with a strip agase. Identical volumes were transferred into tubes. After heat treatment in a water bath thermostated (Memertt) under continuous agitation at temperatures of $56 \,^{\circ}$ C to $89 \,^{\circ}$ C with a step of $3 \,^{\circ}$ C for 3 min ,10 min and 20 min, the tubes were immediately immersed in an ice bath for 1 minute to stop the denaturation process [16,24].

2.2. Measure of transmittance

It consists in measuring the transmittance of the egg white processed relative to that of untreated; the latter is used to bring back the turbidity at 0 (which corresponds to a transmittance of 100%). This measure is performed at a wavelength of 650 nm [24], immediately after heat treatment and after 24 hours of storage at $4\,^{\circ}$ C.

2.3. Determination of protein solubility

The treated samples were centrifuged at 20000 g for 20 min at $4\,^{\circ}$ C [24]. The proteins of the supernatant were measured by the Bradford method [28]. Solubility S (%) of proteins is expressed relative to an untreated white using the following formula:

S% = Concentration of proteins in supernatant treated

Concentration of proteins in supernatant untreated

The measurements are carried out immediately after the heat and after 24 hours of storage at 4 $^{\circ}$ C.

2.4. Polyacrylamide gel electrophoresis under native conditions (NATIVE-PAGE)

The equipment used is the mini-plate vertical electrophoresis on polyacrylamide gel described by Laemmli (1970) [29]. The protein electrophoresis under non denaturing conditions is done to separate proteins of egg white as a result of an electric field on a separating gel with 7.5% of acrylamide and a staking gel with 5% of acrylamide. The rate of separation depends on the mass and charge of each protein [30].

3. Results and Discussion

3.1. Turbidity

Turbidity is inversely proportional to the transmittance. Figures 1 show a negative slope sigmoidal evolution of protein denaturation of albumen to increasing temperatures ranging from $56\,^{\circ}$ C to $89\,^{\circ}$ C during 3 min, 10 min and 20 min respectively, it has been measured immediately after heat treatment at $650\,\mathrm{nm}$.

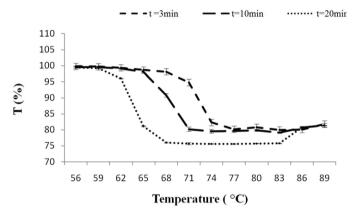


Fig 1. Effect of heat time on egg white turbidity, measures done immediately after heat.

This sigmoid with negative slope includes four distinct parts:

- Stationary Phase 1: The transmittance is steady according to the temperature. This phase could be considered as the phase of denaturation of proteins.
- Downward Phase: Corresponds to a negative evolution of the transmittance according to temperature. It is about the transition phase between the stage of denaturation and the stage of aggregation. The slope of this part informs us on the speed of transition between the stage of denaturation and the one of aggregation.
- Stationary Phase 2: This phase could correspond to the phase of aggregation of proteins.
- Ascending Phase: All curves include a positive phase from 86°C. This reduction of trouble could be associated to the sedimentation of the aggregates formed or to the formation of the S-Ovalbumine of which the ray of Stock is weaker than the one of native protein of a factor of 2, what drives to the formation of small size aggregates.

Statistical analysis reveals that the percentage of transmittance measured immediately and after 24 hours storage at 4 $^{\circ}$ C of heat change over time was significant (P <0.05) and that the disorder is less important at 3 min than 10 min and 20 min respectively. So while extending heat, the trouble appears rapidly (lower temperature) ;for transmittance measured immediately after treatment, the trouble appears from 59 $^{\circ}$ C at 20 min. whereas at 3 min and 10 min, it appears respectively at 68 $^{\circ}$ C and 65 $^{\circ}$ C.

Student test indicates no significant differences between the transmittance measured immediately after heat and after 24 hours storage at 4 ° C treated samples. So the phenomenon of protein denaturation in albumen; is irreversible and interactions occurring between them are covalent (disulfide bridge) and non covalent, (hydrophobic and electrostatic interactions). Statistical analysis reveals that the evolution of transmittance as a function of temperature is significantly different (P <0.05). In fixing the time of heat treatment, the temperature is increasing turbidity increases. This could be explained by protein aggregation most resistant to heat such as ovalbumin, which represents 54% of proteins in egg white [16, 24]. At temperatures understood between 55°C and 65°C, transmittance decrease could be explained by denaturation and aggregation of the most sensitive proteins to heat as the ovotransferrin (13% of proteins in white) and at temperatures more elevated (\geq 74C°), the fall of the transmittance could be due to the denaturation and aggregation of the most heat-resistant proteins such as ovalbumin .

3.2. Solubility

According to figure 2, the heat applied to solutions of egg white lead to irreversible loss of solubility for the three times studied as a function of temperature.

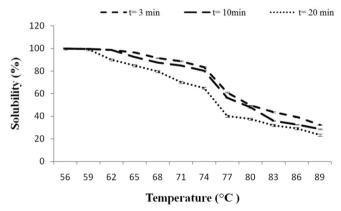


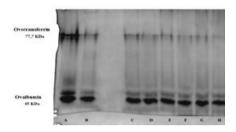
Fig 2. Effect of heat time on egg white solubility, measures done immediately after heat.

After heat during 20 min at 89 °C , only (23.44 ± 0.21) % soluble proteins remained, while after 10 min and 3 min at 89 °C (28.64 ± 0.4) % and (32.43 ± 0.27) %, respectively remained soluble. The strong decrease in solubility at higher temperatures (74-89 °C) is probably due to denaturation and aggregation of heat-resistant proteins such as ovalbumin [31]. While the weak loss in solubility

ity in the range of lower temperatures (56-71° C), could be due to denaturation and aggregation of egg white protein more sensitive to heat, such as ovotransferrin [16, 24, 31].

3.3. Native - PAGE

Figures 3 and 4 show the electrophoretic patterns on polyacrylamide gel (7.5%) in native conditions of egg white proteins after heat during 20 min.



Ovalbumi 48 KDa

Fig 3. Native-Page of albumen heated during 20 min. A: control (Oval, Ovot, Lys) 1mg/ml, B: unheated white, C: 56°C, D: 59°C, E: 62°C, F: 65°C, G: 68°C, H: 71°C.

Fig 4. Native-Page of albumen heated during 20 min. A: 74°C, B: 77°C, C: 80°C, D: 83°C, E: 86°C, F: 89°C.

On the first gel (figure 3), all samples are similar to the control; but ovotransferrin band intensity decreases from 59°C, which could be due to its gelation due to its heat-sensitivity. As the highest temperature here is 71°C, we can conclude that its effect on the structure of the egg white proteins is minimal-or at least it does not cause major changes in its structure; also denaturation protein phase cannot be observed on a Blue Native gel. In the second gel (figure 4),, high temperatures are clearly a major effect because the proteins all migrated to a higher position, suggesting an aggregation-resistant electrophoresis conditions. The comparison between the two gels suggests that under the chosen conditions, coagulation and /or precipitation begins to be observed around 74°C.

4. Conclusion

Egg products are used as ingredients in many food sectors for their techno-unique feature. Stabilization treatments applied to egg white provide good control of hygiene but also induce an alteration of functional properties. Indeed, when egg white is subjected to heat, its globular proteins are prone to changes in structure and conformation. Depending on the extent of the temperature and duration of the treatment, these changes can range from denaturation at the gelation or coagulation. Our study shows effect of heat on some physico-chemical properties of egg white proteins such a sigmoidal evolution of transmittance and an irreversible loss of solubility. Analysis of electrophoretic profiles of native proteins heated at different temperatures, shows appearance of several bands, indicating diversity of proteins in egg white. The comparison of these profiles with those who did not undergo any heat shows appearance of new bands and different relative mobility due to the formation of protein aggregates. Coagulation and / or precipitation of egg white proteins around 74°C/20 min were observed.

5. References

- C.Allemeersch. The ovoproduits in France. Veterinary doctorate thesis. Veterinary National school Of Aflort. 1983; 70:1-151.
- [2] M. Hirose .Globular proteins. In: Progress Biotechnology .ISSN 0921-0423. Published by Elsevier Science B.V. 2003; 23.
- [3] D. Lorient, E. Matringe, F. Vey, O. Bourgeois-Adragna. Functional Properties, sensory and nutritional of the ovoproducts. In: Egg and ovoproducts, Paris, Tec and Doc. Lavoisier. ISBN: 2-85206-903-2. 1994: 368.
- [4] J. Adrian, J. Potus, A. Poiffait. Introduction to the nutritional analysis of the food commodities. Tech ed And Doc, Lavoisier. Paris .1998; 254-260.
- [5] C.Alais, G. Linden. Food biochemistry Abstract. Masson ed. Paris. 1997: 248-250.
- [6] Y.Nys, B.Sauveur. Nutritional Value of eggs. INRA Prod. Anim. 2004; 17 (5): 385-393.
- [7] Ph. Lafon, F. Lafon. The egg and the ovoproducts. Technique of the engineer. 2009; F7010: 1-21.
- [8] S.Rehault, M.Anton, F.Nau, J.Gautron, Y.Nys. The biologic activities of egg. INRA Prod. Anim, 2007; 20 (4): 337-348.
- [9] R. Huopalahti, R. Lopez-Fandino, M. Anton, R. Schade. Bioactive Egg Compounds. ISBN-13:978-3-540-37883-9. Springer Berlin Heidelberg New York. 2007.
- [10] F. Nau, A. Marc, N. Yves. Egg of hen: a mine of molecules whith biological activities. Fifth Day of the Poultry Research, Tours. 2003.
- [11] F. Nau, L. Gestin, J. Protais, A. Awade, JL. Thapon. Comparative study of the functional properties and the composition of thick and liquid fractions of the hen's egg white. ENSAR Agro-Food Department, Reindeer. 1995: 113.
- [12] JC .Cheftel, JL .Cuq, D .Lorient. Food Proteins: biochemistry, functional properties, nutritional value. *Tec ed and Doc. Lavoisier*, Paris. 1985: 306-308.
- [13] Y.Mine .Recent advances in the understanding of egg white protein fonctionality. Trends in Food Science and Technology. 1995; 6: 225-232.
- [14] Y.Mine .Egg Bioscience and Biotechnology. Department of Food Science. University Guelph of. Willy - Interscience. ISBN: 978-0-470-03998-4. 2008.
- [15] B. Sauveur .Structure, composition and nutritional value of hen egg. In: Reproduction of poultries and production of egg. Edition: INRA. ISBN: 2853409619. 1988: 347-436.
- [16] I. Van der Plancken, A. Van Loey, ME. Hendrickx. Combined effect of high pressure and temperature one selected properties of egg white proteins. Innovate Food Science and Ennerging Technology. 2005; 6:11-20.
- [17] JL .Thapon, JC .Bourgeois .Egg and ovoproducts. In: Collection science and agroalimentary techniques. Edition: technique and documentation, Lavoisier. ISBN: 2852069032. 1994: 1-344.
- [18] L.Campbell, V.Raikos, SR. Euston. Modification of functional properties of egg-white proteins. Nahrung/Food. 2003; 47(6): 369-376.

- [19] C. Sanchez, S. Fremont. Consequences of the thermal treatments and formulation on the structure and the allergenic of the food proteins, Direct Science. 2003; 43:13-20.
- [20] M-S .Appavou. Survey of the influence of the temperature and the pressure on the structure and the dynamics of the inhibitor of the bovine pancreatic trypsine. A survey by diffusion of neutrons. Thesis of doctorate. University of Paris XI Orsay. Specialty: Biophysical. 2005; 1-228.
- [21] H.Neurath, HB.Bull.Denaturation and hydration of proteins. From the Division of Agricultural Biochemistry, University Minnesota, St. Paul. 1936: 519-527.
- [22] D. Lorient. Biochemical Modifications of the constituent food. Techniques of engineer. 2009; 3400: 1-20.
- [23] ISL .Navizet. Modelling and analysis of the mechanical properties of the proteins. Thesis of doctorate. Paris University 6 Pierre & Marie Curie. 2004.
- [24] I. Van der Plancken, A. Van Loey, ME. Hendrickx. Effect of heat-treatment one the physico-chemical properties of egg white proteins: A kinetic study. Newspaper of Food Engineering. 2006: 75: 316-326.
- [25] V. Lechevalier. Ciaisaillement, creation of interfaces and thermal treatment. Thesis of physicochemistry Doctorate and quality of the byproducts. School of life-Agro-health doctorate. 2005.
- [26] M-M. Filion. Improvement of the thermal stability of milk by modulation of the potential of oxydoréduction. Faculty of the Sciences of agriculture and food. University Laval, Quebec. 2006.
- [27] E. Hagerman, L. G. Butler. Protein precipitation Method to Quantitative and Determination of tannins. Newspaper of Agriculture. Food Chem. 1978; 26(4): 809-812.
- [28] MM .Bradford. A rapid and sensory method to the quantification of microgram quantities of protein utilizing the principle of protein-dye biding. Anal Bioch. 1976; 72: 248-254.
- [29] UK. Laemmli. Cleavage of structural proteins during the assembly of the head of T4 bacteriophage. Nature. 1970; 227(259): 680-685.
- [30] GM .Walker .The Protein Protocols Handbook (second edition). SD Polyacrylmide Frost Electrophoresis of proteins. University of Hertfordshire, Hatfield, UK. 2002: 1-1103.
- [31] C. Desert, C. Guérin-Dubiard, F. Nau, G. Jan, F. Valley, J. Mallard. Comparison of Defers Electophoretic Separation of Hen Egg White Proteins., School National Superior Agronomic of Rennes, Food Chem. 2001; 49:4553-4561.