

Electrochemical Sensors Based on Electropolymerized Films

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1. Introduction

Chemical sensors are the devices that provide a certain type of responses related to the quantity of a specific species. All chemical sensors consist of a transducer and a chemically selective layer. The transducers transform the response into a detectable signal on modern instrumentation, and the chemically selective layer isolates the response of the analyte from its immediate environment. According to the properties to be determined, chemical sensors can be classified as electrical, optical, mass or thermal sensors. Among all of them electrochemical sensors obtained more attention because they are sensitive and selective, fast and accurate, portable and inexpensive.

The modification of electrodes surfaces by some special layers has been the major growth area in electrochemical sensors in recent years. Compared to conventional electrodes, greater control of electrode characteristics and reactivity is achieved by surface modification, since the immobilization transfers the physicochemical properties of the modifier to the electrode surface. This process could impart a high degree of selectivity or sensitivity to the electrochemical transducers. Different procedures such as chemical reaction, chemisorption, composite formation or polymer coating have been used to modify electrodes. Several papers have reviewed the application of the modified electrodes in electrochemical sensors areas [1]

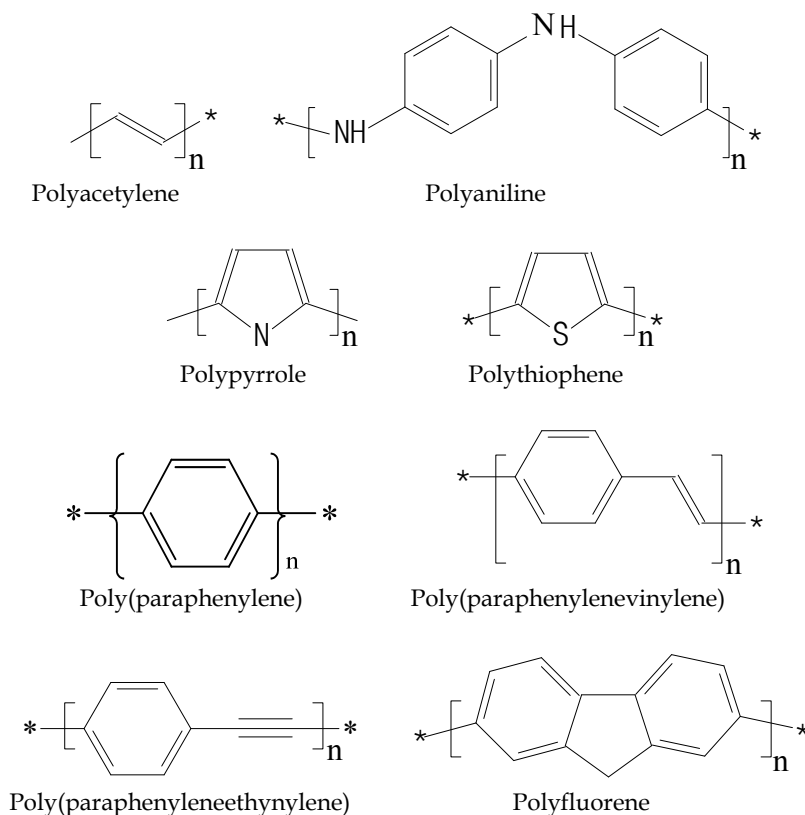
Electrochemical polymerization (ECP) refers to the application of electrochemical methods in the cathode or anode during the polymerization reaction process. The polymerization method provides a new current or potential control factor, so some important advantages were recognized at the beginning of its development such as: short process and low cost. The film thickness and composition can be achieved easily by controlling the electrochemical parameters during the electrochemical process. In addition, electrochemical polymerization can make raw monomer aggregate directly on the substrate film to avoid the use of a large number of volatile organic solvents to achieve the aim of clean production. Because of these excellent features, electropolymerization method holds a promising future for construct simple design, high stable, rapid response and enhanced selectivity sensors. This review will briefly present the development and application of the sensors prepared by electropolymerization method in the recent ten years. The main goal of this contribution is not to collect all papers published recently, but to discuss the development and advantages of the electropolymerized films for analytical purpose. For application, we specifically focused on the most recent and promising applications of those sensors in environmental and clinical monitoring.

2. Classification of films obtained by electrochemical polymerization method

Polymerization is a reaction in which large molecules are created from many small monomers. Normally it is a process which must be controlled carefully under strict conditions. Recently electropolymerization method has been successfully used for the controllable preparation of films because of the advantages we referred in the introduction section. Three types of electrochemical methods are generally employed for the polymerization of different monomers: (1) a constant current (galvanostatic); (b) a constant potential (potentiostatic) and (3) a potential scanning/cycling or sweeping. Different types of polymer films were applied for the construction of sensors. They are classified as conductive and nonconductive films.

2.1 Conducting films prepared by electropolymerization method

Conducting polymers have the polyconjugated structures with electronic properties similar to metals, but retaining the properties of conventional organic polymers. They have gained much attention in sensor areas [2,3] in recent years because of their unique characters [4]. A wide variety of organic molecules have been used as the monomers for the preparation of conducting polymers, such as polycyclic benzenoid, nonbenzenoid hydrocarbons, acetylene, polyaromatic and heterocyclic compounds. Fig. 1 gives some examples of the conductive



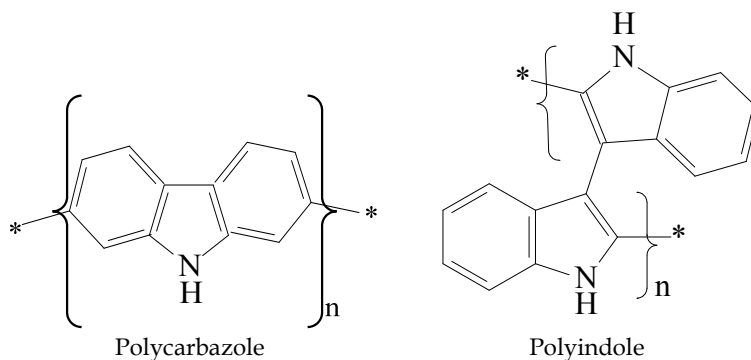


Fig. 1. Main classes of conductive polymers

polymer films. Their unusual electrochemical properties are caused by the conjugated π -electron backbones. A large number of reviews devoted to the fabrication and description of the properties of conducting polymers have been published. Numbers of reviews focus on their use as electrochemical sensors. However there is still considerable interest in the development of new conductive polymers by the electropolymerization method, and new application of the films continually appear. Novel work in the literature from 2000 up to present will be reviewed.

2.2 Non-conducting films prepared by electropolymerization method

Non-conducting films prepared by electropolymerization method are also important. The resulting non-conducting film usually has a small thickness and is self-controlled by the increase in electrical resistance during its growth on the electrode. Because non-conducting polymers are always thin (10-100 nm), substrates and products can diffuse rapidly to and from the film modified electrodes. Therefore, fast response time and high selectivity could be expected for non-conducting polymer based electrochemical sensors. In most cases, phenol and its derivatives are always used for the synthesis of non-conductive films by electrochemical methods [5]. Fig. 2 illustrated the process for the preparing of the phenol related films. Phenolate will be oxidized to generate phenolate radicals which would couple together by ortho- or para- coupling way. Subsequent reactions produce oligomers and, finally, poly(phenylene oxide) films are polymerized on the surface of the electrode. Mahmoudian MR et al prepared poly (pyrrole-co-phenol) (co-PyPh) film by using cyclic voltammetry in the mixture electrolytes of dodecyl benzene sulphonic acid (DBSA) and oxalic acid solution on steel electrodes [6]. It can be used to protect the corrosion of steel. Tahar NB and Savall A [7] have studied the electrochemical oxidation of phenol at different temperatures in basic aqueous solution on a vitreous carbon electrode at different temperatures by cyclic voltammetry and chronoamperometry techniques. Other phenol derivatives have also been prepared by the electropolymerization method. Matsushita et al reported the electropolymerization of coniferyl alcohol in an aqueous system (0.2 M NaOH) and in an organic solvent system [CH_2Cl_2 /methanol (4:1 v/v) in the presence of 0.2 M LiClO_4] [8]. Ciriello R et al. investigated the electrosynthesis mechanism of 2-naphthol (2-NAP) in phosphate buffer at pH 7 on Pt electrodes. The voltammetric behavior suggested the formation of a non-conducting polymer (poly(2-NAP)) through an irreversible electrochemical process complicated by 2-NAP adsorption and fast electrode passivation [9].

Non-conductive polymers obtained from amino acid or their derivatives have also obtained particular interest because they bear specific groups which can interact with some electroactive species through the formation of covalent bonds between either the amino and aldehyde or amino and carboxyl groups. We have used cyclic voltammetric method to form the L-cysteine modified electrode for the detection of sinomenine [10], dopamine [11], terbinafine [12] and adenine [13]. Cysteine was electropolymerized on a glassy carbon electrode in 0.04 M HCl solution in the scan range from -1.20 to 2.60 V at the scan rate of 100 mV/s [13].

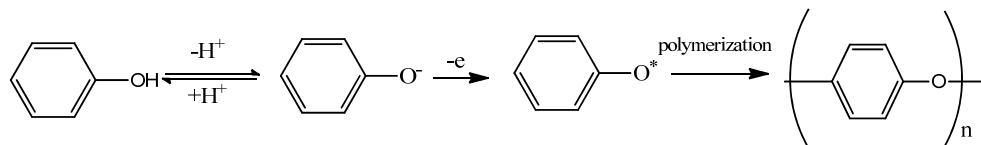


Fig. 2. The process for the preparing of the Phenol film by electrochemical method.

Fig. 3 illustrated the structures of some of the non-conductive films obtained by electropolymerization method. They included the already discussed polyphenol, polymers of phenyldiamines and the overoxidized polypyrrole (PPy). These polymers can be used as a novel support matrix for the immobilization of biomolecules to construct different electrochemical sensors. We would discuss the preparation and application of these films in the following section.

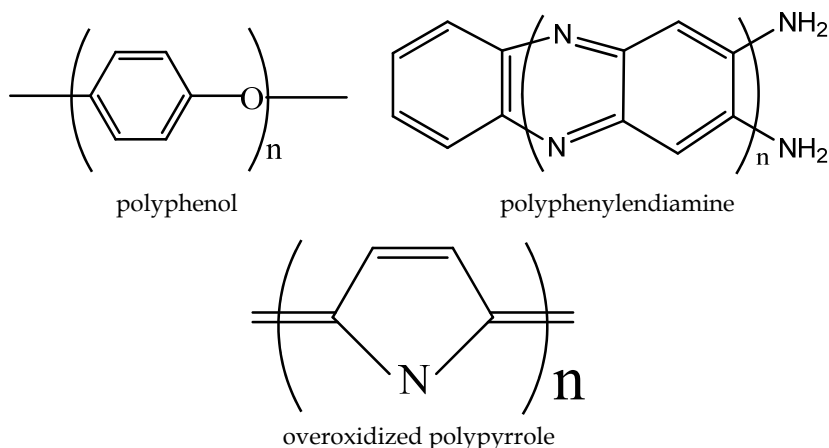


Fig. 3. Structures of some non-conductive polymerfilms

3. Application of the electropolymerized film in analytical areas

Significant advances in electropolymerization areas during the 1990's are certain to facilitate the application of the sensors to different analytical areas. To date, electropolymerized films have been widely used for clinical and environmental detection purpose.

3.1 Electropolymerized films for clinical monitoring

Electrochemical sensors in clinical assay was developed from 1962 by Clark and Lyons who used the glucose oxidase (GOx) enzyme to construct an amperometric electrode for dissolved oxygen detection [14]. From that time, the application of electrochemical sensors to determine the concentration of substances and other parameters of biological interest has represented a rapidly expanding field of instrument. The electrochemical sensors have been widely used in clinical analysis because of their high sensitivity and selectivity, portable field-based size, rapid response time and low-cost. Some of these sensor devices have been routinely used in clinical, industrial, environmental, and agricultural areas. Many works and reviews related on this area have been reported. Lakshmi D et al has reviewed the application of electrochemical sensors for uric acid detection in mixed and clinical samples[15]. Ronkainen et al reviewed the application of electrochemical biosensors from two points: biocatalyst and affinity [16].

Since the original work reported by Diaz et al. [17], the films prepared by electropolymerization method have attracted considerable interest due to their versatility. Polymers have gain considerable interest in the clinical analysis area because of their unique and biochemical properties. In 1992, Davies et al have extensively reviewed the application of the polymer membranes in clinical sensor application [18]. In this work, the authors were concerned with the relationship between the polymer design and the proposed application. They highlighted the permeability, permselectivity and transmembrane potential of the polymer membranes and the role of polymer membranes as matrices for the immobilization of reactive chemical and biological agents. Cosnier reviewed the application of the electropolymerized films on the construction of affinity sensor[19]. He compared the different strategies for the immobilization of biomolecules on electropolymerized films to construct affinity sensors which can be used as clinical sensors. Table 1 [20-52] summarized the numerous of recent applications in clinical areas based on the electropolymerized films. The information on the analytes, the polymer films, and the characters of the sensors has been listed.

Non-conductive polymer from polyphenylenediamines has been used as a matrix for the entrapment of enzymes. Glucose oxidase has been caged into the microtubule structures of polycarbonate membrane by using poly (1, 3- phenylenediamine) to fill the pores. The sensitivity of the sensor increased 60 times [23]. Different techniques for the electropolymerization of 1,2-, 1,3- and 1,4-phenylenediamine, such as cyclic voltammetry and chronoamperometry, were compared by Currulli et al [53]. When heparin was co-immobilized with glucose oxidase during the electropolymerization of a non-conductive poly (1,2- phenylenediamine) film, an implantable glucose biosensor could be constructed [54]. This sensor could prevent the fibrin formation and clotting when the glucose sensor was exposed to blood.

Phenol and its derivative have also been widely used for clinical analysis. The electropolymerization of phenol derivatives is similar to that of phenol. We reported the polymerization of the acid chrome blue K on a glassy carbon electrode by cyclic voltammetric method in 0.05 M pH 7.0 phosphate buffer solution in the potential range from -0.4 V to 1.5 V at the scan range of 100 mV/s by 25 cycles [28]. This film can be used to separate the electrochemical response of dopamine (DA), ascorbic acid (AA) and uric acid (UA). Under the optimum conditions, the calibration curves for DA, AA and UA were obtained in the range of 1.0–200.0, 50.0–1000.0 and 1.0–120.0 μ M, respectively. Both poly

analyte	monomer	linearity or detection limit	Ref
glucose	toluidine blue O	0.1-1.2 mM	20
	preoxidized catecholamines	0.3 μ M	21
	thioaniline functionalized gold nanoparticle	0-200 mM	22
	1,3-phenylenediamine	0.25 μ M - 18 mM	23
	thioaniline-modified glucose oxidase		24
dopamine	N-methylpyrrole	0.1- 10 μ M	25
	aniline/gold nanoparticles	3-115 μ M	26
	1-aminoanthracene	0.56-100 μ M	27
	acid chrome blue K	1.0 - 200.0 μ M	28
prostate specific antigen	poly(1,2-diaminobenzene) as the template for the electropolymerization of polyaniline	1-100 pg/mL	29
uric acid	3-aminophenol		30
	2-aminophenol	0.5 - 0.9 mM	31
DNA	pyrrole		32
		3.7-370 nM	33
		0.16 -3.5 fmol	34
	ferrocene-functionalized pyrrole	0.1-200 nM	35
	gold nanoparticles/p-aminobenzoic acid/carbon nanotubes	1.0 fmol -50 nM	36
	gold nanoparticles/L-lysine	0.1 - 10 fmol	37
	silver nanoparticles/3-(3-pyridyl) acrylic acid/carbon nanotubes	9.0 fmol-9.0 nM	38
leptin	Au-pyrrole propylic acid-pyrrole nanocomposite	10-100000 ng/mL	39
Human IgG	pyrrole-3-carboxylic acid		40
urea	styrene sulphonate-aniline	0-75 mM	41
hemoglobin	pyrrole-gold nanoparticles	60-180 μ g/mL	42
Myeloperoxidase	o-phenylenediamine/multi-wall carbon nanotubes -ionic liquid/gold nanoparticles	0.25-350 ng/mL	43
17- β estradiol	3,4-ethylenedioxythiophene/ gold Nanocomposite		44
nitric oxide	eugenol or o-phenylenediamine		45
	poly(toluidine blue)	0.18-86 μ M	46
	m-phenylenediamine, 2,3-diaminonaphthalene, and 5-amino-1-naphthol polymers	From nM to μ M	47
	meldola blue/chitosan	10 nM-600 μ M	48
cholesterol	2-mercaptobenzimidazole	5-30 μ M	49
nicotinamide	phenothiazine	70 nM	50
adenine dinucleotide	pyronin B	1.0 - 500 μ M	51
interleukin 5	pyrrole-pyrrolepropylic acid-gold nanocomposite	10 fg/mL	52

Table 1. Examples of electropolymerized films for clinical analysis

(3-aminophenol) film [30] and poly (2-aminophenol) [31] film have been used for the selectively detection of uric acid. The poly (2-aminophenol) film was electrochemically prepared on Pt electrodes at a constant potential of 0.3 V from a deoxygenated aqueous solution of monomer dissolved in 0.1 M KCl. This film modified electrode allows the penetration of large amounts of uric acid while blocking the electrochemical activity of ascorbic acid in the potential region examined.

3.2 Electropolymerized film for environmental monitoring

Electrochemical sensors play very important roles in the protection of our environment. They can monitor the pollutant on-site and address some other environmental needs. Several electrochemical devices, such as pH- or oxygen electrodes based on the polymerized films, have been used routinely for years in environmental analysis. The electropolymerized pyrrole [55,56], aniline, thiophene, benzene derivatives and others [57,58] have been used for the preparation of pH chemical sensors. Herlem, G et al prepared the polyglycine-like thin film on platinum electrode by anodic oxidation. The film can be used as a pH sensor in the pH range 2-12 because of the proton affinity towards amino groups of polyglycine [59].

Analyte	monomer	linearity or detection limit	Ref
Hg ²⁺	2-mercaptobenzothiazole	1.0-160.0 nM	62
	2,6-diaminopyridine	10 μM -0.1 M	63
	3-methylthiophene	1.4 μg/ L	64
nitrite	methylene blue-carbon nanotubes-ionic liquid	0.5-67.9 μM	65
	carbon nanotube-anillin	0.2μM -3.1 mM	66
	pyrrole	10μM -1 mM	65
	functionalized thiadiazole	0.05 -16μM	68
ammonia	pyrrole	10- 200 μM	69
Ca ²⁺	melatonin	6.2×10 ⁻⁷ - 1.0×10 ⁻⁴ M	70
Cu ²⁺	2-aminothiazole)-multi-walled carbon nanotubes	0.1 - 20 μM	71
	pyrrole	50 nM- 0.01 M	72
sulfite	copper salen (salen=N,N'-ethylenebis(salicylideneiminato))	4.0 -69μM	73
	aniline	0.006-5 mM	74
methyl-parathion para-nitrophenol	para-phenylenediamine	0.01 to 10 mg/L	75
microbial	4-(2,5-di(thiophen-2-yl)-1H-pyrrol-1-yl)benzenamine (SNS-NH2) polymer	0.1-2.5 mM	76
chloride	3-octylthiophene	10 ⁻⁸ - 10 ⁻¹ M	77
4-nitrophenol	carmin	50 nM - 10μM	78

Table 2. Examples of electropolymerized films for environmental analysis

The scope of the electropolymerized films have also been expanded towards a wide range of organic and inorganic contaminants including pesticides, polychlorinated biphenyls, and heavy metals. A new kind of polymer monomer, bis(terthiophene)-appended uranyl-salophen complex, comprising N,N'-bis[4-(5,2':5',2''-terthiophen-3'-yl)salicylidene]-1,2-ethanediamine-uranyl complexes (TUS), has been modified on a glassy carbon electrode by electrochemical polymerizations. This polymer film has both the functionality of ion-to-electron transducers (solid contact) and Lewis-acidic binding sites to construct a monohydrogen phosphate (MHP) ion-selective electrode (ISE). The detection limit was down to 10^{-5} M and the response time is less than 5s [60]. In some cases, the polymers acted as the matrix for the immobilization of enzyme for environmental analytical application. Chen et al reported the entrapment of glucose oxidase into poly-(L-noradrenalin) films (PNA). They studied the inhibition effects of Hg^{2+} , Cu^{2+} , and Co^{2+} on the activity of glucose oxidase. The electrosynthesized PNA matrix to entrap GOX for an inhibitive assay of Ag^+ shows the lowest competitive affinity to heavy metal ions and gives the highest sensitivity, so it can be used for Ag^+ detection⁶¹. The applications of the electropolymerized films applied for environmental monitoring were listed in Table 2 [62-78].

4. Conclusion and future perspective

There has been an enormous increase of the preparation and application of the polymer films in analytical areas. Some of the advances and fields of the electropolymerized films have been outlined in the review. The electropolymerization methods have important advantages over the conventional techniques for the modification and preparation of microelectrodes, permitting the regulation of the spatial location and selective control of the film properties. Selective immobilization of biomoleculars in array of microelectrodes can be implanted in biological tissues for the simultaneous detection of several compounds. Minimization arrays, fast responding electrochemical sensors and on-line detection are the developing tendency of the electropolymerized films in sensor areas. Some progress will be necessary to achieve the appearance of the commercial electrochemical sensors based on the electropolymerized films. A lot of future research into the development of new polymer films by electrochemical method can be expected.

5. Acknowledgements

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6. References

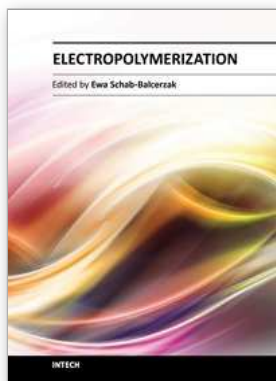
- [1] B.J. Privett, J.H. Shin, M.H. Schoenfish, *Electrochemical Sensors, Analytical Chemistry* 82 (2010) 4723-4741.

- [2] A. Ramanavicius, A. Ramanaviciene, A. Malinauskas, Electrochemical sensors based on conducting polymer- polypyrrole, *Electrochimica Acta* 51 (2006) 6025-6037.
- [3] H. Peng, L.J. Zhang, C. Soeller, J. Travas-Sejdic, *Biomaterials* 30 (2009) 2132-2148.
- [4] C.Y. Wang, J. Guan, Q.S. Qu, G.J. Yang, X.Y. Hu, *Combinatorial Chemistry & High Throughput Screening* 10 (2007) 595-603.
- [5] Z. Ežerskis, Z. Jusys, *Journal of Applied Electrochemistry* 31 (2001) 1117-1124.
- [6] M.R. Mahmoudian, Y. Alias, W.J. Basirun, *Materials Chemistry and Physics* 124 (2010) 1022-1028.
- [7] N.B. Tahar, A. Savall, *Electrochimica Acta* 55 (2009) 465-469.
- [8] Y. Matsushita, T. Sekiguchi, R. Ichino, K. Fukushima, *Journal of Wood Science* 55 (2009) 344-349.
- [9] R. Ciriello, A. Guerrieri, F. Pavese, A. M. Salvi, *Analytical and Bioanalytical Chemistry* 392 (2008) 913-926.
- [10] J. Guan, Z.X. Wang, C.Y. Wang, Q.S. Qu, G.J. Yang, X.Y. Hu, *International Journal of Electrochemical Science* 2 (2007) 572-582.
- [11] C.Y. Wang, Q.X. Liu, X.Q. Shao, X.Y. Hu, *Analytical Letters* 40 (2007) 689-704.
- [12] C.Y. Wang, Y.D. Mao, D.Y. Wang, G.J. Yang, Q.H. Qu, X.Y. Hu, *Bioelectrochemistry* 72 (2008) 107-115.
- [13] Q. Xu, M. Sun, Q.X. Du, X.J. Bian, D. Chen, X.Y. Hu, *Current Pharmaceutical Analysis* 5 (2009) 190-196.
- [14] L.C. Clark, C. Lyons, *Annals of the New York Academy of Sciences* 102 (1962) 29-&.
- [15] D. Lakshmi, M.J. Whitcombe, F. Davis, P.S. Sharma, B.B. Prasad, *Electroanalysis* 23 (2011) 305-320.
- [16] N.J. Ronkainen, H.B. Halsall, W.R. Heineman, *Chemical Society Reviews* 39 (2010) 1747-1763.
- [17] A F. Diaz, K. K. Kanazawa, G. P. Gardini, *Journal of the Chemical Society, Chemical Communications* (1979) 635-636
- [18] M.L. Davies, C.J. Hamilton, S.M. Murphy, B.J. Tighe, *Biomaterials* 13 (1992) 971-978.
- [19] S. Cosnier, *Electroanalysis* 17 (2005) 1701-1715.
- [20] W.J. Wang, F. Wang, Y.L. Yao, S.S. Hu, K.K. Shiu, *Electrochimica Acta* 55 (2010) 7055-7060.
- [21] C. Chen, Y.C. Fu, C.H. Xiang, Q.J. Xie, Q.F. Zhang, Y.H. Su, L.H. Wang, S.Z. Yao, *Biosensors & Bioelectronics* 24 (2009) 2726-2729.
- [22] O. Yehezkeli, R. Tel-Vered, S. Reichlin, I. Willner, *Acs Nano* 5 (2011) 2385-2391.
- [23] M. Lee, Y. Son, J. Park, Y. Lee, *Molecular Crystals and Liquid Crystals* 492 (2008) 155-164.
- [24] O. Yehezkeli, Y.M. Yan, I. Baravik, R. Tel-Vered, I. Willner, *Chemistry-a European Journal* 15 (2009) 2674-2679.
- [25] N.F. Atta, M.F. El-Kady, A. Galal, *Analytical Biochemistry* 400 (2010) 78-88.
- [26] A.J. Wang, J.J. Feng, Y.F. Li, J.L. Xi, W.J. Dong, *Microchimica Acta* 171 (2010) 431-436.

- [27] E.D. Troiani, R.C. Faria, *Electroanalysis* 22 (2010) 2284-2289.
- [28] R. Zhang, G.-D. Jin, D. Chen, X.-Y. Hu, *Sensors and Actuators B* 138 (2009) 174-181.
- [29] A.C. Barton, F. Davis, S.P.J. Higson, *Analytical Chemistry* 80 (2008) 6198-6205.
- [30] S. Kursun, A. Pasahan, B.Z. Ekinci, E. Ekinci, *International Journal of Polymeric Materials* 60 (2011) 365-373.
- [31] S. Kursun, B.Z. Ekinci, A. Pasahan, E. Ekinci, *Journal of Applied Polymer Science* 120 (2011) 406-410.
- [32] J.H. Jin, E.C. Alocilja, D.L. Grooms, *Journal of Porous Materials* 17 (2010) 169-176.
- [33] H. Peng, C. Soeller, M.B. Cannell, G.A. Bowmaker, R.P. Cooney, J. Travas-Sejdic, *Biosensors & Bioelectronics* 21 (2006) 1727-1736.
- [34] C.D. Riccardi, H. Yamanaka, M. Josowicz, J. Kowalik, B. Mizaiakoff, C. Kranz, *Analytical Chemistry* 78 (2006) 1139-1145.
- [35] H.Q.A. Le, S. Chebil, B. Makrouf, H. Sauriat-Dorizon, B. Mandrand, H. Korri-Youssoufi, *Talanta* 81 (2010) 1250-1257.
- [36] Y.Z. Zhang, J. Wang, M.L. Xu, *Colloids and Surfaces B-Biointerfaces* 75 (2010) 179-185.
- [37] J. Wang, S.J. Zhang, Y.Z. Zhang, *Analytical Biochemistry* 396 (2010) 304-309.
- [38] Y.Z. Zhang, K.Y. Zhang, H.Y. Ma, *Analytical Biochemistry* 387 (2009) 13-19.
- [39] W. Chen, Y. Lei, C.M. Li, *Electroanalysis* 22 (2010) 1078-1083.
- [40] R. Janmanee, A. Baba, S. Phanichphant, S. Sriwichai, K. Shinbo, K. Kato, F. Kaneko, *Japanese Journal of Applied Physics* 50 (2011) Part 3 Sp. Iss. SI, 01BK02.
- [41] S.K. Jha, M. Kanungo, A. Nath, S.F. D'Souza, *Biosensors & Bioelectronics* 24 (2009) 2637-2642.
- [42] L. Qu, S.H. Xia, C. Bian, J.Z. Sun, J.H. Han, *Biosensors & Bioelectronics* 24 (2009) 3419-3424.
- [43] B. Liu, L.S. Lu, C.G. Liu, G.M. Xie, *Acta Chimica Sinica* 69 (2011) 438-444.
- [44] R.A. Olowu, O. Arotiba, S.N. Mailu, T.T. Waryo, P. Baker, E. Iwuoha, *Sensors* 10 (2010) 9872-9890.
- [45] B.A. Patel, M. Arundell, K.H. Parker, M.S. Yeoman, D. O'Hare, *Analytical Chemistry* 78 (2006) 7643-7648.
- [46] Y.Z. Wang, S.S. Hu, *Biosensors & Bioelectronics* 22 (2006) 10-17.
- [47] J.H. Shim, H. Do, Y. Lee, *Electroanalysis* 22 (2010) 359-366.
- [48] J. Njagi, J.S. Erlichman, J.W. Aston, J.C. Leiter, S. Andreescu, *Sensors and Actuators B-Chemical* 143 (2010) 673-680.
- [49] A. Aghaei, M.R.M. Hosseini, M. Najafi, *Electrochimica Acta* 55 (2010) 1503-1508.
- [50] Q. Gao, M. Sun, P. Peng, H.L. Qi, C.X. Zhang, *Microchimica Acta* 168 (2010) 299-307.
- [51] S.A. Kumar, S.L. Chen, S.M. Chen, *Electroanalysis* 21 (2009) 1379-1386.
- [52] W. Chen, Z.S. Lu, C.M. Li, *Analytical Chemistry* 80 (2008) 8485-8492.
- [53] A. Curulli, G. Palleschi, In *Proceedings of The 2nd Workshop on Chemical Sensors and Biosensors* (Mazzei, F. and Pilloton, R., eds), (2000) 439-444

- [54] J.Wang, L. Chen, S.B. Hocevar, B. Ogorevc, *Analyst* 125 (2000)1431-1434
- [55] W. Prissanaroon-Ouajai, P.J. Pigram, R. Jones, A. Sirivat, *Sensors and Actuators B-Chemical* 138 (2009) 504-511.
- [56] W. Prissanaroon-Ouajai, P.J. Pigram, R. Jones, A. Sirivat, *Sensors and Actuators B-Chemical* 135 (2008) 366-374.
- [57] G. Herlem, B. Lakard, M. Herlem, B. Fahys, *Journal of The Electrochemical Society* 148 (2001) E435-E43
- [58] R. Aoun, A. Yassin, M. El Jamal, A. Kanj, J. Rault-Berthelot, C. Poriel, *Synthetic Metals* 158 (2008) 790-795.
- [59] G. Herlem, R. Zeggari, J.Y. Rauch, S. Monney, F.T. Anzola, Y. Guillaume, C. Andre, T. Gharbi, *Talanta* 82 (2010) 417-421.
- [60] J. Kim, D.M. Kang, S.C. Shin, M.Y. Choi, S.S. Lee, J.S. Kim, *Analytica Chimica Acta* 614 (2008) 85-92.
- [61] C. Chen, Q.J. Xie, L.H. Wang, C. Qin, F.Y. Xie, S.Z. Yao, J.H. Chen, *Analytical Chemistry* 83 (2011) 2660-2666.
- [62] X.C. Fu, X. Chen, Z. Guo, C.G. Xie, L.T. Kong, J.H. Liu, X.J. Huang, *Analytica Chimica Acta* 685 (2011) 21-28.
- [63] F. Bakhtiarzadeh, S. Ab Ghani, *Electroanalysis* 22 (2010) 549-555.
- [64] H. Zejli, P. Sharrock, J. de Cisneros, I. Naranjo-Rodriguez, K.R. Temsamani, *Talanta* 68 (2005) 79-85.
- [65] Y.H. Li, X.L. Liu, X.D. Zeng, X.Y. Liu, L. Tao, W.Z. Wei, S.L. Luo, *Sensor Letters* 8 (2010) 584-590.
- [66] D.Y. Zheng, C.G. Hu, Y.F. Peng, S.S. Hu, *Electrochimica Acta* 54 (2009) 4910-4915.
- [67] S. Aravamudhan, S. Bhansali, *Sensors and Actuators B-Chemical* 132 (2008) 623-630.
- [68] P. Kalimuthu, S.A. John, *Electrochemistry Communications* 11 (2009) 1065-1068.
- [69] L. Zhang, F.L. Meng, Y. Chen, J.Y. Liu, Y.F. Sun, T. Luo, M.Q. Li, J.H. Liu, *Sensors and Actuators B-Chemical* 142 (2009) 204-209.
- [70] X.P. Wu, W. Liu, H. Dai, G.N. Chen, *Electrochemistry Communications* 11 (2009) 393-396.
- [17] H. Zhao, Z.J. Wu, Y. Xue, Q.A. Cao, Y.J. He, X.J. Li, Z.B. Yuan, *Journal of Nanoscience and Nanotechnology* 11 (2011) 3381-3384.
- [72] A.R. Zanganeh, M.K. Amini, *Sensors and Actuators B-Chemical* 135 (2008) 358-365.
- [73] T.R.L. Dadamos, M.F.S. Teixeira, *Electrochimica Acta* 54 (2009) 4552-4558.
- [74] B. Bahmani, F. Moztarzadeh, M. Hossini, M. Rabiee, M. Tahriri, M. Rezvannia, M. Alizadeh, *Asian Journal of Chemistry* 21 (2009) 923-930.
- [75] I. Tapsoba, S. Bourhis, T. Feng, M. Pontie, *Electroanalysis* 21 (2009) 1167-1176.
- [76] S. Tuncagil, D. Odaci, E. Yidiz, S. Timur, L. Toppare, *Sensors and Actuators B-Chemical* 137 (2009) 42-47.
- [77] P. Sjoberg-Eerola, J. Nylund, J. Bobacka, A. Lewenstam, A. Ivaska, *Sensors and Actuators B-Chemical* 134 (2008) 878-886.

[78] C.Y. Li, Journal of Applied Polymer Science 103 (2007) 3271-3277.



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In recent years, great focus has been placed upon polymer thin films. These polymer thin films are important in many technological applications, ranging from coatings and adhesives to organic electronic devices, including sensors and detectors. Electrochemical polymerization is preferable, especially if the polymeric product is intended for use as polymer thin films, because electrogeneration allows fine control over the film thickness, an important parameter for fabrication of devices. Moreover, it was demonstrated that it is possible to modify the material properties by parameter control of the electrodeposition process. Electrochemistry is an excellent tool, not only for synthesis, but also for characterization and application of various types of materials. This book provides a timely overview of a current state of knowledge regarding the use of electropolymerization for new materials preparation, including conducting polymers and various possibilities of applications.

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