

Molecular Microbiology Applied to the Study of Phytopathogenic Fungi

Carlos Garrido, Francisco J. Fernández-Acero, María Carbú,
Victoria E. González-Rodríguez, Eva Liñeiro, and Jesús M. Cantoral
*Microbiology Laboratory, Faculty of Marine and Environmental Sciences
University of Cádiz, Puerto Real
Spain*

1. Introduction

Fungi is an extensive group of eukaryotic microorganisms, generally they are microscopic and usually filamentous. It is estimated that there are between 70,000 and 1.5 millions species of fungi, most of them are being discovering and describing (Agrios, 2005). Most of the known hundred thousand fungal species are strictly saprophytic, living on decomposing dead organic matter. About fifty species cause disease in human, and more than ten thousand species can cause disease in one (obligate parasite) or many kinds of plants (non-obligate parasites) (Fernández-Acero et al., 2007a).

Phytopathogenic fungi are able to infect any tissue at any stage of plant growth. Plant pathogenic fungi show a complex life cycles, including both sexual and asexual reproduction stages (Agrios, 2005). Moreover, complex infection cycles and carbon assimilation is displayed (Garrido et al., 2010). These biological variability give them the possibility to develop its biological role from very climatologically different environments, since dry and desert zones until wet and hot regions in the tropic and equatorial area to the capacity to attack all plant tissues, from leaves to roots (Agrios, 2005).

During the last decades, the development of molecular methods has lead the Scientifics community to accumulate a high quantity of information from different molecular approaches (Fernández-Acero et al., 2011; Garrido et al., 2009b). Advances into Genomics, Transcriptomics, Proteomics, and more recently, Metabolomics are transforming research into fungal plant pathology, providing better and more accurate knowledge about the molecular biology and infection mechanisms showed by these fungi (Garrido et al., 2010).

Since 1992, our research group has been working with two of the most aggressive plant pathogens, which have been established such a model organisms for molecular and phytaphology studies: *Botrytis cinerea* and *Colletotrichum acutatum* (Fernández-Acero et al., 2006b, 2007a; Garrido et al., 2009b, 2010; Perfect et al., 1999). These genera include some of the most destructive plant pathogen species known. They induce worldwide diseases as, between others, the grey mould on grapes and the anthracnose on strawberries, respectively (Coley-Smith et al., 1980; Elad et al., 2004; Sutton, 1992). The losses caused by the phytopathogenic fungi *Botrytis cinerea* and *Colletotrichum acutatum* have been quantified

between 10 and 100 million of Euros per year in Europe (Fernández-Acero et al., 2007a). Losses caused by *B. cinerea* in French vineyards oscillate between 15% and 40%; in Holland, *B. cinerea* generates losses of about 20% of the flower crop; and, in Spain, the losses fluctuate between 20% and 25% of the strawberry crops (Fernández-Acero et al., 2007a). *Colletotrichum* spp. causes up to 80% plant death in nurseries and yield losses of >50%, being a major disease of cultivated strawberry (Denoyes-Rothan et al., 2004; Garrido et al., 2009a).

Our group has carried out an intense research activity of the molecular microbiology of these plant pathogens. These studies involve several molecular approaches in which the gel electrophoresis plays an important role. In this chapter, we will summarize the results obtained, and the molecular methods used for the study and characterization of the phytopathogen fungi *Botrytis cinerea* and *Colletotrichum* spp., all of them strongly related with different types of gel electrophoresis approaches and downstream protocols, including, between others, Pulse Field Gel Electrophoresis, agarose gel electrophoresis of DNA, Restriction Fragment Polymorphism Analyses, Southern-blot, Polyacrylamide Gel Electrophoresis and Two dimensional gel electrophoresis of proteins. These electrophoretic methods will be used to structure the development of chapter, describing the technical bases of each method and showing the approaches carried out and the results obtained.

2. Chromosomal polymorphism and genome organization in *Botrytis cinerea* and *Colletotrichum* spp.

Botrytis cinerea and *Colletotrichum acutatum* are two species of phytopathogenic fungi that show a very high level of phenotypic diversity among isolates. These fungi show complex cycles of life and infection, including both sexual and asexual forms (Garrido et al., 2008; Vallejo et al., 2002). Also high levels of somatic variability appear when the fungi are grown “*in vitro*”, depending on the medium, temperature, light and other factors, which even determine differences in cultural characteristics, production of reproductive structures and pathogenicity between others (Bailey & Jeger, 1992; Carbu, 2006; Garrido et al., 2009b; Rebordinos et al., 1997, 2000; Vallejo et al., 1996, 2001). These fungi do not show a high level of host specificity and they infect many different genera of hosts, adapting their infection strategy and metabolism to the environment conditions and kind of plant colonized. They are notoriously variable genera about which many fundamental questions relating to taxonomy, evolution, origin of variation, host specificity and mechanisms of pathogenesis remain to be answered (Bailey & Jeger, 1992; Elad et al., 2004).

Many research projects are aimed to study the genome organization and chromosomal polymorphism trying to find the origin of phenotypic variability showed by these fungi. In the past decades, several strategies have been tested on lower fungi such as *B. cinerea* and *Colletotrichum* spp., i.e. cytological karyotyping, analysis of progeny from crosses between strains, sexual hybridizations, etc. These assays looked for a relation between molecular and phenotypic variability (Carbu, 2006; Faretra & Antonacci, 1987; Faretra et al., 1988; Vallejo et al., 1996). Cytological studies showed a very high level of difficulty in this group of microorganisms due to small size and/or the difficulty to condense sufficiently the chromosomes to make them visible by microscope. These characteristics made difficult to obtain reliable information about the genome organization of these fungi, and the obtaining of conclusive results about their biological mechanisms of recombination and chromosomal polymorphisms.

The development of Pulse- field gel electrophoresis (PFGE) resolved many problems found with cytogenetic studies in filamentous fungi. This technique has been widely used since the 90s for genomic characterization into fungal plant pathogens. PFGE allows the separation of large DNA molecules (DNAs from 100 bases to over 10 megabases (Mb) may be effectively resolved) which would all co-migrate in conventional agarose gels. This technique has proved to be a very useful tool to study aspects of genome organization in several yeast and fungi. It has led to the discovery that most species exhibit chromosome-length polymorphisms (CLPs), revealing a high level of intraspecific, and even population-level variability (Vallejo et al., 2002).

Technically, PFGE resolves chromosome-sized DNAs by alternating the electric field between spatially distinct pairs of electrodes. The electrophoresis cell consists of an array with 24 horizontal electrodes arranged in a hexagon. Agarose gels are electrophoresed horizontally, submerged under recirculated buffer. The system (CHEF-“Clamped Homogeneous Electric Field” and PACE “Programmable Autonomously Controlled Electrodes”, from BIO-RAD) provides highly uniform, or homogenous, electric fields within the gel, using an array of 24 electrodes, which are held to intermediate potentials to eliminate lane distortion. Thus, lanes are straight. The system maintains uniform field using patented Dynamic Regulation. The electrodes sense changes in local buffer conductivity due to buffer breakdown, change in buffer type, gel thickness, or temperature, and potentials.

The preparation of samples for resolving chromosomal karyotypes by PFGE is not exempt of difficulty due to the biological characteristic of fungal cells. Fungus has to be grown in an optimal culture medium and mycelium harvest after determinate time which depends of the fungal species. This time is very important because is necessary to obtain the highest number of fungal cells in metaphase stage (Carbu, 2006; Garrido et al., 2009b). Chromosomes are condensed and highly coiled in metaphase, which makes them most suitable for visual analysis. After young mycelium is harvested, it is necessary to produce protoplasts using different mixes of lysing enzymes, which digest the fungal cell wall after incubation. Protoplast suspensions are mixed with low melting point agarose, adjusted to final concentration of 1×10^8 protoplast ml^{-1} , and solidified plugs of agarose containing protoplast are digested with proteinase K. The digestion produces pores in the plasma membrane, providing the possibility to extract the chromosomal by PFGE (Garrido et al., 2009b).

Gels are prepared with a special type of agarose. It depends of the DNA molecules sizes because there are different commercial preparations, some of them for DNA molecules higher than 10 Mb, i.e. PFGE™Megabase agarose (Bio-Rad). Plugs are cast in the gel, and this is placed in the center of the hexagon formed by the 24 electrodes. Many parameters of the electrophoresis have to be optimized, since the type and concentration of running buffer, temperature of buffer, voltage and time of pulses, angles of electric fields. Depending of instrument setup, we can resolve the electrophoretic karyotype (EK) only with one experiment, like in the case of *Botrytis cinerea*; or even it could be necessary two different steps/running conditions, due to the high differences in sizes of the chromosomal DNA molecules. After electrophoresis, gels are stained using i.e. ethidium bromide and visualized using a UV light system.

PFGE has been widely used by our group to study the genome organization and Chromosomal Polymorphisms (CPL) in *B. cinerea* and *C. acutatum*. We have determined the number and sizes of chromosomes in both species, and therefore we have estimated the

genome size for these fungi; the high level of CPL displayed by them, represented in the different EK profiles showed by the strains; and PFGE has made possible downstream applications such as Southern-blot analysis using different probes. All the results accumulated during the last years have provided a better understanding about the genome organization and the molecular bases of asexual and sexual reproduction of these fungi. They proved that polymorphism has been observed in both asexual and sexual fungi and most likely results from both mitotic and meiotic processes, especially in the case of *Botrytis cinerea* (Vallejo et al., 2002).

When a study of PFGE has made, it is usual to find chromosomal bands of different intensity and therefore it is important to consider several technical aspects that can have influence in the interpretation of the final results, and the conclusions obtained: i) a double band could be composed of two compounds of a couple of homologous chromosomes or of two heterologous chromosomes of similar size, and ii) two homologous chromosomes can differ in size and appear like two heterologous ones. Due to this fact, depending on the aims of the study, sometimes further hybridization studies are necessary in order to determine the linkage groups of each of the bands (Carbu, 2006; Vallejo et al., 1996).

Botrytis cinerea strains studied by our group were isolated from different hosts and geographical origins. We found different EK profiles between isolates, which did not follow any correlation with the host, year of isolation, or phenotypical characteristics. We have found that the number of chromosomal bands varied between 5 and 12, and they ranged between 1.80 and 3.8 Mb. These results made possible to estimate the minimal genome size of *B. cinerea* genome, found between 14.5 and 22.7 Mbp (Carbu, 2006; Vallejo et al., 1996, 2002) (Fig. 1a).

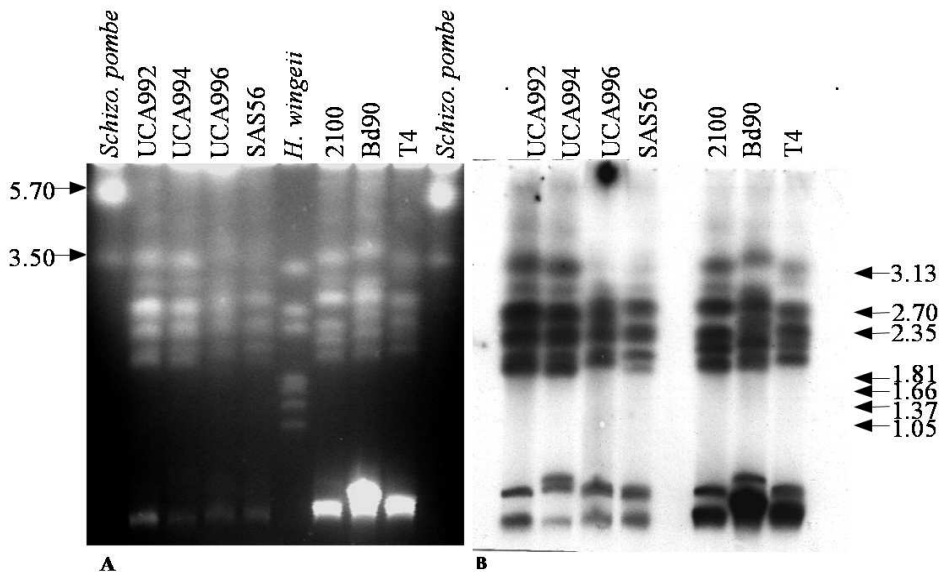


Fig. 1. A.- PFGE chromosomal separation of selected *B. cinerea* isolates. The molecular sizes were estimated using *Schizosaccharomyces pombe* (line 1 and 10), and *Hansenula wingeii* (line 6) chromosomes as reference molecular markers (Bio-Rad). B.- Southern-blot hybridization using a telomeric DNA probe to hybridise the PFGE separated chromosomal bands.

The *B. cinerea* strains showed a high level of CLPs, revealing the facility to support chromosomal rearrangements in this species, and could be the basis of the high degree of adaptability to the environmental conditions. Our group has also studied crosses between strains with different EK profiles. This study had as main aim to analyze the chromosomal rearrangements and chromosomal segregation in the crossed strains, in order to clarify the controversy appeared about the possibility that a high level of CLPs between strains, could inhibit meiosis (Zeigler, 1998), and therefore to be one possible reason to explain the low level of sexual reproduction that take place in *B. cinerea* under natural conditions (Carbu, 2006; Giraud et al., 1997).

The crosses between strains produced fertile strains (more than 100 ascospores studied) and our results demonstrated that chromosomal rearrangements did not affect the capacity to reproduce sexually in *B. cinerea*. It was observed that only several isolates recovered the parental EKs. New chromosomes sizes were identified and some bands were lost from the parental to descendants EKs. All these results, along with a segregation analyses carried out in the descendants, represented strong evidence that some strains might not be haploid, and that aneuploidy and differences in ploidy levels are present in this species (Vallejo et al., 2002). Our group has also studied how during a long period of time, reproducing the fungus "in vitro", there were not detected changes in the EK of a given strain. All results together, proved that mitotic growth does not provide EK variability in this fungus, being the chromosomal rearrangements generated after meiotic recombination the causal agent of EK variability in *B. cinerea* (Carbu, 2006).

In the case of the species *C. acutatum*, there were not data published about the EKs and CLPs among isolates until the last 2009 (Garrido et al., 2009b). PFGE had been used with other species of this genus, like *C. gloeosporioides* (Masel et al., 1993) and *C. lindemouthianum* (O'Sullivan et al., 1998). *Colletotrichum* spp. displayed an estimated genome sizes higher than *B. cinerea*. Protocols to separate the chromosomes molecules were carried out in two different experimental setups, including variations in the pulse of electric field, percentage of agarose gels and duration of the assays (Masel et al., 1993), i.e. for separation of larger chromosomal molecules in *C. gloeosporioides*, Masel et al. (1993) optimized an PFGE approach running a electrophoresis of seven days long. During this experiment, it was necessary to replace the running buffer each two days to obtain a better resolution in the final image. Similar protocols were used to resolve EK from *C. lindemouthianum* strains (O'Sullivan et al., 1998).

The karyotype of *C. acutatum* was studied by our group in several strains isolated from different geographical origins. They had showed differences in the morphological characteristics in relation to the color and texture of mycelium, ratio of growth in different medium, pathogenicity and level of conidia production (Garrido et al., 2008, 2009b). Protocol to obtain *C. acutatum* protoplasts and the PFGE conditions to separate chromosomes were optimized based on our previous experience with *B. cinerea*. We optimized a PFGE running conditions to separate chromosomes between approximatele 0.1 and 9 Mb after only 72 h. of running. This protocol improved substantially those previously described for *Colletotrichum* spp., which took longer due to the two steps needed to resolve the complete karyotypes. Those longer protocols (Masel et al., 1993) were also tested, and we got the same number and sizes of chromosomal bands, proving the improvement of our optimized 72-hours protocol (Garrido et al., 2009b).

C. acutatum strains showed EK profiles containing between six and nine chromosomal bands with different sizes ranging from 0.1 and 8 Mb. The total minimal genome size estimated for *C. acutatum* ranged between 29 and 36 Mb, which is similar to that previously described for other species of *Colletotrichum* (Masel et al., 1993; O'Sullivan et al., 1998). We observed CLPs between strains studies but further analyses with a high number of isolates could be necessary in order to obtain strong conclusions about the CLPs showed by the species and how this variability could affect the sexual and asexual reproduction of this species in the environment (Garrido et al., 2009b).

PFGE gels from *B. cinerea* and *C. acutatum* were used in downstream applications, like Southern-blot analyses. Gels were transferred to Hybond-N membranes and they hybridised with a telomeric probe confirming that all the bands represented chromosomes. The description of Southern-blot analyses will be described in the next section, but it proved how PFGE, not only provides the possibility to obtain interesting conclusions about the biology and genome organization of these fungi, but also gel electrophoresis techniques are often the starting point for interesting downstream applications that provide more information in the researches of these fungi (Fig. 1b).

In our PFGE studies in *B. cinerea* and *C. acutatum*, it has not been observed a higher EKs variability that showed by phenotypic characteristics among strains (Carbu, 2006; Garrido et al., 2008, 2009a, 2009b; Rebordinos et al., 2000; Vallejo et al., 1996). Phenotypic features were very highly variable between strains with the same EKs. Therefore, we cannot conclude that there is a direct relation between morphological, physiological and pathogenic variability directly related with heterokaryosis, aneuploidy and a variable level of ploidy among strains. New proteomics approaches to *B. cinerea* and *Colletotrichum* spp., which will be described during next pages, is contributing with very interesting data, that in conjunction with genomic information, disclose that phenotypic variation is more related with the synthesis of proteins and their post-transductional modifications, and not only by genotypes encoding them (Fernández-Acero et al., 2011).

3. Phylogenetic relationships between strains of *Colletotrichum* spp. using telomeric fingerprinting

Colletotrichum acutatum is a widely spread species that can be found throughout the world (Whitelaw-Weckert et al., 2007). *C. acutatum* causes anthracnose on a number of economically important crops, including woody and herbaceous crops, ornamentals, fruits, conifers and forage plants (Sreenivasaprasad & Talhinhas, 2005). It was classified as an organism of quarantine significance in Canada from 1991 to 1997, in the UK and the EU since 1993, and it can be found widely spread in the southwest region of USA (EPPO/CABI, 1997; Garrido et al., 2009a; Mertely and Legard, 2004). Investigations of *C. acutatum* were focused in two main aspects of the pathogen: i) cultural and morphological studies (Afanador-Kafuri et al., 2003; Denoyes-Rothan & Baudry, 1995; Garrido et al., 2008); and ii) molecular approaches using molecular techniques including isoenzyme comparisons, Restriction Fragment Length Polymorphism (RFLP) analyses of mitochondrial DNA, Amplified Fragment Length Polymorphism (AFLP), AT rich analyses, Random Amplified Polymorphic DNA (RAPD), and ITS sequences analyses for specific PCR sequencing and identification (Buddie et al., 1999; Freeman et al., 1993; Garrido et al., 2009a, 2009b; Sreenivasaprasad et al., 1996; Talhinhas et al., 2005).

Sreenivasaprasad & Talhinhas (2005) studied *C. acutatum* populations from several hosts and different geographical origins. They established molecular groups based on sequences analyses of the internal transcribed spacers (ITS) of ribosomal DNA polymorphic regions (Sreenivasaprasad & Talhinhas, 2005). ITS regions have been widely used on molecular approaches for studying relationship between microorganisms, and it is also very useful regions for designing molecular approaches to identification and diagnostic protocols, due to the high variability showed by the sequences among species and even strains (Garrido et al., 2009a). The classification carried out by Sreenivasaprasad & Talhinhas (2005), established eight molecular groups for *C. acutatum* species. These molecular groups have been widely used to study the genotypic and phenotypic diversity of this fungus, and to classify isolates from different origin (Whitelaw-Weckert et al., 2007).

During the last years, we carried out a study to classify a worldwide collection of *C. acutatum* strains isolated from thirteen countries (Australia, Canada, France, Germany, Japan, The Netherlands, New Zealand, Norway, Portugal, Spain, Switzerland, USA and UK). For this purpose we used two different molecular approaches in order to study the phylogenetic relationship between strains: i) a sequencing analysis of the internal transcribed spacers (ITS) of the 5.8S ribosomal DNA polymorphic regions; ii) a telomeric fingerprinting study by Southern-blot hybridization, using a telomeric probe after RFLP digestions of genomic DNA (Garrido et al., 2009b).

In total, eighty-one 5.8S-ITS sequences were studied, several strains were sequenced by our group, and other ones used from databases such as reference sequences for allocating our strains in the previously established molecular groups for *C. acutatum*. ITS regions, including 5.8S rDNA, were amplified by conventional PCR using universal primers ITS1 and ITS4 (White et al., 1990). After PCR amplification, products were loaded in a conventional 1% agarose gel for conventional DNA electrophoresis. Products were cut from the gels using a purification kit, DNA was quantified, and subsequently sequenced in both directions (Garrido et al., 2009b).

The phylogenetic study carried out with the sequences allowed us to allocate the strains into *C. acutatum* molecular groups described by Sreenivasaprasad & Talhinhas (2005), but the analysis of bootstrap in the neighbour-joining phylogenetic tree, published by Garrido et al. (2009), showed interesting data about the molecular groups. In base of that analyses, the nine molecular groups previously described (Whitelaw-Weckert et al., 2007), could be grouped in only four groups. Our results proved that A1, A2, A5 A8 and A9 subgroups showed a bootstrap support of 90%, and therefore could be considered such as large group in base to the analyses of the sequences of ITS regions (Garrido et al., 2009b). The same result was observed for subgroups A6 and A4, since these subgroups clustered together with a strong bootstrap support of 91% (Garrido et al. 2009). Our results supported a new classification into four molecular groups instead the nine previously described for this species in base to the ITS sequences (Garrido et al., 2009b).

The phylogenetic analyses showed that the majority of the strains studied grouped in the group A2. This happened because many strains from Spain were included in the analyses. The results proved the high level of similarity between *C. acutatum* strains isolated from Spain. It is also interesting that the A2 group included, principally, isolates from Spain, Portugal, France, UK and USA. *C. acutatum* was first described in the southwest region of

the USA, and then it was observed in France and UK at the beginning of the 80s. It is not clear how the pathogen was introduced into production fields in Europe, but it is thought that the pathogen could have arrived since the American nurseries to the EU (Freeman & Katan, 1997). It should have arrived to France, UK and the Iberian Peninsula fields. The arrival of the pathogen was facilitated by the intense international trade between these countries related with strawberry crop. Therefore the fungus could be introduced by infected plants, contaminated soil associated with strawberry crowns at planting, and quiescent infections on strawberry leaves or fruits (Garrido et al., 2008, 2009b; Leandro et al., 2001, 2003).

In order to complete the phylogenetic classification of our *C. acutatum* strain collection, a different molecular approach was carried out. The results obtained were compared with those from the ITS sequences analyses. We used the profiles obtained after restriction enzymes digestions of genomic DNA, and then hybridized with a telomeric probe by Southern-blot hybridization. Genomic DNA of *C. acutatum* strains were digested to completion with several restriction enzymes in independent experiments (*Bam*HI, *Eco*RI, *Hind*III and *Pst*I). Gel electrophoresis is an intermediate point of the complete protocol. It make possible to separate the DNA fragments obtained after the restriction enzymes digestions. In this case, we used a 1.5% agarose gel, and electrophoresis was carried out in a conventional horizontal tray for DNA electrophoresis (Bio-Rad). After separation of digested fragments, gels were blotting to Hybond-N membrans, being ready for subsequently hybridization (Garrido et al., 2009b).

For Southern-blot hybridization we used a telomeric probe to get hybridization in the telomeric regions. These regions are located at the end of the lineal chromosomes of most eukaryotic organisms, and they are named telomeres. Telomeres are regions of repetitive DNA sequences that protect the end of the chromosome from deterioration or from fusion with neighboring chromosomes. The repeated sequences is dependent of the species. For *C. acutatum* telomeres, we produced our telomeric probe, (TTAGGG)_n, by PCR in the absence of a template using (TTAGGG)₅ and (CCCTAA)₅ primers as it was described by Ijdo et al. (Ijdo et al., 1991). The Hybond-N membranes were allowed to hybridize with the telomeric probe; films images were digitalized and telomeric profiles were analysed using Fingerprinting II software v3.0 (Bio-Rad).

The experimental setup described provided the possibility to obtain two different kinds of results/conclusions from the study: I) Selected restriction enzymes used for RFLP did not produce any cut in the telomeric regions of *C. acutatum* strains. Each band represents a physically distinct telomere extremity. Therefore, taking into consideration the higher number of telomeric extremities and then divided into two, we can determine the number of chromosomes among strains studied. II) The fingerprinting analyses of the telomeric profiles, carried out using Fingerprinting II software, make possible to produce phylogenetic trees based in the similarity of the profiles showed among the strains. Therefore, these results could be compared with those obtained from phylogenetic groups based on ITS sequences.

Among the fifty-two isolates analysed by telomeric fingerprinting, the number of band or telomeres oscillated between twelve and eighteen. Therefore, the minimum number of estimated chromosomes was from six to nine among *C. acutatum* isolates (Garrido et al.,

2009b). In this study the number of strains studied was higher than those studied by FPGE, and although fingerprinting analyses did not make possible to study the chromosomal length polymorphisms among the isolates, the minimum numbers of estimated chromosomes are coincident with those obtained from FPGE analyses, showed in the last section of this chapter.

The telomeric profiles obtained for each isolate of *C. acutatum* were analysed. UPGMA dendrogram showed a representative grouping among the isolates, which was coincident with the grouping in the neighbour-joining phylogenetic tree based on sequences of rDNA ITS regions (Garrido et al., 2009b). All the strains previously classified in the A2 molecular groups, also clustered in a large group with more than 70% of similarity based in this case in the telomeric fingerprinting profiles. These results proved the high level of similarity shows by these isolated, not only based in sequence similarity of one specific region but also in their genotypes and genome organization among *C. acutatum* strains, which suggests a common origin of the strains among the different molecular groups (Garrido et al., 2009b; Talhinhas et al., 2005).

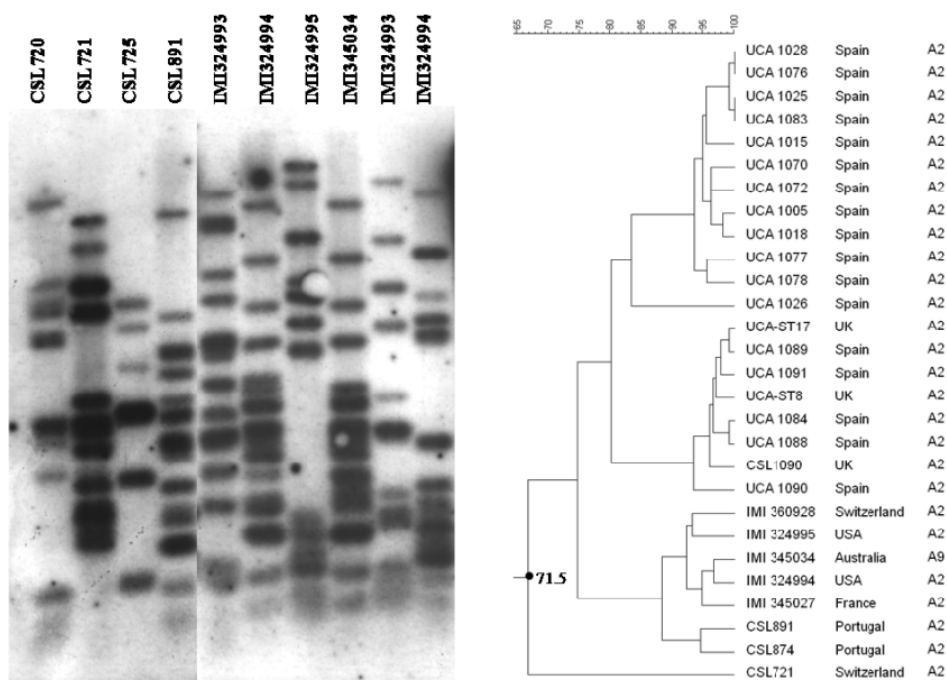


Fig. 2. Left.- Telomeric fingerprinting patterns obtained by telomeric hybridisation of Southern blots from HindIII-DNA digestions. Right.- Combined UPGMA dendograms with the *C. acutatum* isolates belonging to A2 group, based on Dice coefficients generated using a composite data set from individual experiments of each enzyme digestion (*Bam*HI, *Eco*RI, *Hind*III and *Pst*I) hybridised with a telomeric probe.

4. Develop of molecular methods for detection and identification of phytopathogenic fungi – Monitoring of the diseases causing by *Colletotrichum* spp.

Many fungal plant pathogens produce similar symptoms when they develop diseases among different hosts. Currently, the ability to detect, identify and quantify plant pathogens accurately is the cornerstone of plant pathology (Garrido et al., 2011). The reliable identification of the organism(s) responsible for a crop disease is an essential prerequisite to apply the correct disease management strategies and the most appropriate control measures to take. Besides, many pathogens are subjected to special regulation through quarantine programs agreed among producer countries. For all these reasons, pathogen identification is crucial to all aspect of fungal diagnostics and epidemiology in the field of plant pathology, but also in medical science, environmental studies and biological control (Alastair McCartney et al., 2003; Atkins et al., 2003).

Since the 1990's, new methods based on molecular biology have provided new tools for more accurate and reliable detection, identification and quantification of plant pathogens. These methods are based on immunological and DNA/RNA study strategies, including, amongst others: RFLP analyses of mitochondrial DNA (Garrido et al., 2008; Sreenivasaprasad et al., 1992), AFLP, AT-rich analyses (Freeman et al., 2000a, 2000b), RAPD-DNA (Whitelaw-Weckert et al., 2007), genus-specific and species-specific PCR primers (Garrido et al., 2008; Martínez-Culebras et al., 2003; Mills et al., 1992; Sreenivasaprasad et al., 1996), real-time PCR studies (Garrido et al., 2009a), and ELISA assays (Hughes et al., 1997). Diagnosis time can be reduced from a period of weeks, typically experienced with culture plating, to only a few days, thus allowing the appropriate control methods to implemented much sooner and more effectively (Atkins et al., 2003).

Advances in polymerase chain reaction technology have opened alternative approaches to the detection and identification of fungal pathogens. The development of PCR technology relies on three fundamental steps: i) the selection of a specific target region of DNA/RNA to identify the fungus; ii) extraction of total community DNA/RNA from the environmental sample; iii) a method for identifying the presence of the target DNA/RNA region in the sample (Garrido et al., 2011). Our group have optimised a very high sensitive protocol for diagnosis and identification of the fungal genus *Colletotrichum*, and the species *C. acutatum* and *C. gloeosporioides* (Garrido et al., 2009a).

The sensitivity of PCR-based protocols depends mainly on the instrumentation and technique used (i.e. conventional PCR *vs.* real-time PCR), but in a high proportion of cases this sensitivity depends on the quality of the total community DNA/RNA extracted from the environmental samples. Garrido et al. (2009) optimized a DNA extraction protocol that can be used for samples of strawberry plant material directly, or from fungal colonies removed from an agar plate. This method uses sample material physically ground using a grinding machine, in the presence of CTAB lysis buffer. The lysated samples are washed in various chemical products (chloroform, isopropanol, ethanol, etc.) and then the final step involves using Magnesil® beads and GITC lysis buffer (guanidinium thiocyanate buffer) in a Kingfisher robotic processor (Kingfisher ML, Thermo Scientific). The new method was tested with roots, crowns, petioles, leaves and fruits and the extraction methods always showed very high yields of DNA in both quantity and quality. Although, a wide range of

commercial kits are available for extraction of fungal DNA, they can represent a high cost per sample analysed, and they are not always totally reliable in not co-extracting PCR inhibitors, needed a dilution of samples prior to PCR reactions. The optimised protocol did not co-extract PCR-inhibitors from any samples, and therefore, the sensitivity of the detection protocol is improved using this DNA extraction protocols (Garrido et al., 2011).

To date, conventional PCR has been a fundamental part of fungal molecular diagnosis, but it shows several limitations: i.e. gel-based methods, possibility of quantification, sensitivity, etc. The development of real-time PCR has been a valuable response to these limitations (Garrido et al., 2011). This technology improve the sensitivity, accuracy and it is less time-consuming than conventional end-point PCR. For development and optimization of *Colletotrichum* diagnosis protocols, the commonly-used ribosomal RNA genes were used, because of the highly variable sequences of the internal transcribed spacers ITS1 and ITS2, which separate the 18S/5.8S and 5.8S/28S ribosomal RNA genes, respectively (Garrido et al., 2009a). Specific genus and species sets of primers and probes were designed for real-time PCR amplifications using TaqMan® chemistry technology. This system consists of a fluorogenic probe specific to the DNA target, which anneals to the target between the PCR primers; TaqMan® tends to be the most sensitive and simple methods for real-time PCR detection (Garrido et al., 2009a, 2011).

The specificity of all assays was tested using DNA from isolates of six species of *Colletotrichum* and from DNA of another nine fungal species commonly found associated with strawberry material. All the new assays were highly specific for *Colletotrichum* spp., *C. acutatum* and *C. gloeosporioides*, no cross-reactions were observed with either related plant pathogens or healthy strawberry plant material. The sensitivity of the new real-time PCR assays was compared with that of previously published conventional PCR assays; they were confirmed to be 100 times more sensitive than the latter. The *C. acutatum*-specific real-time PCR assay was also compared with an existing ELISA assay for the diagnosis of this pathogen. Real-time PCR permitted the detection of the pathogen in samples that gave negative results for *C. acutatum* using ELISA. The real-time PCR assay detected the equivalent of 7.2 conidia per plant inoculated with a serial dilution of *C. acutatum* spores, demonstrating the high degree of sensitivity of the method (Garrido et al., 2009a).

The new protocols were tested for monitoring the development of anthracnose disease in strawberry in the field in the south of Spain. The real-time PCR results showed a progressive increase of target DNA between January and June. The results showed that an increase in lesion development was accompanied by an increase in the amount and incidence of the pathogen as the season progressed. These results showed that new methods are suitable for diagnosis, identification and monitoring of the disease using field samples of strawberry and also, they permitted the detection of the pathogens from artificially infected symptomless plant material. Therefore, the methods described, based on real-time PCR, proved useful for studying the epidemiological routes of these strawberry pathogens in fields and nurseries (Garrido et al., 2009a, 2011).

5. Proteomics approaches of phytopathogenic fungi

In spite of the advances done by the described techniques above, nowadays proteomics is the most realistic and effective set of tools to unravel complex mixtures of proteins,

describing the current molecular biology age as “post-genomic era”. The term proteome was coined in 1995 by Wilkins et al (Wilkins et al., 1995), later the term proteomics appeared by James et al. (James, 1997). Proteome is defined as the complete set of proteins expressed by an organism, in a particular biological state. Proteomics may be introduced as a set of techniques that allow to study and to describe the proteome. The impact of the proteomic approaches is mainly based in a group of widely used techniques such as liquid chromatography or two dimensional gel electrophoresis, to separate complex protein mixtures, defining the proteome. However, the increasing relevance of these studies has been pushed by the improvements done in mass spectrometry system, allowing the analysis of peptides and proteins and/or by the increase number of proteins entries in the databases, making easier protein analysis and identification.

Main proteome characteristic is that it is a high dynamic system. It is even more complex than genomics, due to while the genome of an organism is more or less constant, the number of obtained proteomes from a specific genome is almost infinite. It depends of the assayed cell, tissue, culture conditions, etc. Each change produces a modification in the observed proteome. An additional factor of complexity is that there are changes that occur in proteome that are not encoded in the genome. These changes are mainly based on two sources, (i) the editing of the mRNA and (ii) post-translational modifications (PTMs) that normally serve to modify or modulate the activity, function or location of a protein in different contexts physiological or metabolic. There are more than 200 different described PTMs (phosphorylation, methylation, acetylation, etc.). They transform each single gene into tens or hundreds of different biological functions. Before proteomics achievements, the differential analysis of the genes, that were expressed in different cell types and tissues in different physiological contexts, was done mainly through analysis of mRNA. However, for wine yeast it has been proved that there is no direct correlation between mRNA transcripts and protein content (Rossignol et al., 2006). It is known that mRNA is not always translated into protein, and the amount of protein produced by a given amount of mRNA depends on the physiological state of the cell. Proteomics confirms the presence of the protein and provides a direct measure of its abundance and diversity.

In terms of methodology, proteomics approaches are classified in two groups, (i) gel free systems, based in the use of different chromatography methods, and (ii) gel based methods, using mainly two dimensional polyacrylamide gel electrophoresis (2DE), that will be the core of our discussion. As a schematic summary, the typical workflow of a proteomic experiment begins with the experimental design. It must be deeply studied, and it will delimit the obtained conclusions, even more when comparison between two strains, cultures or physiological stages between others, are done. From an optimal point of view, only one factor must change between the different assayed conditions (Fernández-Acero et al., 2007a, 2007b). It must contain the use of different biological replicates depending of the used strategy, usually from 3 to 5. The next key step is to obtain a protein extract with enough quality to separate the complex mixture of proteins. Usually, the protein extraction is done in sequential steps (Garrido et al., 2010). First, the biological sample is disrupted using mechanical or chemical techniques. Then, proteins are precipitated and cleaned. Most of the protocol use acetone and trichloroacetic acid. During the next step the proteome is defined and visualized using electrophoretic techniques. 2DE has been widely used for this purpose. Using this technique proteins are separated using two different parameters. During the first dimension, proteins are separated by their isoelectric point using an isoelectrofocusing (IEF)

device. Then, the focused strips are used to load in a polyacrylamide gel, where the proteins are separated by their molecular weight. This system allows the separation of hundreds of proteins from a complex mixture. The gels are visualized with unspecific protein stains (those that stain total proteins, such as Coomassie, Sypro, Silver, etc.), or specific ones (those staining solution prepared to detect specific groups of proteins, mainly post-translational modifications, i.e. Phospho ProQ diamond). The gels are digitalized and analyzed with specific software to reveal the significant spots. Those spots are identified using mass spectrometry. MALDI TOF/TOF is commonly used for 2DE approaches. The huge list of identified proteins obtained is studied to reveal the biological relevance of each identification.

Unfortunately, the number of papers related to fungal proteomics is still poor compared with the application of this technology to other biological sources. As an example, a simple search in WOK website (web of knowledge, <http://www.accesowok.fecyt.es/>) get 809 entries when the terms "proteom*" and "fung*" are used, whereas 51237 entries are displayed when "proteom*" is used alone. In spite of the numerical results obtained may vary depending of the used keywords and web resource, the fact is that there is a lot of work to do to bring fungal proteomic information at the same level that is obtained with other biological sources. This lack is mainly caused by (i) the difficulties to obtain proteins with enough quality to 2DE separations and (ii) the lack of protein sequences listed in the databases. Our research group was pioneer solving these problems and preparing the first proteomic approaches to the phytopathogenic fungi *Botrytis cinerea* (Fernández-Acero et al. 2006).

Fungi posse strong cell walls. This makes difficult the cell breakage using standard protocols. Moreover, fungal proteins extract are characterised by its high concentration of glycosylated proteins that produces dense extracts, dragging a lot of impurities that disturb protein electrofocusing. We optimized a protocol based on a first phosphate buffer solubilisation followed by a typical TCA/Acetone precipitation. Using this protocol we developed the first proteomic map of *Botrytis cinerea* (Fernández-Acero et al., 2006b). Using this optimized approach we prepared a differential proteomics approach based on 2DE, comparing the proteomes of two *B. cinerea* strains differing in virulence (Fernández-Acero et al., 2007b). In spite of this protocol has been widely cited and used (Cobos et al., 2010; Fernández-Acero et al., 2010, 2011; Michielse et al., 2011; Moreira et al., 2011; Sharma et al., 2010; Yang et al., 2011), our recent data suggest that the phosphate buffer solubilisation produces an artificial enrichment of soluble proteins in our assayed extracts. For this reason, we improved our method using a phenol based protocol preparing a *Botrytis cinerea* map during cellulose degradation (Fernández-Acero et al., 2010). Based on this protocol, adding a previous step of precipitation with DOC, we developed the analysis of the main fungal subproteome, the secretome. We identified 76 secreted proteins from cultures where the virulence was induced with different plant-based elicitors (Fernández-Acero et al., 2010). New projects to unravel proteome content of *Botrytis cinerea* and *Colletotrichum acutatum* are running.

All the proteomic approaches developed on *B. cinerea* has been facilitated by the availability of fungal genome sequence (Amselem et al., 2011) (<http://urgi.versailles.inra.fr/Species/Botrytis>, and http://www.broadinstitute.org/annotation/genome/botrytis_cinerea/Home.html). Summarizing all our identified spots, we do not get the 3% of the

predicted genome. The method to capture new fungal proteins, its identification by mass spectrometry and to determine their biological relevance needs to be determined yet. By using our previous experience with *B. cinerea*, we are developing proteomic approaches to *C. acutatum*. Its conidial germination, mycelia dataset and secretome are characterized by 2DE. The key challenge is in our opinion, the use of the collected information to develop new methodologies to fight against plants pathogens. As a future prospect, the development of new environmental friendly proteomics-based fungicides has been discussed (Fernández-Acero et al., 2011).

6. Acknowledgements

This research has been financed by the Spanish Government DGICYT - AGL2009-13359-CO2/AGR, by the Andalusian Government: Junta de Andalucía, PO7-FQM-002689, <http://www.juntadeandalucia.es/innovacioncienciayempresa>; Programa Operativo 2007-2013 (FEDER-FSE) (18INSV2407, 18INSV2610), and by the CeiA3 International Campus of excellence in Agrifood (18INACO177.002AA, <http://www.uco.es/cei-A3/>). Victoria E. González-Rodríguez was financed by the grant FPU of the Ministerio de Educación, Government of Spain, Ref. AP2009-1309; Eva Liñeiro was financed by the grant of the University of Cádiz Ref. 2010-152.

7. References

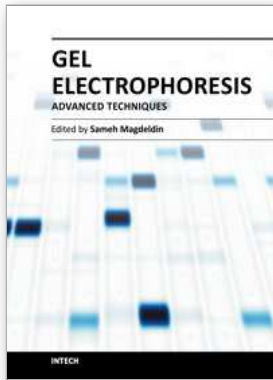
- Afanador-Kafuri, L., Minz, D., Maymon, M., Freeman, S. (2003). Characterization of *Colletotrichum* isolates from Tamarillo, Passiflora, and Mango in Colombia and identification of a unique species from the genus. *Phytopathology*, Vol. 93, pp. 579-587, 0031-949X
- Agrios, G.N. (2005). *Plant Pathology*. (5th Ed). Elsevier Academic Press, ISBN: 13: 978-0120445653, San Diego, CA
- Alastair McCartney, H., Foster, S.J., Fraaije, B.A., Ward, E. (2003). Molecular diagnostics for fungal plant pathogens. *Pest Management Science*, Vol. 59, pp. 129-142, 1526-498X
- Amselem, J., Cuomo, C.A., van Kan, J.A.L., Viaud, M., Benito, E.P., Couloux, A., Coutinho, P.M., de Vries, R.P., Dyer, P.S., Fillinger, S., Fournier, E., Gout, L., Hahn, M., Kohn, L., Lapalu, N., Plummer, K.M., Pradier, J.-., Quévillon, E., Sharon, A., Simon, A., Have, A., Tudzynski, B., Tudzynski, P., Wincker, P., Andrew, M., Anthouard, V., Beever, R.E., Beffa, R., Benoit, I., Bouzid, O., Brault, B., Chen, Z., Choquer, M., Collémare, J., Cotton, P., Danchin, E.G., Da Silva, C., Gautier, A., Giraud, C., Giraud, T., Gonzalez, C., Grossetete, S., Güldener, U., Henrissat, B., Howlett, B.J., Kodira, C., Kretschmer, M., Lappartient, A., Leroy, M., Levis, C., Mauceli, E., Neuvéglise, C., Oeser, B., Pearson, M., Poulain, J., Poussereau, N., Quesneville, H., Rasclé, C., Schumacher, J., Ségurens, B., Sexton, A., Silva, E., Sirven, C., Soanes, D.M., Talbot, N.J., Templeton, M., Yandava, C., Yarden, O., Zeng, Q., Rollins, J.A., Lebrun, M.-., Dickman, M. (2011). Genomic analysis of the necrotrophic fungal pathogens *Sclerotinia sclerotiorum* and *Botrytis cinerea*. *PLoS Genetics*, Vol. 7, No. 8, e1002230. doi:10.1371/journal.pgen.1002230, 1553-7390
- Atkins, S.D., Clark, I.M., Sosnowska, D., Hirsch, P.R., Kerry, B.R. (2003). Detection and quantification of *Plectosphaerella cucumerina*, a potential biological control agent of

- potato cyst nematodes, by using conventional PCR, Real-Time PCR, Selective Media, and Baiting. *Applied and Environmental Microbiology*, Vol. 69, pp. 4788-4793, 0099-2240
- Bailey, J.A., Jeger, M.J. (1992). *Colletotrichum: biology, pathology and control*. CAB international, ISBN: 0-85198-756-7, Wallingford
- Buddie, A.G., Martínez-Culebras, P.V., Bridge, P., García-López, M.D., Querol, A., Cannon, P.F., Monte, E. (1999). Molecular characterization of *Colletotrichum* strains derived from strawberry. *Mycological Research*, Vol. 103, pp. 385-394, 0953-7562
- Carbu, M. (2006). *Estudio de la variabilidad genética y organización cromosómica en el hongo fitopatogeno "Botrytis cinerea"*. Thesis doctoral. University of Cádiz, Cádiz
- Cobos, R., Barreiro, C., Mateos, R.M., Coque, J.R. (2010). Cytoplasmic- and extracellular-proteome analysis of *Diplodia seriata*: A phytopathogenic fungus involved in grapevine decline. *Proteome Sciences*, Vol. 8, doi: 10.1186/1477-5956-8-46, 1477-5956
- Coley-Smith, J.R., Verhoheff, K., Jarvis, W.R. (1980). *The Biology of Botrytis*. Academic Press, ISBN 0-1217 98508X, London, UK
- Denoyes-Rothan, B., Guerin, G., Lerceteau-Kohler, E., Risser, G. (2004). Inheritance of resistance to *Colletotrichum acutatum* in *Fragaria x ananassa*. *Phytopathology*, Vol. 95, pp. 405-412, 0031-949X
- Denoyes-Rothan, B., Baudry, A. (1995). Species identification and pathogenicity study of French *Colletotrichum* strains isolated from strawberry using morphological and cultural characteristics. *Phytopathology*, Vol. 85, pp. 53-57, 0031-949X
- Elad, Y., Williamson, B., Tudzynski, P., Delen, N. (2004). *Botrytis: Biology, Pathology and Control*. Springer, ISBN: 978-1-4020-6586-1, Dordrecht, The Netherland.
- EPPO/CABI (1997). *Quarantine Pests for Europe* (2nd edition). Edited by Smith IM, McNamara DG, Scott PR, Holderness M. CABI International, Wallingford, UK, 1425 pp.
- Faretra, F., Antonacci, E., Pollastro, S. (1988). Sexual behaviour and mating system of *Botryotinia fuckeliana* (*Botrytis cinerea*). *Microbiology*, Vol. 134, No. 9, pp. 2543-2550, 1350-0872
- Faretra, F., Antonacci, E. (1987). Production of apothecia of *Botryotinia fuckeliana* (de Bary) Whetz under controlled environmental conditions. *Phytopathologia Mediterranea*, Vol. 26, pp. 29-35, 0031-9465
- Fernández-Acero, F.J., Colby, T., Harzen, A., Carbú, M., Wieneke, U., Cantoral, J.M., Schmidt, J. (2010). 2-DE proteomic approach to the *Botrytis cinerea* secretome induced with different carbon sources and plant-based elicitors. *Proteomics*, Vol. 10, pp. 2270-2280, 1615-9853
- Fernández-Acero, F.J., Carbú, M., Garrido, C., Vallejo, I., Cantoral, J.M. (2007a). Proteomic Advances in Phytopathogenic Fungi. *Current Proteomics*, Vol. 4, pp. 79-88, 1570-1646
- Fernández-Acero, F.J., Jorge, I., Calvo, E., Vallejo, I., Carbú, M., Camafeita, L.E., Garrido, C., López, J.A., Cantoral, J.M., Jorriñ, J. (2007b). Proteomic analysis of phytopathogenic fungus *Botrytis cinerea* as a potential tool for identifying pathogenicity factors, therapeutic targets and for basic research. *Archives of Microbiology*, Vol. 187, pp. 207-215, 1432-072X
- Fernández-Acero, F.J., Carbú, M., Garrido, C., Collado, I.G., Cantoral, J.M., Vallejo, I. (2006a). Screening Study of Potential Lead Compounds for Natural Product-based Fungicides Against *Phytophthora* Species. *Journal of Phytopathology*, Vol. 154, pp. 616-621, 0931-1785
- Fernández-Acero, F.J., Jorge, I., Calvo, E., Vallejo, I., Carbú, M., Camafeita, E., Lopez, J.A., Cantoral, J.M., Jorriñ, J. (2006b). Two-dimensional electrophoresis protein profile of

- the phytopathogenic fungus *Botrytis cinerea*. *Proteomics*, Vol. 6, No. 1, pp. 88-96, 1615-9853
- Fernández-Acero, F.J., Carbú, M., El-Akhal, M.R., Garrido, C., González-Rodríguez, V.E., Cantoral, J.M. (2011). Development of proteomics-based fungicides: New strategies for environmentally friendly control of fungal plant diseases. *International Journal of Molecular Sciences*, Vol. 12, pp. 795-816, 1422-0067
- Freeman, S., Katan, T. (1997). Identification of *Colletotrichum* species responsible for anthracnose and root necrosis of strawberry in Israel. *Phytopathology*, Vol. 87, pp. 516-521, 0031-949X
- Freeman, S., Minz, D., Jurkevitch, E., Maymon, M., Shabi, E. (2000a). Molecular analyses of *Colletotrichum* species from Almond and other fruit. *Phytopathology*, Vol. 90, pp. 608-614, 0031-949X
- Freeman, S., Shabi, E., Katan, T. (2000b). Characterization of *Colletotrichum acutatum* causing anthracnose of Anemone (*Anemone coronaria* L.). *Phytopathology*, Vol. 66, No. 12, pp. 5267-5272, 0031-949X
- Freeman, S., Pham, M., Rodríguez, R.J. (1993). Molecular genotyping of *Colletotrichum* species based on arbitrarily primed PCR, A+T-rich DNA, and nuclear DNA analyses. *Experimental Mycology*, Vol. 17, pp. 309-322, 0147-5975
- Garrido, C., Carbú, M., Fernández-Acero, F.J., González-Rodríguez, V.E., Cantoral, J.M. (2011). New Insight in the Study of Strawberry Fungal Pathogens. *Genes, Genomes and Genomics*, Vol. 5, No. 1, pp. 24-39, 1749-0383
- Garrido, C., Cantoral, J.M., Carbú, M., González-Rodríguez, V.E., Fernández-Acero, F.J. (2010). New proteomic approaches to plant pathogenic fungi. *Current Proteomics*, Vol. 7, pp. 306-315, 1570-1646
- Garrido, C., Carbú, M., Fernández-Acero, F.J., Boonham, N., Colyer, A., Cantoral, J.M., Budge, G. (2009a). Development of protocols for detection of *Colletotrichum acutatum* and monitoring of strawberry anthracnose using real-time PCR. *Plant Pathology*, Vol. 58, pp. 43-51, 1365-3059
- Garrido, C., Carbú, M., Fernández-Acero, F.J., Vallejo, I., Cantoral, J.M. (2009b). Phylogenetic relationships and genome organization of *Colletotrichum acutatum* causing anthracnose in strawberry. *European Journal of Plant Pathology*, Vol. 125, pp. 397-411, 0929-1873
- Garrido, C., Carbú, M., Fernández-Acero, F.J., Budge, G., Vallejo, I., Colyer, A., Cantoral, J.M. (2008). Isolation and pathogenicity of *Colletotrichum* spp. causing anthracnose of strawberry in south west Spain. *European Journal of Plant Pathology*, Vol. 120, pp. 409-415, 0929-1873
- Giraud, T., Fortini, D., Levis, C., Leroux, P., Brygoo, Y. (1997). RFLP markers show genetic recombination in *Botryotinia fuckeliana* (*Botrytis cinerea*) and transposable elements reveal two sympatric species. *Molecular Biology and Evolution*, Vol. 14, pp. 1177-1185, 1537-1719
- Hughes, K.J.D., Lane, C.R., Cook, R.T.A. (1997). Development of a rapid method for the detection and identification of *Colletotrichum acutatum*, In: *Diagnosis and Identification of Plant Pathogens*, Dehne HW, Adam G, Diekmann M, Frahm J, Mauler-Machnik A & van Halteren P), pp. 113-116. Kluwer Academic, ISBN: 0792347714, Dordrecht (NL).
- Ijdo, J.W., Wells, R.A., Baldini, A., Reeders, S.T. (1991). Improved telomere detection using a telomere repeat probe (TTAGGG)_n generated by PCR. *Nucleic Acids Research*, Vol. 19, pp. 4780, 0305-1048

- James, P. (1997). Protein identification in the post-genome era: the rapid rise of proteomics. *Quarterly Reviews of Biophysics*, Vol. 30, pp. 279-331, 0033-5835
- Kim, Y., Nandakumar, M.P., Marten, M.R. (2007). Proteomics of filamentous fungi. *Trends in Biotechnology*, Vol. 25, pp. 395-400, 0167-7799
- Leandro, L.F.S., Gleason, M.L., Nutter, F.W., Wegulo, S.N., Dixon, P.M. (2003). Strawberry plant extracts stimulate secondary conidiation by *Colletotrichum acutatum* on symptomless leaves. *Phytopathology*, Vol. 93, pp. 1285-1291, 0031-949X
- Leandro, L.F.S., Gleason, M.L., Nutter, F.W., Wegulo, S.N., Dixon, P.M. (2001). Germination and Sporulation of *Colletotrichum acutatum* on symptomless strawberry leaves. *Phytopathology*, Vol. 91, pp. 659-664, 0031-949X
- Martínez-Culebras, P.V., Querol, A., Suarez-Fernández, M.B., García-López, M.D., Barrio, E. (2003). Phylogenetic relationships among *Colletotrichum* pathogens of strawberry and design of PCR primers for their identification. *Journal of Phytopathology*, Vol. 151, pp. 135-143, 0931-1785
- Masel, A., Irwin, J.A.G., Manners, J.M. (1993). DNA addition or deletion is associated with a major karyotype polymorphism in the fungal phytopathogen *Colletotrichum gloeosporioides*. *Molecular Genetics and Genomics*, Vol. 237, pp. 73-80, 1617-4615
- Mertely, J.C., Legard, D.E. (2004). Detection, isolation, and pathogenicity of *Colletotrichum* spp. from strawberry petioles. *Plant Disease*, Vol. 88, pp. 407-412, 0191-2917
- Michielse, C.B., Becker, M., Heller, J., Moraga, J., Collado, I.G., Tudzynski, P. (2011). The *Botrytis cinerea* Reg1 protein, a putative transcriptional regulator, is required for pathogenicity, conidiogenesis and for the production of secondary metabolites. *Molecular Plant-Microbe Interaction*, Vol 24, No. 9, pp. 1074-1085, 0894-0282
- Mills, P.R., Sreenivasaprasad, S., Brown, A.E. (1992). Detection and differentiation of *Colletotrichum gloeosporioides* isolates using PCR. *FEMS Microbiology Letters*, Vol. 98, pp. 137-144, 0378-1097
- Moreira, J., Almeida, R., Tavares, L., Santos, M., Viccini, L., Vasconcelos, I., Oliveira, J., Raposo, N., Dias, S., Franco, O. (2011). Identification of Botryticidal Proteins with Similarity to NBS-LRR Proteins in Rosemary Pepper (*Lippia sidoides* Cham.) Flowers. *The Protein Journal*, Vol. 30, pp. 32-38, 1572-3887
- O'Sullivan, D., Tosi, P., Creusot, F., Cooke, M., Phan, T., Dron, M., Langin, T. (1998). Variation in genome organization of the plant pathogenic fungus *Colletotrichum lindemuthianum*. *Current Genetic*, Vol. 33, pp. 291-298, 0172-8083
- Perfect, S.E., Bleddyn Hughes, H., O'Connell, R.J., Green, J.R. (1999). REVIEW: *Colletotrichum*: A model genus for studies on pathology and fungal-plant interactions. *Fungal Genetics and Biology*, Vol. 27, pp. 186-198, 1087-1845
- Rebordinos, L., Vallejo, I., Santos, M., Collado, I.G., Carbú, M., Cantoral, J.M. (2000). Análisis genético y relación con patogenicidad en *Botrytis cinerea*. *Revista Iberoamericana de Micología*, Vol. 17, pp. 37-42, 1130-1406
- Rebordinos, L., Santos, M., Vallejo, I., Collado, I.G., Cantoral, J.M. (1997). Molecular characterization of the phytopathogenic fungus *Botrytis cinerea*. *Recent Research Developments in Phytochemistry*, Vol. 1, pp. 293-307, 81-308-0052-7
- Rossignol, M., Peltier, J., Mock, H., Matros, A., Maldonado, A.M., Jorrín, J.V. (2006). Plant proteome analysis: A 2004-2006 update. *Proteomics*, Vol. 6, pp. 5529-5548, 1615-9853

- Sharma, N., Rahman, M.H., Kav, N.N.V. (2010). A possible proteome-level explanation for differences in virulence of two isolates of a fungal pathogen *Alternaria brassicae*. *Journal of Plant Biochemistry and Biotechnology*, Vol. 19, pp. 40-49, 0971-7811
- Sreenivasaprasad, S., Talhinhas, P. (2005). Genotypic and phenotypic diversity in *Colletotrichum acutatum*, a cosmopolitan pathogen causing anthracnose on a wide range of hosts. *Molecular Plant Pathology*, Vol. 6, pp. 361-378, 0885-5765
- Sreenivasaprasad, S., Sharada, K., Brown, A.E., Mills, P.R. (1996). PCR-based detection of *Colletotrichum acutatum* on strawberry. *Plant Pathology*, Vol. 45, pp. 650-655, 1365-3059
- Sreenivasaprasad, S., Brown, A.E., Mills, P.R. (1992). DNA sequence variation and interrelationships among *Colletotrichum* species causing strawberry anthracnose. *Physiological and Molecular Plant Pathology*, Vol. 41, pp. 265-281, 0885-5765
- Sutton, B.C. (1992). The genus *Glomerella* and its anamorph *Colletotrichum* , In: *Colletotrichum: Biology, Pathology and Control*, Bailey, J.A. & Jeger, M.J., pp. 1-26, CBA International, ISBN: 0-85198-756-7, Wallingford, UK.
- Talhinhas, P., Sreenivasaprasad, S., Neves-Martins, J., Oliveira, H. (2005). Molecular and phenotypic analyses reveal association of diverse *Colletotrichum acutatum* groups and a low level of *C. gloeosporioides* with olive anthracnose. *Applied and Environmental Microbiology*, Vol. 71, pp. 2987-2998, 0099-2240
- Vallejo, I., Carbú, M., Muñoz, F., Rebordinos, L., Cantoral, J.M. (2002). Inheritance of chromosome-length polymorphisms in the phytopathogenic ascomycete *Botryotinia fuckeliana* (anam. *Botrytis cinerea*). *Mycological Research*, Vol. 106, pp. 1075-1085, 0953-7562
- Vallejo, I., Rebordinos, L., Collado, I.G., Cantoral, J.M. (2001). Differential behaviour of mycelial growth of several *Botrytis cinerea* strains on either Patchoulol- or Globulol-amended media. *Journal of Phytopathology*, Vol. 149, pp. 113-118, 0931-1785
- Vallejo, I., Santos, M., Cantoral, J.M., Collado, I.G., Rebordinos, L. (1996). Chromosomal polymorphism in *Botrytis cinerea* strains. *Hereditas*, Vol. 124, pp. 31-38, 0018-0661
- White, T.J., Bruns, S.L., Taylor, J.W. (1990). Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics, In: *PCR Protocols: A Guide to Methods and Applications*, Innis, M. A., Gelfand, D. H., Sninsky, J. J., and White, T. J., pp. 315, Academic Press, Inc., New York
- Whitelaw-Weckert, M.A., Curtin, S.J., Huang, R., Steel, C.C., Blanchard, C.L., Roffey, P.E. (2007). Phylogenetic relationships and pathogenicity of *Colletotrichum acutatum* isolates from grape in subtropical Australia. *Plant Pathology*, Vol. 56, pp. 448-463, 1365-3059
- Wilkins, M.R., Sanchez, J.C., Gooley, A.A., Appel, R.D., Humphery-Smith, I., Hochstrasser, D.F., Williams, K.L. (1995). Progress with proteome projects: why all proteins expressed by a genome should be identified and how to do it. *Biotechnology & Genetic Engineering Reviews*, Vol. 13, pp. 19-50, 0264-8725
- Yang, J., - Wang, L., - Ji, X., - Feng, Y., - Li, X., - Zou, C., - Xu, J., - Ren, Y., - Mi, Q., - Wu, J., - Liu, S., - Liu, Y., - Huang, X., - Wang, H., - Niu, X., - Li, J., - Liang, L., - Luo, Y., - Ji, K., - Zhou, W., - Yu, Z., - Li, G., - Liu, Y., - Li, L., - Qiao, M., - Feng, L., - Zhang, K. (2011). Genomic and Proteomic Analyses of the Fungus *Arthrotrichy oligospora* Provide Insights into Nematode-Trap Formation. *PLoS Pathogens*, Vol. 7, e1002179, 1553-7366.
- Zeigler, R.S. (1998). Recombination in *Magnaporthe grisea*. *Annual review of phytopathology*, Vol. 36, pp. 249-275, 0066-4286



Gel Electrophoresis - Advanced Techniques

Edited by Dr. Sameh Magdeldin

ISBN 978-953-51-0457-5

Hard cover, 500 pages

Publisher InTech

Published online 04, April, 2012

Published in print edition April, 2012

As a basic concept, gel electrophoresis is a biotechnology technique in which macromolecules such as DNA, RNA or protein are fractionated according to their physical properties such as molecular weight or charge. These molecules are forced through a porous gel matrix under electric field enabling uncounted applications and uses. Delivered between your hands, a second book of this Gel electrophoresis series (Gel Electrophoresis- Advanced Techniques) covers a part, but not all, applications of this versatile technique in both medical and life science fields. We try to keep the contents of the book crisp and comprehensive, and hope that it will receive overwhelming interest and deliver benefits and valuable information to the readers.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Carlos Garrido, Francisco J. Fernández-Acero, María Carbú, Victoria E. González-Rodríguez, Eva Liñeiro and Jesús M. Cantoral (2012). Molecular Microbiology Applied to the Study of Phytopathogenic Fungi, Gel Electrophoresis - Advanced Techniques, Dr. Sameh Magdeldin (Ed.), ISBN: 978-953-51-0457-5, InTech, Available from: <http://www.intechopen.com/books/gel-electrophoresis-advanced-techniques/molecular-microbiology-applied-to-the-study-of-phytopathogenic-fungi>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.