

Roles of SWI/SNF Complex Genes in Breast Cancer

Esra Gunduz¹, Mehmet Gunduz²,
Bunyamin Isik³ and Omer Faruk Hatipoglu⁴

¹*Departments of Medical Genetics, School of Medicine,
Fatih University, Ankara,*

²*Departments of Medical Genetics and Otolaryngology, School of Medicine,
Fatih University, Ankara*

³*Department of Family Medicine, School of Medicine,
Fatih University, Ankara*

⁴*Department of Molecular Biology and Biochemistry, Graduate School of Medicine,
Dentistry and Pharmaceutical Sciences, Okayama
^{1,2,3}Turkey
⁴Japan*

1. Introduction

Cancer is a multifactorial genetic disease which is characterized by uncontrolled proliferation of the cells. Cells undergo mutational changes in a multistep process. Cancer develops from a tumor clone though the firstly mutated cell doesn't present all the features of a cancer cell. Accumulation of the mutations lead cells to display the properties of the cancer. The proliferating cells which have the capacity to survive and invade result in hyperplasia followed by dysplasia and invasion and metastasis at the end [1].

Breast cancer is the most common cancer type and one of the leading cause of cancer mortality in women. Various factors including estrogens and its signaling, EGFR signaling pathway, other oncogenes and tumor suppressor genes including chromatin remodeling factors contribute to development of breast cancer.

At molecular level two major group of genes are responsible for cancer development. These genes, proto- oncogenes and tumor suppressor genes (TSG) control cell growth together in cells at a balance. They are normally required for cell survival and have a direct role in carcinogenesis and cancer progression. When the balance is broken between oncogenes and TSGs due to activation of proto-oncogene or inactivation tumor suppressor genes, cancer develops (**Figure 1, 2**).

In cellular functions proto-oncogenes serve as growth factors, growth factor receptors, transcription factors and signal transduction elements. The mutated proto-oncogenes are named as oncogenes. An oncogene, when mutated or altered, contributes to conversion of a normal cell into a cancer cell. The activation of a proto-oncogene may occur during replication; by a translocation; by gene amplification or by the alterations in mRNA expression. TSGs are also normal cellular genes taking part in regulation of the cell cycle,

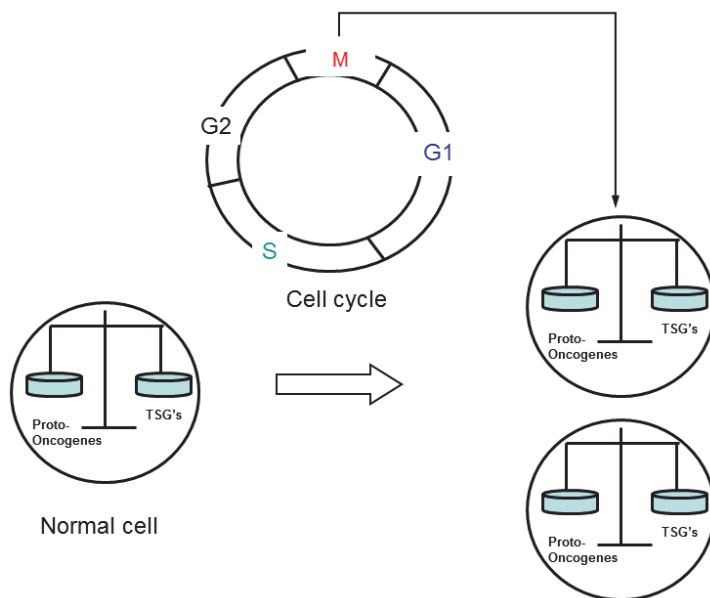


Fig. 1. In human cells proto-oncogenes and tumor suppressor genes are at a balance. There exists a controlled cell division and proliferation.

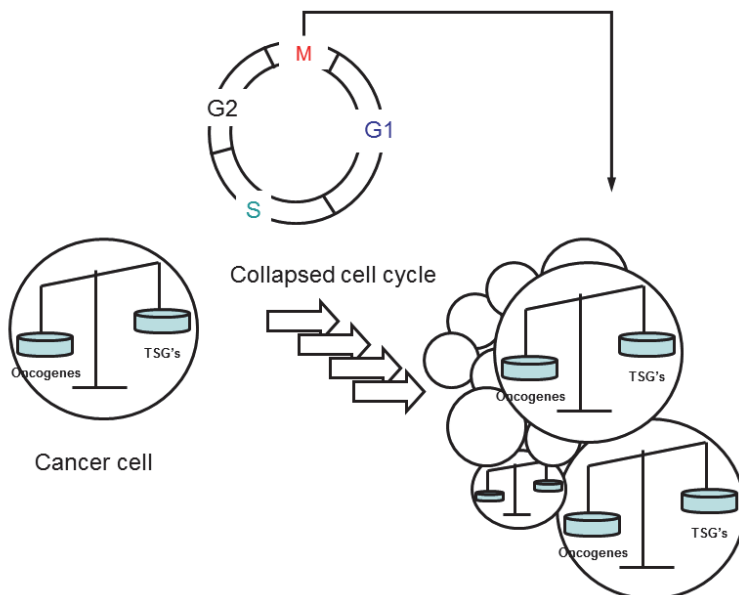


Fig. 2. In a cancer cell, over expression of oncogenes (activation) or low expression of TSGs (inactivation) leads cells to uncontrolled proliferation.

apoptosis, differentiation, surveillance of genomic integrity and repair of DNA errors, chromatin remodeling, signal transduction, and cell adhesion. The activation of the oncogenes and the inactivation of tumor suppressor genes lead cells to proliferate in an uncontrolled manner. Usually one mutation is sufficient for the activation mechanism of oncogenes whereas two hits are necessary for the inactivation of tumor suppressor genes [2,3]. However, a new class of tumor suppressor gene, in which one of the alleles is lost while the rest allele is kept, has recently been defined. Such a tumor suppressor gene is called as haploinsufficient and supposed to be in a cancer-prone state [4-6]. These patients develop cancer when they are exposed to the various carcinogens such as smoking, x-ray and chemicals.

In eukaryotic cells, genetic information encoded by DNA is packaged into chromatin and kept in the nucleus. Thus chromatin is composed of DNA and proteins. The primary proteins of chromatin are histones. A nucleosome, basic unit of chromatin, consists of 146 base-pairs of duplex DNA wrapped around a histone octamer composed of two of each of the conventional histone proteins: H2A, H2B, H3 and H4. Another histon, H1, provides compaction of neighboring nucleosomes by linking them. These compact situation of chromatin reversibly changes in an open and closed situation by various molecules such as histon acetyl transferases (HAT), histon deacetyl transferases (HDAC) and chromatin remodeling molecules, which then influence on transcriptional regulation of gene expression through accessibility of transcription factors by these molecules.

Transcription is an important step to control gene expression from the very early step of life to the end. To maintain transcription every human cell has to deal with the step of an access to DNA either through histone acetylases or chromating remodelling complexes. Many activator proteins of transcription use both of these mechanisms. Histone Acetyl Transferases (HATs) add acetyl groups to the tails of the histones that protrude out of nucleosomes which lead to the binding of the transcription factors. Chromatin remodeling complexes use ATP to open or close the chromatin (**Figure 3**).

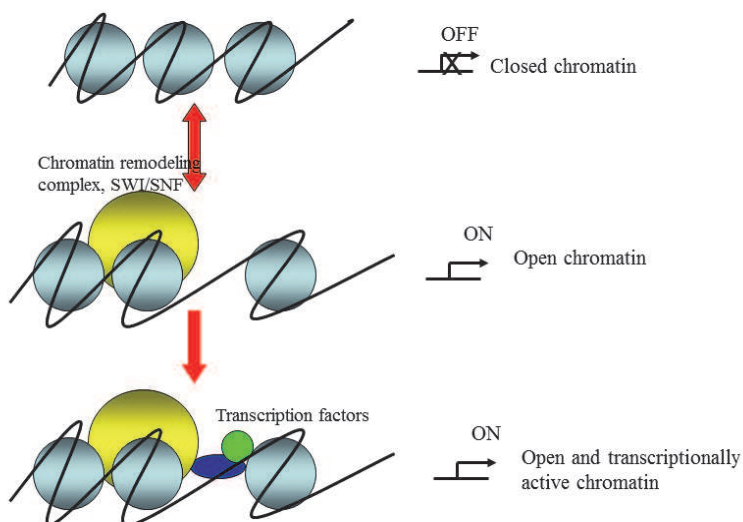


Fig. 3. The binding of chromatin remodeling complex changes conformationally closed chromatin to open chromatin that enables the transcription factors to bind and start transcription.

By cooperation of members of these two classes of complexes, the structure of chromatin is dynamically regulated and thus they play important roles in the control of gene expression. ATP-dependent chromatin remodelers are divided into families according to the subunit composition and biochemical activity such as SWI/SNF, ISWI, INO80, SWR1 and NURD/Mi2/CHD complexes. Of these in particular, some of the members SWI/SNF complexes are emerging tumor suppressors, as genetic and epigenetic inactivation events in several SWI/SNF subunits have been detected in various human cancers [7-10].

2. Function of SWI/SNF family members

Transcription factor action and then the targeted gene expression are mainly regulated by SWI/SNF family of chromatin remodeling complexes. SWI/SNF complexes are large 2-MDa (1.14 MDa in yeast) multi-subunit conglomerates that are involved in either enhancement or suppression of the downstream genes [7-12]. SWI/SNF complex genes were identified through two screens in yeast *Saccharomyces cerevisiae*. The first identified gene that is required for the expression of SUC2 for sucrose metabolism (sucrose non-fermenting (SNF) mutants), and the second screen showed another gene required for the activation of HO for mating-type switching (switch (SWI) mutants) [7, 13-15].

SWI/SNF complex is composed of three groups of subunits; 1) enzymatic (ATPase), 2) core subunits, and 3) accessory subunits [8,11]. Though the exact mechanisms for modification of chromatin structure by SWI/SNF complexes remain incompletely understood, current knowledge suggests that ATPase-dependent disruption of histone-DNA association and resultant nucleosome “sliding” is the main mechanism [8,12]. The mammalian genome encodes 29 different SWI/SNF-like ATPases [12]. Accordingly, each SWI/SNF complex consists of only one of two ATPases, BRM (Brahma) or BRG1 (Brahma-Related Gene 1), which show 74% homology.

SWI/SNF complexes are classified into two major classes as BAF (BRG1 or BRM-Associated Factor; also known as SWI/SNF-A) or PBAF (Polybromo-Associated BAF; also known as SWI/SNF-B) complexes (**Figure 4**). BAF complexes contain either BRG1 (also known as SMARCA4, SNF2b, BAF190) or BRM (also known as SMARCA2, SNF2a) and PBAF complexes include only BRG1 as ATPase subunit. Each ATPase is accompanied with 10 to 12 proteins as core and accessory subunits. The core subunits include BAF155 (also known as SWI3, SRG3, SMARC1), BAF170 (also known as SMARCC2), and SNF5 (also known as SMARCB1, BAF47, INI1). Accessory subunits consist of BAF45 (a,b,c,d; encoded gene names PHF10, DPF1, DPF2, DPF3), BAF53 (a,b; encoded gene names ACTL6A, ACTL6B), BAF57 (encoded gene name SMARCE1), BAF60 (a,b,c; encoded gene name SMARCD1, SMARCD2, SMARCD3), BAF180 (encoded gene name PBRM1), BAF200 (encoded gene name ARID2), BRD7 and BAF250 (a,b; a: also known as ARID1A, SMARCF1, OSA1; b: also known as ARID1B, OSA2) [7,8]. ARID1A (BAF250a) and ARID1B (BAF250b) subunits are mutually exclusive and exist only in BAF complexes. BAF180, BAF200 and BRD7 are exclusively present in PBAF complexes [7,8] [**Figure 4**].

SWI/SNF complexes were found to be based on their roles in the transcription activation. However, studies show that mammalian SWI/SNF complexes have function to both repression and activation of the targeted genes. For development of mammalian T lymphocyte, BRG1 and BAF57 are necessary both for silencing CD4 and activating CD8 expression [7,16,17]. Specific combinations of individual SWI/SNF components were reported to generate sub-complexes with specialized functions that are involved in

SWI/SNF Complexes

SWI/SNF-A (BAF) Complex

SWI/SNF-B (PBAF) Complex

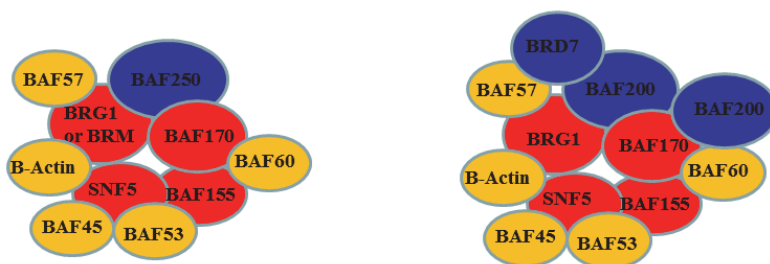


Fig. 4. SWI/SNF complexes are classified into two major classes as BAF (SWI/SNF-A) or PBAF (SWI/SNF-B) complexes. BAF complexes contain either BRG1 or BRM and PBAF complexes include only BRG1 as ATPase subunit. The core subunits include BAF155, BAF170, and SNF5. Accessory subunits consist of BAF45, BAF53, BAF57, BAF60, BAF180, BAF200, BRD7 and BAF250. BAF250a and BAF250b subunits are mutually exclusive and exist only in BAF complexes. BAF180, BAF200 and BRD7 are exclusively present in PBAF complexes.

sequential stages of muscle-gene activation--i.e., initial displacement of the nucleosome followed by the loading of the complete myogenic transcriptosome that promotes gene transcription [18]. Immunoprecipitation analysis of osteocalcin promoter showed that BRM- and BRG1-containing complexes have different roles on it. BRG1 complexes were associated with the promoter induction, while BRM-specific complexes were present only on the repressed promoter and were required for association of the co-repressor HDAC1 [19]. In embryonic stem (ES) cells, BRG1 was reported to act as a repressor to inhibit programmes that are associated with differentiation. On the other hand, it also facilitates the expression of core pluripotency programmes [20,21]. Loss of *Snf5* in murine fibroblasts results in more genes being activated such as E2F targeted genes than repressed [22]. Another example of repression of gene expression is recruitment histone deacetylases (HDACs), which remove activating acetyl marks from histone tails, by SWI/SNF complexes. By this mechanism, SNF5 suppresses cyclin expression in an HDAC1-dependent manner [23]. In conclusion, mammalian SWI/SNF complexes are composed of dynamic units with essential roles in regulating both the activation and the repression of gene expression programmes.

3. Roles of SWI/SNF proteins in cancer

Findings of abnormalities at genetic, epigenetic as well as protein levels of SWI/SNF complexes in various cancers provide a link between chromatin remodelling and tumour suppression. Tumor suppressor role of SWI/SNF complexes was first demonstrated with loss of BRG1 and BRM expression in many cancer cell lines and arrest of growth or slower growth after introduction of BRG1 or hBRM [24]. Brg mutant mice die at early embryonic days due to growth arrest of the inner cell mass and trophoblast [25,26]. Mice with Brg 1

heterozygosity develop mammary adenocarcinomas, suggesting an occurrence of cancer prone state due to haploinsufficiency of Brg1. On the other hand, the mouse with inactivation of BRM by homologous recombination (BRM^{-/-} mice) is born alive and develops normally. Adult mutant mice were approximately 15% heavier than control littermates. This phenomenon was suggested to be caused by increased cell proliferation, because a higher mitotic index was detected in mutant livers and it was further supported by the observation that mutant embryonic fibroblasts were significantly deficient in their ability to arrest in the G0/G1 phase of the cell cycle in response to cell confluency or DNA damage. These studies suggested that BRM plays a role in the regulation of cell proliferation in adult mice and have some defects in control of cellular proliferation [27].

Chromosome transfer studies mapped tumor suppressor gene(s) at 19p13 chromosome locus [28,29]. Studies with microsatellite analysis and functional as well as cancer tissue examination for abnormalities of candidate tumor suppressor gene indicated that chromosome 19p13 locus includes at least two putative tumor suppressor genes namely STK11/LKB1 and BRG1 [30]. STK11 maps about 8.5 Mb distally from BRG1. Loss of heterozygosity of 19p13 was reported in various cancers including thyroid cancer, sex cord stromal tumors, breast cancer, oral carcinoma, prostate cancer, pancreas carcinoma, brain tumors, colorectal carcinoma, gynecological tumors, lung cancers and ovarian carcinoma [31-46]. Some of the studies included genetic analysis of STK11/LKB1 and showed mutation in a subset of tumors especially related with Peutz-Jegher Syndrome such as breast, colorectal, lung, pancreatic, biliary and ovarian cancer [41-49]. On the other hand, quite a lot of studies reported mutations and/or loss or various alterations of BRG1 in human cancer lines and primary tumors [50-61]. Thus genes at this chromosomal locus may involve in various type cancer exclusively or in cooperation in some cancer types. It should be also noticed that some studies showed only LOH without alteration of either one of these genes. In this situation, each of them can still be involved in carcinogenesis due to haploinsufficiency. At least haploinsufficiency of BRG1 is recognized [25-27,62], while further studies are necessary whether such a role exists for STK11/LKB1 or not. Similar to BRG1, abnormalities of BRM in various cancers have been reported [58-61,63-69].

Though the early studies of cell lines and animal models strongly suggested subunits of SWI/SNF proteins as tumor suppressor, the first definitive evidence that members of these complexes function as tumor suppressive was shown by Versteeg and colleagues. They demonstrated occurrence of LOH of BAF47 (SNF5) in almost all cases of pediatric rhabdoid sarcoma, in which the other allele was mutated or silenced by methylation [70]. Inactivation of SNF5 subunit of SWI/SNF is via biallelic mutations, including deletion, nonsense, missense and frameshift mutations was also shown by other studies, supporting SNF5 as a strong tumor suppressor gene at least in this kind of tumors [71-73].

SNF5 alterations have also been shown in other types of tumors though it is much rare as compared to malignant rhabdoid tumors. In a recent study, the effects of Ini1 haploinsufficiency (loss of one allele) on cell growth and immortalization in mouse embryonic fibroblasts were examined. Their results revealed that heterozygosity for Ini1 up-regulated cell growth and immortalization and that exogenous Ini1 down-regulated the growth of primary cells in a Rb-dependent manner. Furthermore, loss of Ini1 was redundant with loss of Rb function in the formation of pituitary tumors in Rb heterozygous mice and gave rise to the formation of large, atypical Rb(+/-) tumor cells lacking adrenocorticotrophic hormone expression, confirming *in vivo* the relationship between Rb and Ini1 in tumor suppression [74]. Mutations and alterations of SNF5 were also reported in familial

schwannomatosis and other cancer types [75-84]. Germ line mutations of SNF5 were detected in brain tumors and rhabdoid tumors, suggesting its link with familial cancers [85-88]. In some other tumors, no alteration of SNF was detected [89,90].

Complete loss of *Snf5* in genetically engineered mouse leads to early embryonic death. However, heterozygote mice with haploinsufficient *Snf5* (*snf5*^{+/-}) develop tumors similar to malignant rhabdoid tumors in about one third of the animals [91-93]. On the other hand, conditional biallelic inactivation of *Snf5* (*Snf5*^{-/-} mice) resulted in tumors including lymphomas and rhabdoid tumors in 100% of mice [94]. Onset of these tumors occurred in a median period of 11 weeks for a single gene inactivation. When compared to this period with most commonly mutated genes in human cancer i.e. p53 and RB1, p53 loss gave rise to lymphomas and sarcomas at 20 weeks and RB1 heterozygosity together with p53 deficiency resulted in similar tumors and other cancers at 16 weeks [95]. Thus shorter onset time for tumor occurrence in *Snf5* inactivation as compared to other well-known tumor suppressors indicates strong tumor suppressor character of this gene. Tumor formation in the absence of SNF5 has been supposed to be due to loss of function of the SWI/SNF complex. However, this view has been challenged by several findings of a recent research. Using both human cell lines and mouse models, Wang et al. [96] showed that cancer formation in the absence of SNF5 does not result from SWI/SNF inactivation but rather that oncogenesis is dependent on continued presence of BRG1 activation than tumor suppressor loss. Thus *Snf5* loss would lead to effects more frequently associated with oncogene activation than tumor suppressor loss.

Other than BRG1 and SNF5, alterations of other member of SWI/SNF complexes have been reported in various cancer types. For example mutations of BAF180 (PBRM1) were identified in 41% of renal cell carcinomas, making this gene as the second most frequently mutated gene in these cancers after VHL50 [97]. The ARID1A subunit of SWI/SNF complexes was also recently shown to have mutation or loss of protein in primary human cancers including ovarian clear cell carcinomas, low and high grade endometrioid carcinomas [98-101]. ARID1A was also rarely mutated in medulloblastoma, breast and lung cancer [102,103].

4. Alterations and roles of SWI/SNF proteins in breast cancer

Breast cancer is among the most common tumors affecting women. It is characterized by a number of genetic aberrations. Some 5-10% of cases are thought to be inherited. Estrogen plays an important role in normal physiology and malignancy of breast tissue. Biological functions of estrogen are mediated by estrogen receptor (ER). ER controls transcription of ER targeted genes by binding to estrogen responsive elements in their promoters. ATP-dependent chromatin remodeling complexes also influence this signaling pathway by changing the chromatin open/close state. In this respect, heterozygous state of a SWI/SNF subunit, Brg1 in mice leads to mammary carcinomas, indicating roles of SWI/SNF proteins in breast cancer [25]. On the other hand, BRCA1 and BRCA2 genes are already known to have roles both in familial and sporadic breast cancers [104-106]. Breast tumors of patients with germ-line mutations in the BRCA1 and BRCA2 genes have more genetic defects than sporadic breast tumors.

Bochar et al. [107] isolated a predominant form of a multiprotein BRCA1-containing complex from human cells displaying chromatin-remodeling activity using a combination of affinity- and conventional chromatographic techniques. Mass spectrometric sequencing of components of this complex proved that BRCA1 is associated with a SWI/SNF-related complex. They also demonstrated that BRCA1 directly interacts with the BRG1 subunit of

the SWI/SNF complex. Furthermore, p53-mediated stimulation of transcription by BRCA1 was completely abrogated by either a dominant-negative mutant of BRG1 or the cancer-causing deletion in exon 11 of BRCA1, revealing that BRCA1 has a direct function in transcriptional control through modulation of chromatin structure [107].

To investigate abnormalities SWI/SNF complex subunits in breast cancer, Decristofaro et al. [108] determined the protein status of the core subunits of BAF170, BAF155, BAF57, BAF53a, and BAF47 in 21 breast cancer cell lines. The authors also determined the protein status of the BRM, BRG1 as well as two other proteins found in human SWI/SNF complexes, BAF180 and BAF250. A breast cancer cell line negative for the BAF57 protein was identified [108].

Deficiency of p270 protein (ARID1A) was shown in a subset of breast cancer. BAF180, a subunit of the PBAP type SWI/SNF chromatin remodeling complex maps to 3p21, in a region where frequent allele loss has been detected in various cancers. A study which used screening for tumor suppressor genes in breast cancer revealed multiple truncating mutations of PB1, which encodes the BAF180 subunit and the mutation was associated with loss of heterozygosity of the wild-type allele [109]. Functional studies showed binding of endogenous wild-type BAF180 to the p21 promoter, which was required for proper p21 expression and G1 arrest after transforming growth factor-beta and gamma-radiation treatment, making BAF180 as a physiologic mediator of p21 expression [109].

In a study, Wang et al. [110] examined the role of BAF57 in breast cancer using the cell line, BT549, which is an invasive human breast carcinoma cell line that lacks expression of BAF57 [111]. They prepared a BT549 stable cell line with expression of the full-length BAF57 protein. The results showed that BT549 clones expressing BAF57 revealed remarkable phenotypic changes, slow growth kinetics, and restoration of contact inhibition. Moreover, microarray analysis showed that BAF57-mediated cell death was associated with up-regulation of proapoptotic genes including the tumor suppressor familial cylindromatosis (CYLD). CYLD was found to be a direct target of BAF57 by chromatin immunoprecipitation analysis. Increased expression of CYLD in BT549 cells induced apoptosis, while its suppression by small interfering RNA inhibited cell death in BAF57 expressing BT549 cells, suggesting the crucial role of BAF57 in cell growth regulation and provided a novel link between hSWI/SNF chromatin remodeling factors and apoptosis [112]. P270 subunit of SWI/SNF complexes was found to be essential for normal cell cycle arrest, providing a direct biological basis to support the implication from tumor tissue screens that deficiency of p270 plays a causative role in carcinogenesis [113]. In a separate study, BAF57 was found to be an ER subtype-selective modulator that specifically regulates ERalpha-mediated transcription, linking ER with SWI/SNF proteins [114].

Harte et al. [115] identified BRD7 as a novel binding partner of BRCA1 with a yeast two-hybrid screen using a BRCA1 bait composed of amino acids 1 to 1142. To determine the functional consequences of the BRCA1-BRD7 interaction, they examined the role of BRD7 in BRCA1-dependent transcription with microarray-based expression profiling. A variety of target genes such as ERalpha was found to be coordinately regulated by BRCA1 and BRD7 complex [115]. In a recent study, two novel mutations were found in one out of 95 breast cancer samples by sequencing BAF57 gene [116].

5. Conclusion and future aspects

Important function of subunits of SWI/SNF complexes arises from their roles in chromatin remodeling and transcription regulation. Mutation and other alterations of these proteins

lead to cancer development. Researches on roles of SWI/SNF subunits in development and cancer are increasingly performed yet much work is necessary for clarifying the exact functions of these genes to provide therapy for various human cancers. Promising results are noticed at the moment for usability of some of these genes as a therapeutic and diagnostic target. Thus progress on the knowledge of functions of subunits of SWI/SNF complexes as well as the relationship with other breast cancer-related molecules such as BRCA1-2 and p53 will clarify their roles in human cancer including breast cancer, which will result in their uses in cancer diagnostics as well as therapy in near future.

6. References

- [1] Renan MJ. How many mutations are required for tumorigenesis? Implications from human cancer data. *Mol Carcinog* 7: 139-146, 1993
- [2] Hinds PW and Weinberg RA. Tumor suppressor genes. *Curr Opin Genet Dev* 4: 135-141, 1994
- [3] Knudson AG Jr. Mutation and cancer: statistical study of retinoblastoma. *Proc Natl Acad Sci USA* 68: 820-823, 1971
- [4] Mduff FK, Hook CE, Tooze RM, Huntly BJ, Pandolfi PP, Turner SD. Determining the contribution of NPM1 heterozygosity to NPM-ALK-induced lymphomagenesis. *Lab Invest*. 2011 Jun 27. doi: 10.1038/labinvest.2011.96. [Epub ahead of print]
- [5] Zhou XZ, Huang P, Shi R, Lee TH, Lu G, Zhang Z, Bronson R, Lu KP. The telomerase inhibitor PinX1 is a major haploinsufficient tumor suppressor essential for chromosome stability in mice. *J Clin Invest* 121(4):1266-82, 2011
- [6] Bouwman P, Drost R, Klijn C, Pieterse M, van der Gulden H, Song JY, Szuhai K, Jonkers J. Loss of p53 partially rescues embryonic development of Palb2 knockout mice but does not foster haploinsufficiency of Palb2 in tumour suppression. *J Pathol* 224:10-21, 2011
- [7] Wilson BG, Roberts CW. SWI/SNF nucleosome remodellers and cancer. *Nat Rev Cancer* 11:481-92, 2011
- [8] Weissman B, Knudsen KE. Hijacking the chromatin remodeling machinery: impact of SWI/SNF perturbations in cancer. *Cancer Res* 69:8223-30, 2009
- [9] Hargreaves DC, Crabtree GR. ATP-dependent chromatin remodeling: genetics, genomics and mechanisms. *Cell Res* 21:396-420, 2011
- [10] Reisman D, Glaros S, Thompson EA. The SWI/SNF complex and cancer. *Oncogene* 28:1653-68, 2009
- [11] Wu JI, Lessard J, Crabtree GR. Understanding the words of chromatin regulation. *Cell* 136:200-6, 2009
- [12] Lessard J, Wu JI, Ranish JA, Wan M, Winslow MM, Staahl BT, Wu H, Aebersold R, Graef IA, Crabtree GR. An essential switch in subunit composition of a chromatin remodeling complex during neural development. *Neuron* 55:201-15, 2007
- [13] Neigeborn L, Carlson M. Genes affecting the regulation of SUC2 gene expression by glucose repression in *Saccharomyces cerevisiae*. *Genetics* 108:845-858, 1984
- [14] Stern M, Jensen R, Herskowitz I. Five SWI genes are required for expression of the HO gene in yeast. *J Mol Biol* 178:853-868, 1984
- [15] Breeden L, Nasmyth K. Cell cycle control of the yeast HO gene: cis- and trans-acting regulators. *Cell* 48:389-397, 1987

- [16] Chi TH, Wan M, Zhao K, Taniuchi I, Chen L, Littman DR, Crabtree GR. Reciprocal regulation of CD4/CD8 expression by SWI/SNF-like BAF complexes. *Nature* 418:195-9, 2002
- [17] Wan M, Zhang J, Lai D, Jani A, Prestone-Hurlburt P, Zhao L, Ramachandran A, Schnitzler GR, Chi T. Molecular basis of CD4 repression by the Swi/Snf-like BAF chromatin remodeling complex. *Eur J Immunol* 39:580-8, 2009
- [18] Albin S, Puri PL. SWI/SNF complexes, chromatin remodeling and skeletal myogenesis: it's time to exchange!. *Exp Cell Res* 316:3073-80, 2010
- [19] Flowers S, Nagl NG Jr, Beck GR Jr, Moran E. Antagonistic roles for BRM and BRG1 SWI/SNF complexes in differentiation. *J Biol Chem* 284:10067-75, 2009
- [20] Ho L, Ronan JL, Wu J, Staahl BT, Chen L, Kuo A, Lessard J, Nesvizhskii AI, Ranish J, Crabtree GR. An embryonic stem cell chromatin remodeling complex, esBAF, is essential for embryonic stem cell self-renewal and pluripotency. *Proc Natl Acad Sci U S A* 106:5181-6, 2009
- [21] Ho L, Jothi R, Ronan JL, Cui K, Zhao K, Crabtree GR. An embryonic stem cell chromatin remodeling complex, esBAF, is an essential component of the core pluripotency transcriptional network. *Proc Natl Acad Sci U S A* 106:5187-91, 2009
- [22] Isakoff MS, Sansam CG, Tamayo P, Subramanian A, Evans JA, Fillmore CM, Wang X, Biegel JA, Pomeroy SL, Mesirov JP, Roberts CW. Inactivation of the Snf5 tumor suppressor stimulates cell cycle progression and cooperates with p53 loss in oncogenic transformation. *Proc Natl Acad Sci U S A* 102:17745-50, 2005
- [23] Zhang ZK, Davies KP, Allen J, Zhu L, Pestell RG, Zagzag D, Kalpana GV. Cell cycle arrest and repression of cyclin D1 transcription by INI1/hSNF5. *Mol Cell Biol* 22:5975-88, 2002
- [24] Dunaief JL, Strober BE, Guha S, Khavari PA, Alin K, Luban J, Begemann M, Crabtree GR, Goff SP. The retinoblastoma protein and BRG1 form a complex and cooperate to induce cell cycle arrest. *Cell* 79:119-30, 1994
- [25] Bultman, S. J. Herschkowitz JI, Godfrey V, Gebuhr TC, Yaniv M, Perou CM, Magnuson T. Characterization of mammary tumors from Brg1 heterozygous mice. *Oncogene* 27:460-468, 2008
- [26] Bultman, S. Gebuhr T, Yee D, La Mantia C, Nicholson J, Gilliam A, Randazzo F, Metzger D, Chambon P, Crabtree G, Magnuson T. A Brg1 null mutation in the mouse reveals functional differences among mammalian SWI/ SNF complexes. *Mol Cell* 6:1287-1295, 2000
- [27] Reyes JC, Barra J, Muchardt C, Camus A, Babinet C, Yaniv M. Altered control of cellular proliferation in the absence of mammalian brahma (SNF2alpha). *EMBO J* 17:6979-91, 1998
- [28] Astbury C, Jackson-Cook CK, Culp SH, Paisley TE, Ware JL. Suppression of tumorigenicity in the human prostate cancer cell line M12 via microcell-mediated restoration of chromosome 19. *Genes Chromosomes Cancer* 31:143 - 55, 2011
- [29] Gao AC, Lou W, Ichikawa T, Denmeade SR, Barrett JC, Isaacs JT. Suppression of the tumorigenicity of prostatic cancer cells by gene(s) located on human chromosome 19p13.1 - 13.2. *Prostate* 38:46 - 54, 1999
- [30] Rodriguez-Nieto S, Sanchez-Céspedes M. BRG1 and LKB1: tales of two tumor suppressor genes on chromosome 19p and lung cancer. *Carcinogenesis* 30:547-54, 2009

- [31] Prazeres HJ, Rodrigues F, Soares P, Naidenov P, Figueiredo P, Campos B, Lacerda M, Martins TC. Loss of heterozygosity at 19p13.2 and 2q21 in tumours from familial clusters of non-medullary thyroid carcinoma. *Fam Cancer* 7:141-9, 2008
- [32] Kato N, Romero M, Catasus L, Prat J. The STK11/LKB1 Peutz-Jegher gene is not involved in the pathogenesis of sporadic sex cord-stromal tumors, although loss of heterozygosity at 19p13.3 indicates other gene alteration in these tumors. *Hum Pathol* 35:1101-4, 2004
- [33] Yang TL, Su YR, Huang CS, Yu JC, Lo YL, Wu PE, Shen CY. High-resolution 19p13.2-13.3 allelotyping of breast carcinomas demonstrates frequent loss of heterozygosity. *Genes Chromosomes Cancer* 41:250-6, 2004
- [34] Dumur CI, Dechsukhum C, Ware JL, Cofield SS, Best AM, Wilkinson DS, Garrett CT, Ferreira-Gonzalez A. Genome-wide detection of LOH in prostate cancer using human SNP microarray technology. *Genomics* 81:260-9, 2003
- [35] Sobottka SB, Haase M, Fitze G, Hahn M, Schackert HK, Schackert G. Frequent loss of heterozygosity at the 19p13.3 locus without LKB1/STK11 mutations in human carcinoma metastases to the brain. *J Neurooncol* 49:187-95, 2000
- [36] Oesterreich S, Allred DC, Mohsin SK, Zhang Q, Wong H, Lee AV, Osborne CK, O'Connell P. High rates of loss of heterozygosity on chromosome 19p13 in human breast cancer. *Br J Cancer* 84:493-8, 2001
- [37] Connolly DC, Katabuchi H, Cliby WA, Cho KR. Somatic mutations in the STK11/LKB1 gene are uncommon in rare gynecological tumor types associated with Peutz-Jegher's syndrome. *Am J Pathol* 156:339-45, 2000
- [38] Gunduz E, Gunduz M, Ouchida M, Nagatsuka H, Beder L, Tsujigiwa H, Fukushima K, Nishizaki K, Shimizu K, Nagai N. Genetic and epigenetic alterations of BRG1 promote oral cancer development. *Int J Oncol* 26:201-10, 2005
- [39] Sentani K, Oue N, Kondo H, Kuraoka K, Motoshita J, Ito R, Yokozaki H, Yasui W. Increased expression but not genetic alteration of BRG1, a component of the SWI/SNF complex, is associated with the advanced stage of human gastric carcinomas. *Pathobiology* 69:315-20, 2001
- [40] Valdman A, Nordenskjöld A, Fang X, Naito A, Al-Shukri S, Larsson C, Ekman P, Li C. Mutation analysis of the BRG1 gene in prostate cancer clinical samples. *Int J Oncol* 22:1003-7, 2003
- [41] Sato N, Rosty C, Jansen M, Fukushima N, Ueki T, Yeo CJ, Cameron JL, Iacobuzio-Donahue CA, Hruban RH, Goggins M. STK11/LKB1 Peutz-Jeghers gene inactivation in intraductal papillary-mucinous neoplasms of the pancreas. *Am J Pathol* 159:2017-22, 2001
- [42] Trojan J, Brieger A, Raedle J, Esteller M, Zeuzem S. 5'-CpG island methylation of the LKB1/STK11 promoter and allelic loss at chromosome 19p13.3 in sporadic colorectal cancer. *Gut* 47:272-6, 2000
- [43] Nishioka Y, Kobayashi K, Sagae S, Sugimura M, Ishioka S, Nagata M, Terasawa K, Tokino T, Kudo R. Mutational analysis of STK11 gene in ovarian carcinomas. *Jpn J Cancer Res* 90:629-32, 1999
- [44] Wang ZJ, Churchman M, Campbell IG, Xu WH, Yan ZY, McCluggage WG, Foulkes WD, Tomlinson IP. Allele loss and mutation screen at the Peutz-Jeghers (LKB1) locus (19p13.3) in sporadic ovarian tumours. *Br J Cancer* 80:70-2, 1999

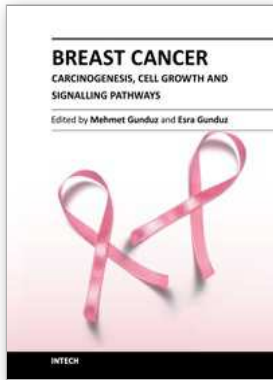
- [45] Gill RK, Yang SH, Meerzaman D, et al. Frequent homozygous deletion of the LKB1/STK11 gene in non-small cell lung cancer. *Oncogene*. 2011 May 2. [Epub ahead of print]
- [46] Papp J, Kovacs ME, Solyom S, Kasler M, Børresen-Dale AL, Olah E. High prevalence of germline STK11 mutations in Hungarian Peutz-Jeghers Syndrome patients. *BMC Med Genet* 11:169, 2010
- [47] Su GH, Hruban RH, Bansal RK, Bova GS, Tang DJ, Shekher MC, Westerman AM, Entius MM, Goggins M, Yeo CJ, Kern SE. Germline and somatic mutations of the STK11/LKB1 Peutz-Jeghers gene in pancreatic and biliary cancers. *Am J Pathol* 154:1835-40, 1999
- [48] Nakanishi C, Yamaguchi T, Iijima T, Saji S, Toi M, Mori T, Miyaki M. Germline mutation of the LKB1/STK11 gene with loss of the normal allele in an aggressive breast cancer of Peutz-Jeghers syndrome. *Oncology* 67:476-9, 2004
- [49] McCarthy A, Lord CJ, Savage K, Grigoriadis A, Smith DP, Weigelt B, Reis-Filho JS, Ashworth A. Conditional deletion of the Lkb1 gene in the mouse mammary gland induces tumour formation. *J Pathol* 219:306-16, 2009
- [50] Wong AK, Shanahan F, Chen Y, et al. BRG1, a component of the SWI-SNF complex, is mutated in multiple human tumor cell lines. *Cancer Res* 60:6171 - 7, 2000
- [51] Rodriguez-Nieto S, Cañada A, Pros E, Pinto AI, Torres-Lanzas J, Lopez-Rios F, Sanchez-Verde L, Pisano DG, Sanchez-Cespedes M. Massive parallel DNA pyrosequencing analysis of the tumor suppressor BRG1/SMARCA4 in lung primary tumors. *Hum Mutat* 32:E1999-2017, 2011
- [52] Schneppenheim R, Frühwald MC, Gesk S, Hasselblatt M, Jeibmann A, Kordes U, Kreuz M, Leuschner I, Martin Subero JL, Obser T, Oyen F, Vater I, Siebert R. Germline nonsense mutation and somatic inactivation of SMARCA4/BRG1 in a family with rhabdoid tumor predisposition syndrome. *Am J Hum Genet* 86:279-84, 2010
- [53] Bartlett C, Orvis TJ, Rosson GS, Weissman BE. BRG1 mutations found in human cancer cell lines inactivate Rb-mediated cell-cycle arrest. *J Cell Physiol* 226:1989-97, 2011
- [54] Medina PP, Romero OA, Kohno T, Montuenga LM, Pio R, Yokota J, Sanchez-Cespedes M. Frequent BRG1/SMARCA4-inactivating mutations in human lung cancer cell lines. *Hum Mutat* 29:617-22, 2008
- [55] Gunduz E, Gunduz M, Nagatsuka H, Beder L, Demircan K, Tamamura R, Hatipoglu OF, Mahmut N, Katase N, Naomoto Y, Nagai N. Epigenetic alterations of BRG1 leads to cancer development through its nuclear-cytoplasmic shuttling abnormalities. *Med Hypotheses* 67:1313-6, 2006
- [56] Medina PP, Carretero J, Fraga MF, Esteller M, Sidransky D, Sanchez-Cespedes M. Genetic and epigenetic screening for gene alterations of the chromatin-remodeling factor, SMARCA4/BRG1, in lung tumors. *Genes Chromosomes Cancer* 41:170-7, 2004. Erratum in: *Genes Chromosomes Cancer*. 2005 Feb;42(2):211.
- [57] Decristofaro MF, Betz BL, Rorie CJ, Reisman DN, Wang W, Weissman BE. Characterization of SWI/SNF protein expression in human breast cancer cell lines and other malignancies. *J Cell Physiol* 186:136 - 45, 2001
- [58] Fukuoka J, Fujii T, Shih JH, Dracheva T, Meerzaman D, Player A, Hong K, Settnek S, Gupta A, Buetow K, Hewitt S, Travis WD, Jen J. Chromatin remodeling factors and BRM/BRG1 expression as prognostic indicators in non-small cell lung cancer. *Clin Cancer Res* 10:4314 - 24, 2004

- [59] Reisman DN, Sciarrotta J, Wang W, Funkhouser WK, Weissman BE. Loss of BRG1/BRM in human lung cancer cell lines and primary lung cancers: correlation with poor prognosis. *Cancer Res* 63:560 - 6, 2003
- [60] Reisman DN, Strobeck MW, Betz BL, Sciarrotta J, Funkhouser W Jr, Murchardt C, Yaniv M, Sherman LS, Knudsen ES, Weissman BE. Concomitant down-regulation of BRM and BRG1 in human tumor cell lines: differential effects on RB-mediated growth arrest vs CD44 expression. *Oncogene* 21:1196 - 207, 2002
- [61] Bock VL, Lyons JG, Huang XX, Jones AM, McDonald LA, Scolyer RA, Moloney FJ, Barnetson RS, Halliday GM. BRM and BRG1 subunits of the SWI/SNF chromatin remodelling complex are downregulated upon progression of benign skin lesions into invasive tumours. *Br J Dermatol* 164:1221-7, 2011
- [62] Glaros S, Cirrincione GM, Palanca A, Metzger D, Reisman D. Targeted knockout of BRG1 potentiates lung cancer development. *Cancer Res* 68:3689-96, 2008
- [63] Liu G, Gramling S, Munoz D, Cheng D, Azad AK, Mirshams M, Chen Z, Xu W, Roberts H, Shepherd FA, Tsao MS, Reisman D. Two novel BRM insertion promoter sequence variants are associated with loss of BRM expression and lung cancer risk. *Oncogene*. 2011 Apr 11. [Epub ahead of print]
- [64] Glaros S, Cirrincione GM, Murchardt C, Kleer CG, Michael CW, Reisman D. The reversible epigenetic silencing of BRM: implications for clinical targeted therapy. *Oncogene* 26:7058 - 66, 2007
- [65] Gunduz E, Gunduz M, Ali MA, Beder L, Tamamura R, Katase N, Tominaga S, Yamanaka N, Shimizu K, Nagatsuka H. Loss of heterozygosity at the 9p21-24 region and identification of BRM as a candidate tumor suppressor gene in head and neck squamous cell carcinoma. *Cancer Invest* 27:661-8, 2009
- [66] Shen H, Powers N, Saini N, et al. The SWI/SNF ATPase Brm is a gatekeeper of proliferative control in prostate cancer. *Cancer Res* 68:10154-62, 2008
- [67] Moloney FJ, Lyons JG, Bock VL, Huang XX, Bugeja MJ, Halliday GM. Hotspot mutation of Brahma in non-melanoma skin cancer. *J Invest Dermatol* 129:1012-5, 2009
- [68] Yamamichi N, Inada K, Ichinose M, et al. Frequent loss of Brm expression in gastric cancer correlates with histologic features and differentiation state. *Cancer Res* 67:10727-35, 2007
- [69] Strobeck MW, Reisman DN, Gunawardena RW, Betz BL, Angus SP, Knudsen KE, Kowalik TF, Weissman BE, Knudsen ES. Compensation of BRG-1 function by Brm: insight into the role of the core SWI-SNF subunits in retinoblastoma tumor suppressor signaling. *J Biol Chem* 277:4782-9, 2002
- [70] Versteeg I, Sévenet N, Lange J, Rousseau-Merck MF, Ambros P, Handgretinger R, Aurias A, Delattre O. Truncating mutations of hSNF5/INI1 in aggressive paediatric cancer. *Nature* 394:203-6, 1998
- [71] Eaton KW, Tooke LS, Wainwright LM, Judkins AR, Biegel JA. Spectrum of SMARCB1/INI1 mutations in familial and sporadic rhabdoid tumors. *Pediatr Blood Cancer*. 56:7-15, 2011
- [72] Jackson, E. M. Sievert AJ, Gai X, Hakonarson H, Judkins AR, Tooke L, Perin JC, Xie H, Shaikh TH, Biegel JA Genomic analysis using high-density single nucleotide polymorphism-based oligonucleotide arrays and multiplex ligation-dependent probe amplification provides a comprehensive analysis of INI1/SMARCB1 in malignant rhabdoid tumors. *Clin Cancer Res* 15:1923-1930, 2009

- [73] Biegel, J. A. Zhou JY, Rorke LB, Stenstrom C, Wainwright LM, Fogelgren B. Germ-line and acquired mutations of INI1 in atypical teratoid and rhabdoid tumors. *Cancer Res* 59:74–79, 1999
- [74] Guidi CJ, Mudhasani R, Hoover K, Koff A, Leav I, Imbalzano AN, Jones SN. Functional interaction of the retinoblastoma and *Ini1/Snf5* tumor suppressors in cell growth and pituitary tumorigenesis. *Cancer Res* 66:8076–82, 2006
- [75] Sevenet N, Lellouch-Tubiana A, Schofield D, Hoang-Xuan K, Gessler M, Birnbaum D, Jeanpierre C, Jouvét A, Delattre O. Constitutional mutations of the *hSNF5/INI1* gene predispose to a variety of cancers. *Am J Hum Genet* 65:1342–1348, 1999
- [76] Trobaugh-Lotrario AD, Tomlinson GE, Finegold MJ, Gore L, Feusner JH. Small cell undifferentiated variant of hepatoblastoma: adverse clinical and molecular features similar to rhabdoid tumors. *Pediatr Blood Cancer* 52:328–334, 2009
- [77] Kohashi K, Izumi T, Oda Y, Yamamoto H, Tamiya S, Taguchi T, Iwamoto Y, Hasegawa T, Tsuneyoshi M. Infrequent *SMARCB1/INI1* gene alteration in epithelioid sarcoma: a useful tool in distinguishing epithelioid sarcoma from malignant rhabdoid tumor. *Hum Pathol* 40:349–55, 2009
- [78] Kohashi, K. Oda Y, Yamamoto H, Tamiya S, Oshiro Y, Izumi T, Taguchi T, Tsuneyoshi M. *SMARCB1/INI1* protein expression in round cell soft tissue sarcomas associated with chromosomal translocations involving *EWS*: a special reference to *SMARCB1/INI1* negative variant extraskeletal myxoid chondrosarcoma. *Am J Surg Pathol* 32:1168–1174, 2008
- [79] Kreiger PA, Judkins AR, Russo PA, Biegel JA, Lestini BJ, Assanasen C, Pawel BR. Loss of *INI1* expression defines a unique subset of pediatric undifferentiated soft tissue sarcomas. *Mod Pathol* 22:142–150, 2009
- [80] Modena P, Lualdi E, Facchinetti F, Galli L, Teixeira MR, Pilotti S, Sozzi G. *SMARCB1/INI1* tumor suppressor gene is frequently inactivated in epithelioid sarcomas. *Cancer Res.* 65:4012–4019, 2005
- [81] Schmitz U, Mueller W, Weber M, Sévenet N, Delattre O, von Deimling A. *INI1* mutations in meningiomas at a potential hotspot in exon 9. *Br J Cancer* 84:199–201, 2001
- [82] Lin H, Wong RP, Martinka M, Li G. Loss of *SNF5* expression correlates with poor patient survival in melanoma. *Clin Cancer Res* 15:6404–11, 2009
- [83] Mobley BC, McKenney JK, Bangs CD, Callahan K, Yeom KW, Schneppenheim R, Hayden MG, Cherry AM, Gokden M, Edwards MS, Fisher PG, Vogel H. Loss of *SMARCB1/INI1* expression in poorly differentiated chordomas. *Acta Neuropathol* 120:745–753, 2010
- [84] Gessi M, Giangaspero F, Pietsch T. Atypical teratoid/rhabdoid tumors and choroid plexus tumors: when genetics "surprise" pathology. *Brain Pathol* 13:409–1, 2003
- [85] Hulsebos TJ, Plomp AS, Wolterman RA, Robanus-Maandag EC, Baas F, Wesseling P. Germline mutation of *INI1/SMARCB1* in familial schwannomatosis. *Am J Hum Genet* 80:805–810, 2007
- [86] Christiaans I, Kenter SB, Brink HC, van Os TA, Baas F, van den Munckhof P, Kidd AM, Hulsebos TJ. Germline *SMARCB1* mutation and somatic *NF2* mutations in familial multiple meningiomas. *J Med Genet* 48:93–7, 2011
- [87] Bourdeaut F, Lequin D, Brugières L, et al. Frequent *hSNF5/INI1* germline mutations in patients with rhabdoid tumor. *Clin Cancer Res* 17:31–8, 2011

- [88] Rousseau G, Noguchi T, Bourdon V, Sobol H, Olschwang S. SMARCB1/INI1 germline mutations contribute to 10% of sporadic schwannomatosis. *BMC Neurol* 11:9, 2011
- [89] Yamamoto H, Kohashi K, Tsuneyoshi M, Oda Y. Heterozygosity Loss at 22q and Lack of INI1 Gene Mutation in Gastrointestinal Stromal Tumor. *Pathobiology* 78:132-9, 2011
- [90] Mori N, Inoue K, Okada M, Motoji T. Absence of Mutations on the SNF5 Gene in Hematological Neoplasms with Chromosome 22 Abnormalities. *Acta Haematol* 126:69-75, 2011
- [91] Roberts CW, Galusha SA, McMenamin ME, Fletcher CD, Orkin SH. Haploinsufficiency of Snf5 (integrase interactor 1) predisposes to malignant rhabdoid tumors in mice. *Proc Natl Acad Sci USA* 97:13796-13800, 2000
- [92] Klochendler-Yeivin, A. Fiette L, Barra J, Muchardt C, Babinet C, Yaniv M. The murine SNF5/INI1 chromatin remodeling factor is essential for embryonic development and tumor suppression. *EMBO Rep* 1:500-506, 2000
- [93] Guidi, C. J. Sands AT, Zambrowicz BP, Turner TK, Demers DA, Webster W, Smith TW, Imbalzano AN, Jones SN. Disruption of *Ini1* leads to peri-implantation lethality and tumorigenesis in mice. *Mol Cell Biol* 21:3598-3603, 2001
- [94] Roberts CW, Leroux MM, Fleming MD, Orkin SH. Highly penetrant, rapid tumorigenesis through conditional inversion of the tumor suppressor gene *Snf5*. *Cancer Cell* 2:415-425, 2002
- [95] Williams BO, Remington L, Albert DM, Mukai S, Bronson RT, Jacks T. Cooperative tumorigenic effects of germline mutations in *Rb* and *p53*. *Nat Genet* 7:480-4, 1994
- [96] Wang X, Sansam CG, Thom CS, Metzger D, Evans JA, Nguyen PT, Roberts CW. Oncogenesis caused by loss of the SNF5 tumor suppressor is dependent on activity of BRG1, the ATPase of the SWI/SNF chromatin remodeling complex. *Cancer Res* 69:8094-101, 2009
- [97] Varela I, Tarpey P, Raine K, et al. Exome sequencing identifies frequent mutation of the SWI/SNF complex gene *PBRM1* in renal carcinoma. *Nature* 469:539-42, 2011
- [98] Wiegand KC, Lee AF, Al-Agha OM, Chow C, Kalloger SE, Scott DW, Steidl C, Wiseman SM, Gascoyne RD, Gilks B, Huntsman DG. Loss of *BAF250a* (*ARID1A*) is frequent in high-grade endometrial carcinomas. *J Pathol* 224:328-33, 2011
- [99] Guan B, Mao TL, Panuganti PK, Kuhn E, Kurman RJ, Maeda D, Chen E, Jeng YM, Wang TL, Shih IeM. Mutation and loss of expression of *ARID1A* in uterine low-grade endometrioid carcinoma. *Am J Surg Pathol* 35:625-32, 2011
- [100] Wiegand KC, Shah SP, Al-Agha OM et al. *ARID1A* mutations in endometriosis-associated ovarian carcinomas. *N Engl J Med* 363:1532-1543, 2010
- [101] Jones, S. Wang TL, Shih IeM, et al. Frequent mutations of chromatin remodeling gene *ARID1A* in ovarian clear carcinoma. *Science* 330:228-231, 2010
- [102] Parsons, D. W. Li M, Zhang X, et al. The Genetic Landscape of the Childhood Cancer Medulloblastoma. *Science* 331:435-439, 2011
- [103] Huang J, Zhao YL, Li Y, Fletcher JA, Xiao S. Genomic and functional evidence for an *ARID1A* tumor suppressor role. *Genes Chromosom Cancer* 46:745-750, 2007
- [104] Wang F, Fang Q, Ge Z, Yu N, Xu S, Fan X. Common *BRCA1* and *BRCA2* mutations in breast cancer families: a meta-analysis from systematic review. *Mol Biol Rep*. 2011 Jun 4. [Epub ahead of print]

- [105] Nanda R. Targeting^h triple-negative breast cancer: the lessons learned from BRCA1-associated breast cancers. *Semin Oncol* 38:254-62, 2011
- [106] Milne RL, Antoniou AC. Genetic modifiers of cancer risk for BRCA1 and BRCA2 mutation carriers. *Ann Oncol Suppl* 1:i11-7, 2011
- [107] Bochar DA, Wang L, Beniya H, Kinev A, Xue Y, Lane WS, Wang W, Kashanchi F, Shiekhhattar R. BRCA1 is associated with a human SWI/SNF-related complex: linking chromatin remodeling to breast cancer. *Cell* 102:257-65, 2000
- [108] Decristofaro MF, Betz BL, Rorie CJ, Reisman DN, Wang W, Weissman BE. Characterization of SWI/SNF protein expression in human breast cancer cell lines and other malignancies. *J Cell Physiol* 186:136-45, 2001
- [109] Wang X, Nagl NG Jr, Flowers S, Zweitzig D, Dallas PB, Moran E. Expression of p270 (ARID1A), a component of human SWI/SNF complexes, in human tumors. *Int J Cancer* 112:636, 2004
- [110] Xia W, Nagase S, Montia AG, Kalachikov SM, Keniry M, Su T, Memeo L, Hibshoosh H, Parsons R. BAF180 is a critical regulator of p21 induction and a tumor suppressor mutated in breast cancer. *Cancer Res* 68:1667-74, 2008
- [111] Kiskinis E, García-Pedrero JM, Villaronga MA, Parker MG, Belandia B. Identification of BAF57 mutations in human breast cancer cell lines. *Breast Cancer Res Treat* 98:191-8, 2006
- [112] Wang L, Baiocchi RA, Pal S, Mosialos G, Caligiuri M, Sif S. The BRG1- and hBRM-associated factor BAF57 induces apoptosis by stimulating expression of the cylindromatosis tumor suppressor gene. *Mol Cell Biol* 25:7953-65, 2005
- [113] Nagl NG Jr, Patsialou A, Haines DS, Dallas PB, Beck GR Jr, Moran E. The p270 (ARID1A/SMARCF1) subunit of mammalian SWI/SNF-related complexes is essential for normal cell cycle arrest. *Cancer Res* 65:9236-44, 2005
- [114] García-Pedrero JM, Kiskinis E, Parker MG, Belandia B. The SWI/SNF chromatin remodeling subunit BAF57 is a critical regulator of estrogen receptor function in breast cancer cells. *J Biol Chem* 281:22656-64, 2006
- [115] Harte MT, O'Brien GJ, Ryan NM, Gorski JJ, Savage KI, Crawford NT, Mullan PB, Harkin DP. BRD7, a subunit of SWI/SNF complexes, binds directly to BRCA1 and regulates BRCA1-dependent transcription. *Cancer Res* 70:2538-47, 2010
- [116] Villaronga MA, López-Mateo I, Markert L, Espinosa E, Fresno Vara JA, Belandia B. Identification and characterization of novel potentially oncogenic mutations in the human BAF57 gene in a breast cancer patient. *Breast Cancer Res Treat*. 2011 Apr 5. [Epub ahead of print]



Breast Cancer - Carcinogenesis, Cell Growth and Signalling Pathways

Edited by Prof. Mehmet Gunduz

ISBN 978-953-307-714-7

Hard cover, 732 pages

Publisher InTech

Published online 30, November, 2011

Published in print edition November, 2011

Cancer is the leading cause of death in most countries and its consequences result in huge economic, social and psychological burden. Breast cancer is the most frequently diagnosed cancer type and the leading cause of cancer death among females. In this book, we discussed various aspects of breast cancer carcinogenesis from clinics to its hormone-based as well as genetic-based etiologies for this deadly cancer. We hope that this book will contribute to the development of novel diagnostic as well as therapeutic approaches.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Esra Gunduz, Mehmet Gunduz, Bunyamin Isik and Omer Faruk Hatipoglu (2011). Roles of SWI/SNF Complex Genes in Breast Cancer, Breast Cancer - Carcinogenesis, Cell Growth and Signalling Pathways, Prof. Mehmet Gunduz (Ed.), ISBN: 978-953-307-714-7, InTech, Available from: <http://www.intechopen.com/books/breast-cancer-carcinogenesis-cell-growth-and-signalling-pathways/roles-of-swi-snf-complex-genes-in-breast-cancer>

INTECH

open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.