

Inflammatory Responses and Regulation in Parkinson's Disease

Lynda J. Peterson and Patrick M. Flood
*The University of North Carolina at Chapel Hill,
U.S.A.*

1. Introduction

Parkinson's disease (PD) is a slow, progressive neurodegenerative disorder affecting an estimated 6 million people worldwide (Litteljohn, Mangano et al. 2011). The etiology of the disease is characterized by increasing loss over time of dopaminergic neurons (DA-neurons) in the substantia nigra (SN) as well as the depletion of dopamine in the striatum, which eventually leads to pathological and clinical symptoms (Jenner and Olanow 2006). PET imaging and post-mortem analyses of the brains of PD patients indicate that the appearance of symptoms, including tremor, bradykinesia, rigidity, slowness of movement, and postural instability (Jellinger 2001), generally are manifest once 60% of DA-neurons have died and a 70% threshold decrease in normal DA activity has been reached (Klockgether 2004; Litteljohn, Mangano et al. 2011). Epidemiological studies indicate that only about 10% of PD cases are early onset, i.e. prior to the age of 50 and occur mainly in familial clusters (Mizuno, Hattori et al. 2001). These cases have established genetic bases due to mutations in several recently identified genes, including parkin, leucine-rich repeat kinase 2 (LRRK2), α -synuclein, PINK-1, or DJ-1 (Polymeropoulos, Lavedan et al. 1997; Lucking, Durr et al. 2000; Abou-Sleiman, Healy et al. 2003; Farrer, Haugarvoll et al. 2006; Sun, Latourelle et al. 2006; Jiang, Wu et al. 2007; Weng, Chou et al. 2007; Bonifati, Wu-Chou et al. 2008). The majority of PD cases (approximately 90%) are late onset and idiopathic (Tanner 2003). Although the etiology of idiopathic PD is uncertain, this form of PD particularly affects the elderly, with average onset of clinical symptoms between 60 and 65 years of age (Litteljohn, Mangano et al. 2011). Idiopathic PD is thus age-associated, with approximately 1% of the population being affected by 65-70 year of age, increasing to 4-5% at 85 years (Fahn 2003; Tansey, McCoy et al. 2007). However, the causes of idiopathic non-familial PD are probably multifactorial, with some form of genetic predisposition, environmental insults and/or aging all likely to be important factors in disease initiation and progression (Nagatsu and Sawada 2006; Dickson 2007; Vilar, Coelho et al. 2007; Singh, Singh et al. 2008).

While the exact cause of chronic neurodegeneration of PD is not known, increasing evidence suggests that chronic inflammation is the fundamental process mediating the progressive nature of the neurodegeneration characteristic of PD (McGeer, Yasojima et al. 2001; Hirsch and Hunot 2009). In animal PD models, long-term inflammation induces progressive loss of DA neurons within the SN brain region (Gao, Jiang et al. 2002; Dauer and Przedborski 2003; Bartels and Leenders 2007). Neuroinflammation, which is characterized by activation of

both the innate and adaptive immune response, has been demonstrated in PD patients by the preponderance of activated microglia within the SN, and the increased production in the CNS of inflammatory mediators, such as cytokines, chemokines, reactive oxygen species (ROS) and reactive nitrites (Cicchetti, Lapointe et al. 2005; Loeffler, Camp et al. 2006; Ghosh, Roy et al. 2007; Cicchetti, Drouin-Ouellet et al. 2009). Post mortem analyses of brains from PD patients provide clear evidence of large numbers of human leukocyte antigen (HLA-DR) and CD11b-positive microglia in the SN, the brain region in which the degeneration of DA neurons is most prominent (McGeer, Itagaki et al. 1988). Expression of the inflammatory markers MHC II, TNF α and cyclooxygenase (COX-2) is also higher in SN tissue of patients with PD than in comparable tissue from unaffected controls (McGeer, Itagaki et al. 1988; Boka, Anglade et al. 1994; Knott, Stern et al. 2000). Additionally, levels of proinflammatory mediators, including TNF α , IL-1 β , IL-6, (ROS), and eicosanoids, are all elevated in the SN, striatum, cerebrospinal fluid (CSF) and/or peripheral blood mononuclear cells (PBMC) of PD patients (McGeer, Itagaki et al. 1988; Qureshi, Baig et al. 1995; Mogi, Harada et al. 1996; Nagatsu, Mogi et al. 2000; Imamura, Hishikawa et al. 2003; Teismann and Schulz 2004; Nagatsu and Sawada 2006; Sawada, Imamura et al. 2006; Hirsch and Hunot 2009; Pisani, Moschella et al. 2010). Nitric oxide (NO) free radicals are also elevated in PD as indicated by increased nitrite (an indicator for NO) present in the CSF, as well as increased expression of inducible nitric oxide synthase (iNOS) within the SN of PD patients (Qureshi, Baig et al. 1995). Oxidative damage measured by the presence of nucleoside oxidation product 8-hydroxyguanosine, is approximately 16-fold higher in SN of PD patients and post-mortem analysis of the brains of PD patients also show evidence of the oxidation of lipids, DNA and proteins (Owen, Schapira et al. 1996; Spencer, Jenner et al. 1998; Halliwell 2006). Reduced levels of antioxidant glutathione (GSH) concomitant with increased concentrations of oxidized GSH, are found in the surviving SN neurons of PD patients compared to normal controls, indicating these neurons were undergoing increased oxidative stress (OS) at the time of death (Sofic, Lange et al. 1992; Pearce, Owen et al. 1997).

In addition to direct evidence for increased inflammatory activity within the CNS of patients with PD, other aspects of the etiology of PD suggest this is primarily an inflammatory disorder. For example, PD patients display decreased activity in mitochondrial complexes I and I/III (Krige, Carroll et al. 1992), and the inhibition of mitochondria respiratory complex-I after exposure to MPTP gives rise to PD-like pathologies in humans (Tetrud, Langston et al. 1989; Langston, Forno et al. 1999), primates, mice and rats (Yazdani, German et al. 2006; Jackson-Lewis and Przedborski 2007). This is because DA neurons appear to have an intrinsic sensitivity to complex I defects based on studies that demonstrate the selective neurotoxic effects of the pesticide rotenone on DA neurons only, despite the fact that rotenone inhibits complex I throughout the brain (Sherer, Betarbet et al. 2002). However, even direct inhibition of complex-I by MPTP in DA neurons results in neurodegeneration that is primarily inflammatory in nature (Ghosh, Roy et al. 2007; Qian, Gao et al. 2007; Qian and Flood 2008). Further evidence of the key role of inflammation in PD is seen in other features of the disease. For example, blood-brain-barrier (BBB) leakage has been found in positron-emission tomography (PET) images of the brains of PD patients (Kortekaas, Leenders et al. 2005), and is likely caused by increased inflammation in PD. In addition, increased angiogenesis also occurs in PD patients (Faucheux, Bonnet et al. 1999), and in the SN of a monkey-model of PD (Barcia, Bautista et al. 2005). Increased expression of the angiogenic-stimulant vascular endothelial growth factor (VEGF) and its major receptor

VEGFR1, have been found post mortem in the SN of PD patients (Wada, Arai et al. 2006), and this is consistent with increased angiogenesis found in PD brains (Faucheux, Bonnet et al. 1999). Excess VEGF induces increased macrophage migration mediated by VEGFR1 (Huusko, Merentie et al. 2009) and VEGF also induces BBB disruption in animal models after brain injury (Zhang, Zhang et al. 2000; Nguyen, Julien et al. 2002; Rite, Machado et al. 2007; Argaw, Gurfein et al. 2009) as well as in a rat model of PD (Rite, Machado et al. 2007). The VEGFR antagonist, Cyclo-VEGI, reduces inflammation and vascular leakiness, and is neuroprotective against excitotoxin-induced neurodegeneration in rats (Ryu and McLarnon 2008). This suggests dysregulated VEGF expression in the PD brain is also a key part of the neuroinflammatory response and increased edema in PD (Kirk and Karlik 2003; Suidan, Dickerson et al. 2010) (Nguyen, Julien et al. 2002; Nguyen, Tatlipinar et al. 2006; Zhang, Wang et al. 2006).

It is also worth noting that the innate immune cells of the brain, including microglia, become more reactive with age and that this might contribute to increased neuroinflammation in the elderly at some time point after activation of the peripheral immune response (Godbout and Johnson 2009). It has been suggested that prolonged exposure over a lifetime to inflammatory cytokines in the brain might eventually lead to various neurologic disorders (Godbout and Johnson 2009). This increased susceptibility to chronic inflammation in the elderly could contribute to the increased risk of PD associated with age. With growing elderly populations in many countries, PD is potentially an expanding public health problem. It is therefore critical that therapeutic interventions be identified and developed, that can halt and eventually reverse the neurodegeneration characteristic in PD. Treatments that focus on reducing chronic inflammation and pro-inflammatory microglial cell activation offer great promise for this sort of therapy. Therapies that foster microglia phenotypes which are non-inflammatory and supportive of neuroregeneration, could also prove beneficial in treating PD in the future.

In this chapter, we will discuss inflammatory mechanisms that play a crucial role in the progression of PD, and the therapeutic efficacy of using several anti-inflammatory compounds to treat progressive PD. We will also discuss the biology of microglial cells, including the cellular and molecular mechanisms that activate and regulate their inflammatory response, the evidence that implicates their different inflammatory mediators in the destruction of DA-neurons, and the efficacy of using different anti-inflammatory cytokines such as IL-10 and 1β , natural anti-inflammatory compounds such as *sinomenine*, *luteolin*, and *curcumin*, and the anti-inflammatory drugs morphinans and cyclosporine A, cell-based treatments such as Treg therapy, and specific anti-inflammatory therapies such as NF- κ B inhibitors and β 2-Adrenergic Receptor (β 2AR) agonists. in halting or reversing the degenerative effects of inflammation within the SN. Finally, we will propose a model for PD as it relates to chronic inflammation, and discuss some possible future directions for the therapeutic treatment of PD.

2. Neuroinflammation in Parkinson's Disease

2.1 Microglia and chronic inflammation in neurodegeneration

Over two decades ago, high levels of activated microglia were discovered in the SN of PD patients (McGeer, Itagaki et al. 1988), first suggesting a connection between inflammation and PD. Since then, a wide range of inflammatory mediators produced by active microglia

have been identified post mortem in PD tissue samples (Hirsch 2000; Orr, Rowe et al. 2002). The presence of activated microglia has also been verified in the SN of PD animal models (rodents and primates) (Akiyama and McGeer 1989; Barcia, Sanchez Bahillo et al. 2004; Gao, Miao et al. 2011). Studies using animal models of PD have begun to elucidate the inflammatory mechanisms that underlie the neurodegeneration of DA neurons and loss of DA production which are hallmarks of the disease. It has been found that intranigral injection of the pathogen product lipopolysaccharide (LPS) induces increased levels of CD11b+ and microglia and neutrophils in the SN of rats when compared to levels found in the cortex (Ji, Kronenberg et al. 2009). This same study also observed pronounced damage to endothelial cells and BBB permeability within the SN of the LPS-injected rats (Ji, Kronenberg et al. 2009). In this model, direct intranigral injection LPS or administration of the neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) induced PD-like symptoms accompanied by increased activation of microglia in the SN. Of particular significance is another finding that activated microglia and elevated levels of pro-inflammatory TNF α could be detected in the SN of monkeys from 1 to 14 years after an initial brief administration of MPTP (McGeer, Schwab et al. 2003; Barcia, Sanchez Bahillo et al. 2004; Barcia, de Pablos et al. 2005). These observations suggested that a brief insult to the brain by toxins could induce a chronic inflammatory state with concomitant progressive neurodegeneration. Additional support for sustained inflammation after insult is provided by an unfortunate human example of accidental exposure to MPTP in contaminated illegal street-drugs. Post-mortem evidence of activated microglia and degeneration in the SN of these human patients remained evident from 3 to 16 years after intravenous injection of the toxin (Langston, Forno et al. 1999).

Among the many inflammatory factors produced by microglia, superoxide is one of the major mediators of neurodegeneration. DA neurons are especially vulnerable to oxidative insults and this strongly supports the association of microglia activation with progressive PD (Block and Hong 2007). It is known that the normal midbrain region encompassing the SN contains nearly five times more microglia than other areas of the brain (Kim, Mohney et al. 2000). Understanding the progressive nature of microglia-mediated neurotoxicity, and the common mechanism of microglia activation by various diverse toxins or insults, thus has prime therapeutic importance for treating PD (Nguyen, Julien et al. 2002).

2.2 Inflammatory mechanisms and dopaminergic-neuron death

Several different animal models of PD have found that both systemic inflammation and direct dopaminergic neurotoxicity can initiate or exacerbate the neuroinflammation and neurodegeneration that are hallmarks of the disease. Injection of MPTP results in chronic neuroinflammation and progressive neurotoxicity in humans (Langston, Forno et al. 1999) and other primates (McGeer, Schwab et al. 2003). In the former case, an accidental intravenous injection of MPTP-contaminated street-drugs left several patients ill with progressive motor-dysfunction which was shown in post mortem analysis to be mediated by DA-neuron loss in the SN, very similar to what is observed following MPTP injection in both rodent and primate models of neurodegeneration (Langston, Forno et al. 1999; Qin, Wu et al. 2007). Similarly a single systemic injection of LPS in an rodent model results in significant loss (23%) of DA neurons in the SN beginning at 7-months post treatment, increasing to a 47% loss at 10-months post injection (Qin, Wu et al. 2007). This delayed, progressive loss of DA neurons in the SN recapitulates some of the hallmark characteristics

of PD. This model is further supported as a model for PD by a clinical case report that a patient displayed PD-related symptoms after accidental peripheral exposure to LPS (Niehaus and Lange 2003). However, the underlying mechanism that advances the progressive nature of the disease remains unclear. It is known that systemically injected LPS cannot readily cross the blood-brain barrier, and studies have shown that pro-inflammatory cytokines, actually mediate the mechanism of systemic LPS-induced DA neurotoxicity (Qin, Wu et al. 2007). Interestingly, systemic administration of LPS enhances motor neuron degeneration in animal models of another neurodegenerative disease, amyotrophic lateral sclerosis (ALS), 6 months after LPS injection (Nguyen, D'Aigle et al. 2004), suggesting this may be a common etiology for a number of chronic neurodegenerative conditions. In addition, systemic exposure to LPS was also shown to significantly exacerbate neuronal-cell death associated with ischemic insult in neonatal rats (Lehnardt, Massillon et al. 2003). The effects of systemic inflammation on neuronal survival may depend upon several factors such as brain region affected, length of time to allow cumulative effects, age of exposure to the inflammagen, presence of systemic TNF α , and severity of the inflammatory stimulation.

It now appears that it is the inflammatory cytokines TNF α , IL-1 β and IL-6 which play major roles in the etiology of PD both systemically and within the CNS. First, TNF α produced in the periphery after systemic LPS injection is transported across BBB to enter the brain through a TNF α -receptor dependent mechanism (Gutierrez, Banks et al. 1993; Pan and Kastin 2002; Qin, Wu et al. 2007). TNF α then initiates an inflammatory cascade by interacting with TNF α receptors on the microglia, leading to the activation of transcription factor NF- κ B within microglial cells, and resulting in the production of additional TNF α and other pro-inflammatory factors, and thus creating a persistent and self-perpetuating neuroinflammatory response that drives delayed and progressive loss of DA neurons in the SN (Park and Bowers 2010). In addition, TNF α is one of the primary pro-inflammatory cytokines that promote overproduction of ROS (Fernandez-Checa, Kaplowitz et al. 1997), a key player in DA-neurodegeneration (Gao and Hong 2008) (Qian, Flood et al. 2011). Another pro-inflammatory cytokine, IL-1 β , is also under transcriptional control of NF- κ B and is consequently up-regulated in neuroinflammation. Direct injection of recombinant IL-1 β into the brains of rats induces astrocytic expression of hypoxia inducible factor (HIF-1 α) which contributes to the induction of oxidative stress (Argaw, Zhang et al. 2006). IL-1 β also induces the HIF-1 α -VEGF pathway that results in increased BBB permeability and increased angiogenesis (Argaw, Zhang et al. 2006; Argaw, Gurfein et al. 2009), which are all found to be increased in PD patients (Kortekaas, Leenders et al. 2005). Similarly, IL-6 expression is controlled by NF- κ B (Lappas, Permezel et al. 2002) and is increased under a variety of inflammatory conditions (Van Snick 1990; Dendorfer 1996), including an MPTP-induced mouse model of PD (Kohutnicka, Lewandowska et al. 1998). IL-6 expression from activated microglia coincides in the SN with loss of TH+ DA neurons after MPTP injection but is absent from the SN in uninjected controls (Kohutnicka, Lewandowska et al. 1998). Neuronal levels of IL-6 are also increased by oxidative stress which is another mediator of microglia activation, neuroinflammation and degeneration (Lee, Cho et al. 2010; Naik and Dixit 2011; Negi, Kumar et al. 2011).

In addition to inflammatory cytokines, oxidative stress appears to play a crucial role in the DA-neurodegeneration seen in PD. Oxidative stress occurs because of intracellular accumulation of reactive oxygen and nitrogen species (ROS & RNS) and, along with

mitochondrial dysfunction and inflammation, has been identified as central in the mechanism underlying the inflammatory pathogenesis of PD (Bartels and Leenders 2007; Monahan, Warren et al. 2008). Impaired mitochondrial function has been observed in PD (Schapira, Cooper et al. 1989; Schapira 1994; Jenner and Olanow 1996; Sherer, Betarbet et al. 2002; Keeney, Xie et al. 2006; Schapira 2006), in which mitochondria generate ROS as by-products of molecular oxygen consumption in the electron transport chain. Oxidative stress is hypothesized to initiate DA neuron loss in the SN (Jenner and Olanow 1998; Lin and Beal 2006). The brain is hypersensitive to oxidative damage in part due to the fact that oxygen consumption by brain cells constitutes 20% of total oxygen consumption in body. Brain tissue is also enriched in peroxidizable fatty acids and has low levels anti-oxidant defenses (catalase, SOD, glutathione, glutathione peroxidase) (Floyd 1999; Floyd 1999). Within the brain, the SN is amongst regions most vulnerable to oxidative insult. The SN is characterized by a pro-oxidative state, and has a high metabolic rate with high levels of the oxidizable species DA, DA-derived ROS, neuromelanin and other molecules that render the SN vulnerable to the possible effects of peroxynitrite and sulfite (Marshall, Reist et al. 1999). Oxidative stress is part of the pro-inflammatory response of microglia within the CNS, and it has been found that NADPH oxidase (PHOX), is the major superoxide-producing enzyme of microglia. It is a molecular complex of membrane-associated cytochrome b558 (composed of 2 subunits: gp91phox and p22phox) and the cytosolic components: p47phox, p67phox, p40phox, and a small GTPase rac2 (Groemping and Rittinger 2005). Upon activation, the cytosolic subunits translocate to the membrane and form a functional enzyme to generate superoxide. Therapy directed against the oxidative stress response has proved beneficial in treatment for PD (Smith and Zigmond 2003; Jackson-Lewis and Smeyne 2005). Anti-inflammatory agents that inhibit NADPH oxidase activity are neuroprotective (Liu and Hong 2003; Qian, Block et al. 2006; Qian, Gao et al. 2007), and reduced DA neurotoxicity induced by LPS or MPTP has been demonstrated in PHOX^{-/-} compared to wild-type mice (Gao, Liu et al. 2003; Qin, Liu et al. 2004). Pharmacological inhibition of PHOX by the specific inhibitor diphenyliodonium (DPI), also shows potent DA-neuroprotection *in vitro* (Qian, Gao et al. 2007). PHOX activity also regulates production of pro-inflammatory cytokines, such as TNF α , by activated microglia *in vitro* following LPS stimulation (Qin, Liu et al. 2004). This suggests that PHOX not only mediates superoxide production, but also controls the levels of other pro-inflammatory neurotoxic factors produced by activated microglia. Cell signaling pathways that regulate PHOX activity are still being elucidated but it has been shown that a microglial adhesion molecule, the integrin MAC1 (macrophage antigen complex 1), is closely linked with PHOX and plays an important role in microglia-mediated neuroinflammation and neurotoxicity (Hu, Zhang et al. 2008). Microglial-cell expressed MAC1 is indispensable for the enhanced neurotoxicity induced by LPS, α -synuclein, or MPTP in neuron-glia mixed cell cultures (Pei, Pang et al. 2007; Zhang, Dallas et al. 2007; Hu, Zhang et al. 2008). Furthermore, MAC1-deficient mice show resistance to MPTP-induced DA neurotoxicity *in vivo*. NADPH oxidase-generated oxygen free radicals are also required for MAC1-mediated phagocytosis in neutrophils (Coxon, Rieu et al. 1996). Therefore, the dual activity between MAC1 and NADPH oxidase might be a central mechanism underlying the reactive microgliosis that mediates oxidative damage and consequent progressive neurodegeneration after microglia activation.

2.3 Peripheral inflammation, microglial-cell activation and neurodegeneration

Neuroinflammation is an important defense mechanism against pathogens and environmental toxins damaging the brain. Under normal conditions, the central nervous

system (CNS) and the brain especially are considered to be immune privileged (Neuwelt and Clark 1978; Smith, DeGirolami et al. 1992; Lotan and Schwartz 1994). Any immune response is very highly regulated with a finely balanced anti-inflammatory environment within the CNS combined with vigilant immune surveillance by circulating immune cells and those that are resident in the CNS. Under pathological conditions, the brain mounts an aggressive, acute inflammatory response to invading pathogens, infection, trauma, stroke or any other threat to homeostasis. These perturbations trigger the activation of microglia, the brain resident macrophages (Ransohoff and Perry 2009), as well as local invasions of circulating immune cells and the production of ROS and RNS (Mosley, Benner et al. 2006; Tansey, McCoy et al. 2007; Whitton 2007). The resident microglia aid in functional regulation of other immune cells particularly the helper T-cells and dendritic cells (DC) which also play critical roles in the initiation and maintenance of any neuroinflammatory response to pathological threats. However, inflammation in the brain is a 'two-edged sword' (Wyss-Coray and Mucke 2002) because if the tight regulation of the inflammatory process breaks down in any manner, the inflammatory response can switch from being beneficial to highly detrimental to brain function. In acute situations short-lived inflammatory mechanisms limit injury and promote healing (Wyss-Coray and Mucke 2002; McGeer and McGeer 2004). Microglia are the critical elements in an acute neuroinflammatory response that minimize tissue damage, restore tissue to homeostasis and promote wound healing processes. Conversely, when inflammation is chronically sustained at high levels, this can lead to brain-tissue damage with concomitant initiation and progression of neuroinflammatory diseases such as PD.

While microglial cells are the major immunocompetent cells in the CNS and are important mediators in the pathogenesis of PD (Herrera, Tomas-Camardiel et al. 2005; Smeyne, Jiao et al. 2005; Zhou, Wang et al. 2005), up-regulation of humoral response and other peripheral immune components has been identified in both idiopathic and genetic cases of PD (Orr, Rowe et al. 2005). Elevated levels of IL-1 β , TNF α , IL-6 have been described in post mortem tissue from PD patients (Mogi, Harada et al. 1994; Mogi, Harada et al. 1994; Mogi, Harada et al. 1996), and PD patients show higher levels of IL-1 β , TNF α , IL-1, CD4 $^{+}$ and CD8 $^{+}$ cells in serum and CSF (Dobbs, Charlett et al. 1999; Bas, Calopa et al. 2001; Hisanaga, Asagi et al. 2001; Reale, Greig et al. 2009). CD8 $^{+}$ T cells have also been found associated with degenerating SN neurons (McGeer, Itagaki et al. 1988), and classical complement components have been isolated in the Lewy bodies that form in the SN of PD patients (Yamada, McGeer et al. 1992). Significantly, compromised BBB integrity has also been observed in PD patients (Kortekaas, Leenders et al. 2005), indicating that factors in peripheral inflammation have ready access to the brain and may also be involved in the neurodegenerative process probably via an effect on the activation of microglia. Complement proteins normally circulate in the blood in an inactive state, but in response to pathogenic threat, these proteins become activated in cascades of events that precipitate destruction and removal of the threat. Importantly for PD, studies have shown that complement factors C1q and mannose-binding lectins (MBL), participate in microglial cell activation and could be important in pathogenesis of neurodegeneration (Yamada, McGeer et al. 1992; Webster, Galvan et al. 2001; Whitton 2007). The importance of the classical complement pathway in PD has been established (Yamada, McGeer et al. 1992; Yamada, Chong et al. 1993), and MBL infiltration into the brain accompanies degeneration of TH $^{+}$ neurons in MPTP-injected mice up to 28 days post MPTP administration in an animal model of PD (Chao, He et al. 2009). In the latter study, clear evidence of BBB breakdown was also

documented, providing further evidence that peripheral immunity probably participates in the activation of microglia and consequent DA-neuron destruction.

Normally, immune surveillance protects the healthy brain from pathogen assault and damage. Memory T-cell surveillance of the CNS is first regulated via migration from the blood into the cerebral spinal fluid (CSF) through the choroid plexus barrier (Engelhardt and Ransohoff 2005), while the recruitment and infiltration of immune cells into brain tissue occurs through the post-capillary venules into the perivascular space that surrounds microvascular endothelial cells (MVEC) which are joined together by tight-junctions that form the blood-brain-barrier (Carrithers, Visintin et al. 2000; Pachter, de Vries et al. 2003). MVEC in the brain closely interact with astrocytes, pericytes, perivascular microglia and neuronal cells such as DA-producing neurons to form the neurovascular unit. Thus, at these crucial checkpoints the specific microenvironment within the CNS tissues determines whether the immune cells survive to mount antigen-specific inflammatory response or if they will be eliminated. CD4⁺ T cells play a critical role in the initiation and maintenance of any immune response in the CNS including the brain, but to survive in this normally anti-inflammatory environment, they need additional stimulatory signals. Generally, CD4⁺ must be activated in the peripheral lymphoid system prior to migration into the CNS. Activated CD4⁺Th cells differentiate into distinct effector subsets that express unique suites of cytokines that perform different immunoregulatory functions. For example Th1 and Th2 produce predominantly INF- γ and IL-4 respectively which have differential effects on the activation state of microglia amongst other disparate outcomes (Appel, Beers et al. 2009)

T cells express many sensor receptors with which they assess the microenvironment milieu. CD4⁺ interact with other immune cells such as antigen presenting cells (APC) that take-up, process and present antigens via the major histocompatibility complex molecules (MHC). MHCII is expressed mainly in cells of the lymphoid system such as T cells, B cells, macrophages, and DC (Matarese, De Rosa et al. 2008). Under non-inflammatory conditions, cells of the CNS express low levels of MCH and in the absence of histocompatibility molecules the survival or further differentiation of activated T cells is low. Constitutively expressed mediators also help prevent differentiation of Th cells, and conversely promote development of Tregs. For example, the neuropeptide Vasoactive Intestinal Peptide (VIP) up-regulates Treg function which has an inhibitory effect on microglia and macrophages (Delgado, Chorny et al. 2005; Fernandez-Martin, Gonzalez-Rey et al. 2006). Additionally, anti-inflammatory cytokines secreted by astrocytes and glial cells, such as IL-10 and TGF β 1, induce differentiation of Tregs (Chen and Wahl 2003). However, during inflammation TGF β 1 works with IL-6 or IL-21 to induce differentiation of pro-inflammatory IL-17-producing T cells (Th17) indicating the distinction between anti-inflammatory and pro-inflammatory is not always straight forward. In an animal model of PD, the DA-neuron neurodegenerative effects of MPTP administration were Th17-cell mediated, and the induction of Tregs by VIP attenuated this effect (Reynolds, Banerjee et al. 2007; Reynolds, Stone et al. 2010). However, as it has also been shown that TGF β 1/IL-6 increases production of IL-10 as well as IL-17, this IL-10 is then capable of suppressing the degenerative effects mediated by Th17 cells (McGeachy, Bak-Jensen et al. 2007).

3. Efficacy of anti-inflammatory agents halting or reversing degeneration

Mounting evidence indicates that numerous types of inflammatory mediators such as TNF α , prostaglandin (PGE₂), NO, free radicals, and other products of activated immune

cells can also play a role in the DA-neuron degeneration in several models of PD (Gao, Jiang et al. 2002; Dauer and Przedborski 2003; Bartels and Leenders 2007). Treatment using anti-inflammatory agents directed at a number of different pro-inflammatory targets could potentially halt, slow or even reverse PD disease progression. For example, steroidal anti-inflammatory drugs (SAIDS) such as dexamethasone have been used to stimulate neuroprotection against MPTP or LPS-induced toxicity (Kurkowska-Jastrzebska, Wronska et al. 1999; Castano, Herrera et al. 2002; Kurkowska-Jastrzebska, Litwin et al. 2004). In addition, non-steroidal anti-inflammatory drugs (NSAIDS), e.g. aspirin and ibuprofen, can reduce inflammation by inhibition of COX activity (Sairam, Saravanan et al. 2003)(Sairam et al. 2003). A prospective study found a significantly lower risk of developing PD in users of ibuprofen compared to non-users, and that the efficacy of taking ibuprofen was dose-dependently related to the number of weekly tablets of the drug taken (Gao, Chen et al. 2011). Microglial cell inhibitors such as minocycline have also shown neuroprotective characteristics in PD animal models (Du, Ma et al. 2001; Wu, Jackson-Lewis et al. 2002). Recently, strategies that directly inactivate the activation of the pro-inflammatory transcription factor NF- κ B (Ghosh, Roy et al. 2007; Zhang, Qian et al. 2010), and/or activate the peroxisome proliferator-activated receptor-c (PPARc) (Bernardo, Ajmone-Cat et al. 2005), have produced beneficial modulation of inflammatory responses. Other strategies, including inhibition of ion channels in microglial cells (Thomas, Chartrand et al. 2007) can also be effective treatments aimed at halting progressive PD in an animal model of the disease. We will focus on five relatively new approaches in anti-inflammatory therapy for PD: (1) the use of exogenous anti-inflammatory cytokines IL-10 or TGF β 1, (2) the use of cell-based therapies such as regulatory T cells (Tregs), which are the major source of IL-10 or TGF β 1 in the resolution of neuroinflammation; (3) the use of morphinan-related compounds; (4) the targeting of NF- κ B, the major transcriptional regulator of inflammation, and (5) the administration of beta2-adrenergic receptor (β 2AR) agonists.

3.1 Therapies using anti-inflammatory cytokines: IL-10 and TGF β

When acute inflammation is not resolved, i.e. the agent or event that originated the inflammatory response has not been eliminated, chronic inflammation can become entrenched and lead to neuroinflammation and degeneration. Normally, endogenous anti-inflammatory cytokines provide a negative-feedback mechanism which controls the continued activation of immune cells from potentially pathological effects (Moore, Lahiri et al. 2001; Strle, Zhou et al. 2001). Therefore, replacing or elevating the level of these anti-inflammatory regulators could be a therapeutic means to halt the chronic, pathological aspects of inflammation. IL10 and TGF β 1 are two major anti-inflammatory cytokines that are normally produced by Treg cells. Studies using exogenously supplied IL10 and TGF β 1 have shown potent effects in reducing neurotoxicity induced by either LPS or MPTP in PD models. In an *in vitro* model of PD, application of IL-10 to a mixed glial cell-neuron cell culture abrogated the degeneration of the neuron cells induced by either LPS or MPTP (Qian, Block et al. 2006). This inhibitory effect of IL10 was mediated through its inhibition of the production of TNF α , nitric oxide, and extracellular superoxide in the microglia cells within the mixed cell culture (Qian, Block et al. 2006). These *in vitro* effects of IL-10 are confirmed by *in vivo* results from a 6-OHDA rat model of PD. In this model, sustained administration of IL10 via a viral-vector, significantly protected DA-neuron from death and ameliorated behavioral deficits induced by intra-striatal delivery of 6-OHDA (Johnston, Su et al. 2008).

Several *in vitro* studies have shown that TGF β 1 can also protect neurons from cell death induced by oxidative stress (Prehn and Kriegelstein 1994), glutamate excitotoxicity (Zhu, Yang et al. 2002), or chemical-induced hypoxia (Ruocco, Nicole et al. 1999). TGF β 1 suppresses the progression of neurodegeneration *in vivo* in an EAE model of MS (Szczepanik, Tutaj et al. 2005). Additionally, recombinant TGF β 1, intracerebrally delivered by a viral-vector, protects against brain injury induced by ischemia (Unsicker and Kriegelstein 2002), excitotoxic-induced death (Ruocco, Nicole et al. 1999), and oxidative stress (Henrich-Noack, Prehn et al. 1996). Although exogenous TGF β 1 has thus been strongly suggested as a neuroprotective treatment, the molecular mechanisms underlying its neuroprotection have not been clearly elucidated. Recent evidence indicates that the neuroprotective effects of both IL10 and TGF β 1 are mainly due to their inhibition of ROS production in microglia during initial activation or in reactive microgliosis (Qian, Block et al. 2006; Qian, Wei et al. 2008). In mixed glial cell-neuronal cell cultures, application of TGF β 1 blocked neuron cell death through the inhibition of PHOX activity in cultures exposed to either LPS or MPTP. TGF β 1 prevented the ERK-dependent phosphorylation on p47phox in the microglial cells and blocked translocation and assembly of the PHOX molecular complex to the plasma membrane. Inhibition of PHOX activation consequently reduced oxidase activities induced by LPS (Qian, Wei et al. 2008). While the complete *in vivo* roles of IL10 and TGF β 1 in the regulation of chronic CNS inflammation in PD remain to be determined, both may provide promising basis for therapeutic use in PD treatments.

3.2 Morphinan-based anti-inflammatory therapeutics

Dextromethorphan (DM) is a well established drug that is used as the active ingredient in many cough suppressants. Over twenty years ago, it was recognized that DM and the metabolite dextrorphan (DX) could have anti-seizure effects in models of convulsive disorders such as epilepsy (Tortella, Ferkany et al. 1988). Dextrorphan was also shown to antagonize the *in vitro* excitation of spinal neurons by application of N-methyl-D-aspartate (NMDA), a glutamate receptor ligand (Church, Lodge et al. 1985). In addition, DX was found to block cortical neuron injury induced by NMDA or by glutamate (Choi 1987; Choi, Maulucci-Gedde et al. 1987; Choi, Peters et al. 1987; Koh and Choi 1987). Because NMDA receptors were known to mediate hypoxic injury in neuronal cell cultures, it was suggested that morphinan drugs, especially DX and DM, might be potential therapeutic treatments for brain injury from hypoxia of ischemia (Goldberg, Weiss et al. 1987). Additionally, since the loss of DA-neuron in PD leads to the secondary effect of hyperactive glutamatergic function, it was postulated that DM and DX might also be useful as adjunct therapies in treating PD (Albin, Young et al. 1989; Greenamyre and O'Brien 1991).

Several studies using PD animal models or *in vitro* cell cultures have shown that dextromethorphan and its metabolites are neuroprotective due to their anti-inflammatory properties and inhibitory function towards microglia activation (Liu, Qin et al. 2003; Zhang, Wang et al. 2004; Zhang, Qin et al. 2005). In the first of these studies, DM treatment protected DA-neurons from LPS-induced neuron death in mixed glial-neuronal cell cultures (Liu, Qin et al. 2003). Furthermore, the neuroprotective effects of DM were mediated through inhibiting microglial cell activation (Liu, Qin et al. 2003). Similarly, DM inhibited microglia activation and was neuroprotective when administered daily to mice that had been injected with MPTP (Zhang, Wang et al. 2004). Another metabolite of DM, 3-hydroxymorphinan (3-HM), was shown to have the greatest potency (of several tested

methorphanins) for attenuating the loss of DA-neurons in the SN, as well as restoring motor functions in an MPTP-injection mouse-model of PD (Zhang, Qin et al. 2005). 3-HM also protected neuronal cells in mixed glial-neuronal cells cultures by reducing MPTP-induced microgliosis and decreasing the production of ROS. In addition to this neuroprotective property, 3-HM was also found to have neurotrophic effects. The neurotrophic effects of 3-HM were mediated by induction of increased expression of neurotrophic factors, including GDNF, EGF, NTF and TGF β 1, by astroglial cells in mixed cultures with neuronal cells (Zhang, Wang et al. 2004; Zhang, Shin et al. 2006). In another study using mixed neuron-glia cultures, both l-morphine and its synthetic stereoisomer, d-morphine, reduced LPS or MPTP-induced DA-neuron death with similar efficacy (Qian, Tan et al. 2007). These results indicated that morphine exerts anti-inflammatory effects either by inhibition of direct microglial cell activation via LPS or through attenuating reactive microgliosis induced by MPTP. A naturally occurring d-morphinan, sinomenine, also has neuroprotective effects and will be discussed below (Qian, Xu et al. 2007).

While DM and other morphinan compounds appear to have strong neuroprotective properties in animal model systems of PD, early human trials using DM to treat PD patients were contradictory. While one study found daily high doses of DM could improve some motor-behavioral deficits in PD patients (Bonuccelli, Del Dotto et al. 1992), another similar trial using the same dose regimen reported no beneficial effects on PD motor-associated symptoms (Montastruc, Fabre et al. 1994). Significantly, more recent double-blind trials have confirmed that DM can improve motor function in PD patients, especially the dyskinesia associated with long-term levodopa treatment (Verhagen Metman, Blanchet et al. 1998; Verhagen Metman, Del Dotto et al. 1998). It has been suggested that the uncertainty over the true effects of DM might be due to the relatively rapid metabolism of DM *in vivo* (Werling, Lauterbach et al. 2007). Although, DX and other DM metabolites have shown similar neuroprotective properties in preclinical studies, only DM seems to function through multiple mechanisms rather than simply blocking activity of the NMDA-receptor (Werling, Lauterbach et al. 2007). *In situ* DM concentrations can be increased if given in conjunction with a low dose of the drug quinidine which retards metabolism of DM and leads to increased DM concentrations in plasma, resulting in greater bioavailability (Pope, Khalil et al. 2004). Clearly, DM presents an attractive potential for use in combinatorial treatment for at least ameliorating the motor deficits characteristic of PD and possibly for attenuating the activation of microglia and chronic inflammation also prominent in PD.

3.3 Sinomenine, luteolin and curcumin as treatments for Parkinson's disease

Several natural compounds isolated from plants, including sinomenine, luteolin and curcumin, are also being investigated for their ability to attenuate inflammation and provide neuroprotection against the loss of DA neurons. Although these three agents belong to different families of organic compounds, they have all shown anti-oxidant, anti-inflammatory and neuroprotective qualities *in vitro* cell-culture models of neurodegenerative diseases including PD. The first of these is sinomenine, a morphinan related alkaloid compound purified from a medicinal plant (Sinomenine acutum) that has been traditionally used to treat inflammatory disorders (Liu, Resch et al. 1994; Liu, Buchner et al. 1996). In a study using rat midbrain mixed glial cell-neuron cell cultures, sinomenine protected the neuronal cells from both LPS and MPP⁺ induced cell death (Qian, Xu et al. 2007). Sinomenine protected neurons by reducing release of extracellular ROS and inhibiting PHOX/p47 from translocation to the plasma membrane where the complex becomes

activated. These effects were mediated by inhibition of microglial cells since sinomenine failed to protect neuronal-cell enriched cultures which lacked microglia from MPP⁺ induced damage and death (Qian, Xu et al. 2007).

Another plant extract, the flavonoid luteolin, has also been used for its anti-oxidant and anti-inflammatory properties. Luteolin was demonstrated to have neuroprotective effects by inhibiting oxidative stress-induced cell death in the DA-producing SH-SY5Y cell line (Kang, Lee et al. 2004). In mixed glial-neuron midbrain cell cultures from rats, luteolin attenuated the loss of TH⁺ DA-neurons after addition of increasing concentrations of LPS. The protective effect of luteolin in these experiments was shown to be mediated through the inhibition of microglial cell activation, and reduced production of TNF α , NO and ROS (Chen, Jin et al. 2008). Similarly, curcumin, the active anti-inflammatory isolate from turmeric (*Curcuma longa*), is known to have anti-inflammatory neuroprotective effects in CNS disorders. In an experimental allergic encephalomyelitis (EAE) mouse model of MS, curcumin inhibited EAE development by attenuating microglia activation and IL-12 production as well as Th1 cell differentiation (Natarajan and Bright 2002). Curcumin also has shown protective effects against LPS-induced DA neuron cell death in mixed rat neuron-glial cell cultures which were mediated by inhibition of the production of proinflammatory factors in microglia (Yang, Zhang et al. 2008). Taken together, the neuroprotective results from these three natural plant products suggest an interesting possibility for PD therapeutics. Since these and other similar plant-based isolates can cross the BBB and have anti-inflammatory as well as anti-oxidant properties, they may provide new directions for adjunct therapy.

4. Anti-inflammatory cell-based strategy for Parkinson's disease therapy: Regulatory T-cells

An anti-inflammatory strategy currently being studied as a cell-based therapy in PD involves the therapeutic introduction of Treg cells. It is now believed that Tregs suppress immune reactivity through multiple mechanisms, such as release of IL-10 and TGF β , induction of apoptotic tolerance, and suppression of metabolic functions in effector immune cells such as microglia and effector T-cells. As the primary source IL10 and TGF1 β *in vivo*, Tregs have been shown to be the major cells which regulate the inflammatory response in a number of disorders through their effects on the innate and adaptive immune responses that have escaped normal pathways of control. Recent studies using models of neurodegeneration demonstrated that induction of an anti-inflammatory Treg response inhibited microglial activation, and promoted neuronal survival (Reynolds, Banerjee et al. 2007; Liu, Gong et al. 2009; Reynolds, Stone et al. 2009). In another report, adoptively transferred Treg cells attenuated a Th17-mediated inflammatory response in mice that had been injected with MPTP and concomitantly vaccinated with nitrated (N) α -synuclein. In this model, injection of N- α -synuclein elicited an adaptive immune response in conjunction with the MPTP-induced neurotoxicity both of which were ameliorated by the transfer of natural or VIP-induced Treg cells in these mice (Reynolds, Stone et al. 2010). Tregs have also been shown to promote neurotrophic factor production from astrocytes (Reynolds, Banerjee et al. 2007; Reynolds, Stone et al. 2010), indicating their potential for neuroregeneration of DA neurons.

Strategies that use Th2 cells are also being employed. Th2 cells inhibit microglial cell activation through the production of IL4 and IL10, and stimulate the production of GDNF by astrocytes,

thereby providing neuroprotection against MPTP-induced neuronal death (Benner, Mosley et al. 2004). Copaxone, a peptide-based therapy approved for patients with MS, is thought to promote the development of Th2 cells which function to decrease CNS inflammation through the release of anti-inflammatory cytokines and neurotrophic factors (Kipnis and Schwartz 2002; Angelov, Waibel et al. 2003; Benner, Mosley et al. 2004; Schwartz 2004). Copaxone-induced T-cells also have neuroprotective effects in animal models of ALS and PD (Angelov, Waibel et al. 2003; Benner, Mosley et al. 2004). However, because the interactions amongst regulatory T cells, glial cells, neurons and other infiltrating leukocytes within the SN is incredibly complex and not well understood, further studies to elucidate the regulatory pathways involved are necessary to develop Th2-cell based therapies for PD patients.

5. Specific anti-inflammatory therapies

5.1 Therapies targeting pro-inflammatory transcription factor NF- κ B

Many of the inflammatory mediators involved in inflammation and DA-neurodegeneration in PD are expressed in microglial cells and their regulation is primarily mediated by the transcription factor NF- κ B. NF- κ B was first described in 1986 as a transcription factor which is essential for the expression of mouse kappa light chain genes (Sen and Baltimore 1986; Sen and Baltimore 1986). It has since been shown that NF- κ B functions to control gene expression of many of pro-inflammatory mediators (Tsoulfas and Geller 2001). Inflammatory cytokines such as TNF α and IL-1 α and β , bacterial products such as lipopolysaccharide (LPS), and products of cellular damage strongly activate inflammatory responses through the activation of NF- κ B. NF- κ B subsequently plays an essential positive-feedback role in the inflammatory response through regulation of genes encoding inflammatory cytokines IL-1 β , TNF α , and IL-12/23, as well as chemokines IL-8, MIP-1 α , and MCP-1 (Xia, Pauza et al. 1997; Roebuck 1999; Roebuck, Carpenter et al. 1999). NF- κ B also mediates nitric oxide production (iNOS), expression of NADPH oxidase subunits p47 and p67 (Gauss, Nelson-Overton et al. 2007; Lawrence 2009), and adhesion molecules ICAM-1, VCAM, and E-selectin (Chen and Manning 1995; Tak and Firestein 2001). Activation of NF- κ B is a key event in many chronic inflammatory diseases such as cardiovascular disease (Van der Heiden, Cuhlmann et al.), tissue reperfusion injury (Latanich and Toledo-Pereyra 2009), experimental autoimmune encephalomyelitis (EAE) (Vandenbroeck, Alloza et al. 2004), rheumatoid arthritis (Criswell 2010), and inflammatory bowel disease (IBD) (Atreya, Atreya et al. 2008). A large number of the therapeutic agents for treating human inflammatory conditions, including sulfasalazine, 5-aminosalicylates, and corticosteroids, as well as some natural anti-inflammatory compounds such as IL-10, TGF β 1, β 2AR agonists, glutamate, and curcumin, may owe their anti-inflammatory effects to inhibition of NF- κ B (Lawrence and Fong ; Wang, Boddapati et al. ; Pereira and Oakley 2008; Lawrence 2009). These anti-inflammatory agents are potent inhibitors of microglial activation, and are neuroprotective to DA neurons *in vitro* and *in vivo*. Clearly, NF- κ B activity presents a key target to ameliorate chronic inflammation in humans. Therapeutic strategies to inhibit NF- κ B activity in microglial cells may lead to more effective treatments for PD (Zhang, Qian et al. 2010; Flood, Qian et al. 2011).

The NF- κ B family consists of dimeric transcription factors which include five members: c-Rel, RelA (p65), RelB, NF- κ B1 (p50/p105), and NF- κ B2 (p52/p100) (Flood, Qian et al. 2011). There are two major activation pathways: (1) the classical or canonical pathway, and (2) the

alternate or noncanonical pathway. While the non-canonical pathway does not appear to play a major role in the activation of inflammation, the classical pathway is thought to regulate the production of most pro-inflammatory mediators and is characterized by activation of a dimer of Rel proteins p50 and p65. In the inactive state this Rel dimer is complexed within the cytosol to the inhibitory protein I κ B α complex (Lawrence 2009). The classical NF- κ B pathway is initiated upon phosphorylation, ubiquitination, and subsequent proteasome-dependent degradation of I κ B α . The phosphorylation of I κ B α on serine residues is mediated by I κ B kinase (IKK), which is a molecular complex of three proteins consisting of a heterodimer of the two catalytic subunits IKK α and IKK β , along with IKK γ (the NF- κ B essential modulator, NEMO) (May, Marienfeld et al. 2002; Huxford and Ghosh 2009; Oeckinghaus and Ghosh 2009). Embryonic cells derived from genetic knock-out mice lacking IKK β , IKK γ , or p65 are unresponsive to classical NF- κ B inducers such as TNF α and IL-1 β (Reuther-Madrid, Kashatus et al. 2002; Sizemore, Lerner et al. 2002; Sizemore, Agarwal et al. 2004). Activation of IKK in response to inflammatory mediators such as TNF α , IL-1 β , and LPS, depends critically on the presence of the IKK γ (NEMO) subunit of the IKK complex (Rudolph, Yeh et al. 2000; May, Marienfeld et al. 2002), which results in the phosphorylation of the I κ B by the kinase activity of IKK β (Huxford and Ghosh 2009; Oeckinghaus and Ghosh 2009). An N-terminal region of NEMO associates with a hexapeptide sequence within the C-terminus of both IKK α and IKK β (NEMO binding domain or NBD), and disruption or mutation of this NEMO-NBD interaction site on either IKK β or IKK γ results in a loss of responsiveness of cells to pro-inflammatory signaling (Flood, Qian et al. 2011). Agents that block the activation of NF- κ B are capable of inhibiting the two major inflammatory pathways in microglia—activation of oxidative stress and production of inflammatory mediators, including cytokines TNF α , IL-1 β , IL-6, as well as chemokines associated with inflammation (Qian, Flood et al. 2010).

Selective IKK β and IKK γ inhibitors that do not target IKK α or the non-canonical P100/p52 pathway, should be promising therapeutic agents for treating chronic inflammatory disorders including PD. Such specific NF- κ B inhibitors have recently been used in murine models of PD to halt the progression of neurodegeneration induced by the neurotoxin MPTP (Ghosh, Roy et al. 2007), or by activation of CNS inflammation by the intracranial injection of LPS (Zhang, Qian et al. 2010). Pretreatment of animals with a peptide against the NEMO-binding domain (NBD peptide) prior to injection of MPTP into mice, significantly inhibits the activation of NF- κ B within the midbrain region. This inhibition of NF- κ B activation is accompanied by a concomitant reduction in inflammatory mediator mRNA expression within the SN, as well as the expression of microglial cell activation marker CD11b. Mice receiving the NBD peptide prior to MPTP injection also showed highly significant protection of the nigrostriatum from MPTP-induced neurodegeneration of the TH $^+$ neurons and the loss of dopamine production, as well as improvement in their locomotor function compared with MPTP-injected mice given mutant peptide. More importantly, administration of NBD peptide 2 days after injection of MPTP showed substantial protection of TH $^+$ neurons, suggesting that NBD peptide can be used therapeutically to slow down or halt the progression of DA-neurodegeneration in MPTP-treated animals (Ghosh, Roy et al. 2007). In addition, infrared analysis of the brains of NBD-treated animals determined that significant levels of the NBD peptide localized within the brain tissue, suggesting that the NBD peptide could cross the BBB and reach sites of

neuroinflammation. The nature of the mechanism of NF- κ B inhibition within the SN remains to be determined, but these data suggest NF- κ B is a viable target for therapy for PD patients (Ghosh, Roy et al. 2007).

A second approach to inhibit inflammation has been to use small molecule inhibitors that specifically block function of IKK β . One such specific inhibitor, called Compound A, is a small molecule inhibitor of the kinase activity of IKK β but not IKK α . Compound A, also known as BAY-65-1942 (7-[2-(cyclopropylmethoxy)-6-hydroxyphenyl]-5-[(3S)-3-piperidinyl]-1,4-dihydro-2Hpyrido [2,3- d] [1,3]-oxazin-2-one hydrochloride), has been shown to specifically and effectively block the catalytic activity of IKK β , inhibiting its ability to phosphorylate I κ B and activate the cytosolic p50/p65 NF- κ B heterodimers (Moss, Stansfield et al. 2007; Zhang, Qian et al. 2010). In one recent study, Zhang and colleagues (2010) used Compound A in an LPS-induced neurodegeneration model to inhibit the canonical NF- κ B pathway and halt inflammation-induced DA-neurodegeneration. In this model, LPS was injected directly into one side of the midbrain of rats, leading to inflammation-induced degeneration of DA-neurons (Zhang, Qian et al. 2010). It was found that Compound A strongly inhibited the activation of NF- κ B *in vitro* and *in vivo*, as well as the mRNA expression and subsequent release of pro-inflammatory mediators. Compound A also significantly inhibited LPS- and MPTP-induced DA-neurotoxicity *in vitro*, and this neuroprotective activity required the presence of microglial cells. Most importantly, administration of Compound A to animals injected intranigally with LPS, attenuated LPS-induced DA neuronal loss and microglia activation within the SN ((Zhang, Qian et al. ; Zhang, Qian et al. 2010). These data provide strong evidence that NF- κ B offers a promising potential therapeutic target to halt DA-neurodegeneration, and that much additional work needs to be performed to determine the optimal approach and agent best suited for the treatment of PD.

5.2 β 2-AR agonists as PD therapeutics

A family of compounds that have recently been shown to potentially reduce inflammation and DA-neurodegeneration in animal models are the β 2 adrenergic receptor agonists. The β 2 adrenergic receptor (β 2AR) is a G-protein coupled receptor (GPCR) which is known to regulate of smooth muscle function in the airway and vasculature. Interestingly, β 2AR expression has also been identified on immune cells such as macrophages, microglia, T cells, and B cells, and signaling through this receptor can inhibit the inflammatory response of these cells (Koff, Fann et al. 1986; Severn, Rapson et al. 1992; van der Poll, Jansen et al. 1994; Sekut, Champion et al. 1995; Panina-Bordignon, Mazzeo et al. 1997; Farmer and Pugin 2000; Kin and Sanders 2006). Both short-acting and long-acting β 2AR agonists have been used for pharmacological studies and clinical therapy, and results have indicated that they possess the ability to inhibit the inflammatory responses by immune cells. Several of these long-acting agonists such as salmeterol (Advair[®]) and formoterol (Symbicort[®]) are currently being used as anti-inflammatory therapeutics to treat asthma and chronic obstructive pulmonary disease (COPD) (Koto, Mak et al. 1996; Tashkin and Cooper 2004; McKeage and Keam 2009). However, potential use of β 2AR agonists in neurodegenerative diseases in the CNS has not been well studied. Since most long-acting β 2AR agonists are highly lipophilic and should readily cross the BBB, it is likely that these compounds could have an immunomodulatory effect on the progression of inflammation in PD patients by inhibiting

the activation of microglia that normally express high levels of β 2AR (Tanaka, Kashima et al. 2002).

When long-acting β 2AR agonists were tested for DA-neuroprotective properties, it was found that the compounds can inhibit DA-neurodegeneration *in vitro*, even if used at extremely low doses. Furthermore, administration of the long-acting β 2AR agonist salmeterol significantly protects DA neurons against LPS- and MPTP-induced cytotoxicity *in vivo* (Qian, Wu et al. 2011). Mechanistic studies using primary midbrain neuron-glia cultures demonstrated that salmeterol, as well as several other long-acting β 2AR agonists, have potent neuroprotective effects through their inhibition of microglial inflammatory mediator production. These anti-inflammatory effects of salmeterol require the presence of β 2AR, are mediated through the inhibition of both MAPK and NF- κ B signaling pathways in activated microglia, and function independently of the canonical GPCR/cAMP/PKA signaling pathway. It was further determined that this inhibition is dependent on the expression of β -arrestin 2, which suggests a novel mechanism for the long-acting β 2AR agonists in regulating CNS inflammatory conditions (Qian, Wu et al. 2011). Therefore, the high specific activity and effectiveness of β 2AR agonists such as salmeterol at inhibiting inflammation and DA-neurodegeneration within the CNS in these animal models suggests they have potential for the treatment of chronic inflammatory disorders and in particular, Parkinson's disease.

6. Proposed model of neuroinflammation in PD

Inflammation associated with PD can be initiated in the brain by internal factors such as a brain injury, a genetic mutation or some other brain insult or dysfunction (Nagatsu and Sawada 2006; Tansey, McCoy et al. 2007; Hirsch and Hunot 2009; Qian, Flood et al. 2010)(Figure 1). These sorts of intracerebral inflammatory stimuli activate the microglia which then up-regulate production of inflammatory factors including inflammatory cytokines such as TNF α , IL-1 β or IL-6, as well as NO and ROS (Nagatsu, Mogi et al. 2000; Nagatsu and Sawada 2006; Tansey, McCoy et al. 2007). These in turn stimulate a further inflammatory response which results in a self-perpetuating chronic inflammatory condition (Qian and Flood 2008; Qian, Flood et al. 2010). Alternatively, inflammation in the periphery might induce the initial inflammatory response within the brain. Normally, the microvascular endothelial cells that line the blood vessels in the brain are tightly joined to each other through cell-cell tight junctions, and thus form the blood-brain-barrier which excludes most substances or cells from entry into the brain (Zlokovic 2008). A chronic peripheral inflammatory response can work to disrupt the cell-cell junctions and the blood-brain barrier, allowing access of inflammatory factors and cells into the brain (Stamatovic, Keep et al. 2008; Zlokovic 2008). These can then activate microglia and initiate the perpetual round of pro-inflammatory factor production leading to chronic neuroinflammation. Once microglia are activated, from whatever the source of initial inflammatory stimulus, the microglial response increases secretion of inflammatory cytokines and release of NO (Qian and Flood 2008). Microglial activation also precipitates an enhanced respiratory burst and release of ROS (Colton and Gilbert 1987). These various inflammatory mediators can trigger cell death in neurons (Chao, Hu et al. 1992; Colton and Gilbert 1993; Taylor, Jones et al. 2005), including DA neurons which seem to be especially vulnerable. Dying neurons then stimulate an intensified inflammatory response as the brain attempts to restore stasis (Qian, Flood et al. 2010). Therapies aimed at halting neurodegeneration are increasingly based on

intervention to top the chronic inflammatory response, including introduction of anti-inflammatory drugs, compounds, cytokines and Treg cells, which inherently release anti-inflammatory cytokines such as TGF β 1 and IL-10.

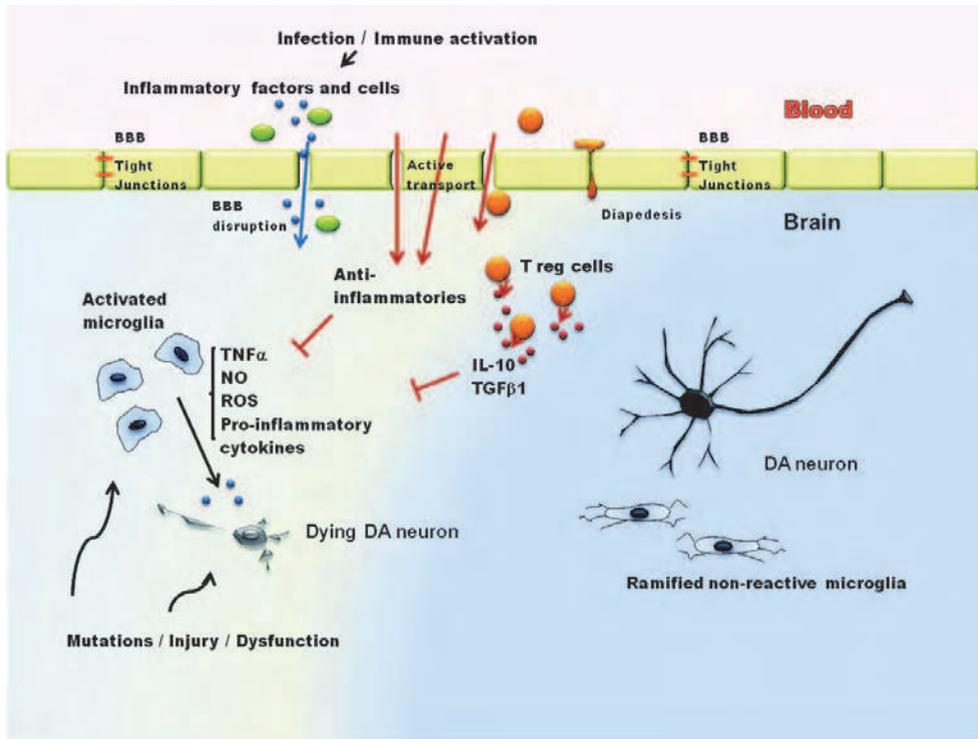


Fig. 1. Model relating chronic inflammation to dopaminergic (DA)-neuron death in Parkinson's Disease.

7. Future directions for PD therapy

Five relatively new approaches in anti-inflammatory therapy described above all show some promise for short-term or long-term therapeutics in PD. The use of exogenous anti-inflammatory cytokines such as IL10 or TGF β 1, has been shown to have potent effects in reducing neurotoxicity in both *in vitro* and in animal models of PD. However, although both cytokines are considered to be predominantly anti-inflammatory, they each can have pro-inflammatory effects in certain contexts. More thorough functional studies are needed for these anti-inflammatory cytokines, especially in the context of PD models before these might be readily used in human therapy. Particularly important factors include what form and what mode of delivery will optimize the anti-inflammatory effects of the cytokines as therapy and ameliorate any unintended negative effects. Cell-based therapies such as regulatory T cells (Tregs) offer great promise for long-term therapy in many degenerative disorders including PD. Both the use of T cells and stem cell transplantation focus on regeneration as well as intervening in neuronal death processes. However, some of the

potential problems with these cell-transplant strategies are obvious since these are invasive therapies and as such carry the attendant problems associated with transplants in general as well as the specific consequences of introducing these cells into PD patients. Much remains to be determined before they become readily applicable for PD therapy. Anti-inflammatory therapies have already been developed in animal models of PD and are being used to a limited extent in human studies. The finding that the chronic use of certain NSAIDs seems to lower the risk of PD is promising for possibly reducing PD incidence. Additionally, NSAIDs (or SAIDs) might be used profitably in combinatorial therapy for PD, but the long-term use of these drugs already has well documented potential health problems. Careful determination of the potential risks to PD patients as compared to what benefits may be gained from long-term use of NSAIDs or SAIDs needs to be addressed. This is particularly important since these drugs might be well suited to combinatorial therapy in conjunction with existing PD treatments. Similarly, morphinan-based drugs, or other drugs already used for treatment of other diseases such as beta2-adrenergic receptor agonists, hold the most immediate potential for development as therapeutics to treat PD. The normal pharmacokinetics and safety profiles have already been established for some of these drugs such as salmeterol, and although preclinical studies indicate potential efficacy in PD, again the specific pharmacokinetics and safety for treating the neuroinflammation underlying PD must be determined. Orally available NF- κ B inhibitors are equally attractive as PD therapies and are currently being studied in preclinical applications in a variety of diseases including PD.

8. References

- Abou-Sleiman, P. M., D. G. Healy, et al. (2003). "The role of pathogenic DJ-1 mutations in Parkinson's disease." *Ann Neurol* 54(3): 283-6.
- Akiyama, H. and P. L. McGeer (1989). "Microglial response to 6-hydroxydopamine-induced substantia nigra lesions." *Brain Res* 489(2): 247-53.
- Albin, R. L., A. B. Young, et al. (1989). "The functional anatomy of basal ganglia disorders." *Trends Neurosci* 12(10): 366-75.
- Angelov, D. N., S. Waibel, et al. (2003). "Therapeutic vaccine for acute and chronic motor neuron diseases: implications for amyotrophic lateral sclerosis." *Proc Natl Acad Sci U S A* 100(8): 4790-5.
- Appel, S. H., D. R. Beers, et al. (2009). "T cell-microglial dialogue in Parkinson's disease and amyotrophic lateral sclerosis: are we listening?" *Trends Immunol* 31(1): 7-17.
- Argaw, A. T., B. T. Gurfein, et al. (2009). "VEGF-mediated disruption of endothelial CLN-5 promotes blood-brain barrier breakdown." *Proc Natl Acad Sci U S A* 106(6): 1977-82.
- Argaw, A. T., Y. Zhang, et al. (2006). "IL-1 β regulates blood-brain barrier permeability via reactivation of the hypoxia-angiogenesis program." *J Immunol* 177(8): 5574-84.
- Atreya, I., R. Atreya, et al. (2008). "NF- κ B in inflammatory bowel disease." *J Intern Med* 263(6): 591-6.
- Barcia, C., V. Bautista, et al. (2005). "Changes in vascularization in substantia nigra pars compacta of monkeys rendered parkinsonian." *J Neural Transm* 112(9): 1237-48.

- Barcia, C., V. de Pablos, et al. (2005). "Increased plasma levels of TNF-alpha but not of IL1-beta in MPTP-treated monkeys one year after the MPTP administration." *Parkinsonism Relat Disord* 11(7): 435-9.
- Barcia, C., A. Sanchez Bahillo, et al. (2004). "Evidence of active microglia in substantia nigra pars compacta of parkinsonian monkeys 1 year after MPTP exposure." *Glia* 46(4): 402-9.
- Bartels, A. L. and K. L. Leenders (2007). "Neuroinflammation in the pathophysiology of Parkinson's disease: evidence from animal models to human in vivo studies with [11C]-PK11195 PET." *Mov Disord* 22(13): 1852-6.
- Bas, J., M. Calopa, et al. (2001). "Lymphocyte populations in Parkinson's disease and in rat models of parkinsonism." *J Neuroimmunol* 113(1): 146-52.
- Benner, E. J., R. L. Mosley, et al. (2004). "Therapeutic immunization protects dopaminergic neurons in a mouse model of Parkinson's disease." *Proc Natl Acad Sci U S A* 101(25): 9435-40.
- Bernardo, A., M. A. Ajmone-Cat, et al. (2005). "Nuclear receptor peroxisome proliferator-activated receptor-gamma is activated in rat microglial cells by the anti-inflammatory drug HCT1026, a derivative of flurbiprofen." *J Neurochem* 92(4): 895-903.
- Block, M. L. and J. S. Hong (2007). "Chronic microglial activation and progressive dopaminergic neurotoxicity." *Biochem Soc Trans* 35(Pt 5): 1127-32.
- Boka, G., P. Anglade, et al. (1994). "Immunocytochemical analysis of tumor necrosis factor and its receptors in Parkinson's disease." *Neurosci Lett* 172(1-2): 151-4.
- Bonifati, V., Y. H. Wu-Chou, et al. (2008). "LRRK2 mutation analysis in Parkinson disease families with evidence of linkage to PARK8." *Neurology* 70(24): 2348; author reply 2348-9.
- Bonuccelli, U., P. Del Dotto, et al. (1992). "Dextromethorphan and parkinsonism." *Lancet* 340(8810): 53.
- Carrithers, M. D., I. Visintin, et al. (2000). "Differential adhesion molecule requirements for immune surveillance and inflammatory recruitment." *Brain* 123 (Pt 6): 1092-101.
- Castano, A., A. J. Herrera, et al. (2002). "The degenerative effect of a single intranigral injection of LPS on the dopaminergic system is prevented by dexamethasone, and not mimicked by rh-TNF-alpha, IL-1beta and IFN-gamma." *J Neurochem* 81(1): 150-7.
- Chao, C. C., S. Hu, et al. (1992). "Activated microglia mediate neuronal cell injury via a nitric oxide mechanism." *J Immunol* 149(8): 2736-41.
- Chao, Y. X., B. P. He, et al. (2009). "Mesenchymal stem cell transplantation attenuates blood brain barrier damage and neuroinflammation and protects dopaminergic neurons against MPTP toxicity in the substantia nigra in a model of Parkinson's disease." *J Neuroimmunol* 216(1-2): 39-50.
- Chen, C. C. and A. M. Manning (1995). "Transcriptional regulation of endothelial cell adhesion molecules: a dominant role for NF-kappa B." *Agents Actions Suppl* 47: 135-41.

- Chen, H. Q., Z. Y. Jin, et al. (2008). "Luteolin protects dopaminergic neurons from inflammation-induced injury through inhibition of microglial activation." *Neurosci Lett* 448(2): 175-9.
- Chen, W. and S. M. Wahl (2003). "TGF-beta: the missing link in CD4+CD25+ regulatory T cell-mediated immunosuppression." *Cytokine Growth Factor Rev* 14(2): 85-9.
- Choi, D. W. (1987). "Dextrorphan and dextromethorphan attenuate glutamate neurotoxicity." *Brain Res* 403(2): 333-6.
- Choi, D. W., M. Maulucci-Gedde, et al. (1987). "Glutamate neurotoxicity in cortical cell culture." *J Neurosci* 7(2): 357-68.
- Choi, D. W., S. Peters, et al. (1987). "Dextrorphan and levorphanol selectively block N-methyl-D-aspartate receptor-mediated neurotoxicity on cortical neurons." *J Pharmacol Exp Ther* 242(2): 713-20.
- Church, J., D. Lodge, et al. (1985). "Differential effects of dextrorphan and levorphanol on the excitation of rat spinal neurons by amino acids." *Eur J Pharmacol* 111(2): 185-90.
- Cicchetti, F., J. Drouin-Ouellet, et al. (2009). "Environmental toxins and Parkinson's disease: what have we learned from pesticide-induced animal models?" *Trends Pharmacol Sci* 30(9): 475-83.
- Cicchetti, F., N. Lapointe, et al. (2005). "Systemic exposure to paraquat and maneb models early Parkinson's disease in young adult rats." *Neurobiol Dis* 20(2): 360-71.
- Colton, C. A. and D. L. Gilbert (1987). "Production of superoxide anions by a CNS macrophage, the microglia." *FEBS Lett* 223(2): 284-8.
- Colton, C. A. and D. L. Gilbert (1993). "Microglia, an in vivo source of reactive oxygen species in the brain." *Adv Neurol* 59: 321-6.
- Coxon, A., P. Rieu, et al. (1996). "A novel role for the beta 2 integrin CD11b/CD18 in neutrophil apoptosis: a homeostatic mechanism in inflammation." *Immunity* 5(6): 653-66.
- Criswell, L. A. (2010). "Gene discovery in rheumatoid arthritis highlights the CD40/NF-kappaB signaling pathway in disease pathogenesis." *Immunol Rev* 233(1): 55-61.
- Dauer, W. and S. Przedborski (2003). "Parkinson's disease: mechanisms and models." *Neuron* 39(6): 889-909.
- Delgado, M., A. Chorny, et al. (2005). "Vasoactive intestinal peptide generates CD4+CD25+ regulatory T cells in vivo." *J Leukoc Biol* 78(6): 1327-38.
- Dendorfer, U. (1996). "Molecular biology of cytokines." *Artif Organs* 20(5): 437-44.
- Dickson, D. W. (2007). "Linking selective vulnerability to cell death mechanisms in Parkinson's disease." *Am J Pathol* 170(1): 16-9.
- Dobbs, R. J., A. Charlett, et al. (1999). "Association of circulating TNF-alpha and IL-6 with ageing and parkinsonism." *Acta Neurol Scand* 100(1): 34-41.
- Du, Y., Z. Ma, et al. (2001). "Minocycline prevents nigrostriatal dopaminergic neurodegeneration in the MPTP model of Parkinson's disease." *Proc Natl Acad Sci U S A* 98(25): 14669-74.
- Engelhardt, B. and R. M. Ransohoff (2005). "The ins and outs of T-lymphocyte trafficking to the CNS: anatomical sites and molecular mechanisms." *Trends Immunol* 26(9): 485-95.

- Fahn, S. (2003). "Description of Parkinson's disease as a clinical syndrome." *Ann N Y Acad Sci* 991: 1-14.
- Farmer, P. and J. Pugin (2000). "beta-adrenergic agonists exert their "anti-inflammatory" effects in monocytic cells through the IkappaB/NF-kappaB pathway." *Am J Physiol Lung Cell Mol Physiol* 279(4): L675-82.
- Farrer, M. J., K. Haugarvoll, et al. (2006). "Genomewide association, Parkinson disease, and PARK10." *Am J Hum Genet* 78(6): 1084-8; author reply 1092-4.
- Faucheux, B. A., A. M. Bonnet, et al. (1999). "Blood vessels change in the mesencephalon of patients with Parkinson's disease." *Lancet* 353(9157): 981-2.
- Fernandez-Checa, J. C., N. Kaplowitz, et al. (1997). "GSH transport in mitochondria: defense against TNF-induced oxidative stress and alcohol-induced defect." *Am J Physiol* 273(1 Pt 1): G7-17.
- Fernandez-Martin, A., E. Gonzalez-Rey, et al. (2006). "Vasoactive intestinal peptide induces regulatory T cells during experimental autoimmune encephalomyelitis." *Eur J Immunol* 36(2): 318-26.
- Flood, P. M., L. Qian, et al. (2011). "Transcriptional Factor NF-kB as a Target for Therapy in Parkinson's Disease." *Inflammation and Parkinson's Disease*, SAGE-Hindawi, pub. in press.
- Floyd, R. A. (1999). "Antioxidants, oxidative stress, and degenerative neurological disorders." *Proc Soc Exp Biol Med* 222(3): 236-45.
- Floyd, R. A. (1999). "Neuroinflammatory processes are important in neurodegenerative diseases: an hypothesis to explain the increased formation of reactive oxygen and nitrogen species as major factors involved in neurodegenerative disease development." *Free Radic Biol Med* 26(9-10): 1346-55.
- Gao, H. M. and J. S. Hong (2008). "Why neurodegenerative diseases are progressive: uncontrolled inflammation drives disease progression." *Trends Immunol* 29(8): 357-65.
- Gao, H. M., J. Jiang, et al. (2002). "Microglial activation-mediated delayed and progressive degeneration of rat nigral dopaminergic neurons: relevance to Parkinson's disease." *J Neurochem* 81(6): 1285-97.
- Gao, H. M., B. Liu, et al. (2003). "Critical role for microglial NADPH oxidase in rotenone-induced degeneration of dopaminergic neurons." *J Neurosci* 23(15): 6181-7.
- Gao, J., H. Miao, et al. (2011). "Influence of aging on the dopaminergic neurons in the substantia nigra pars compacta of rats." *Curr Aging Sci* 4(1): 19-24.
- Gao, X., H. Chen, et al. (2011). "Use of ibuprofen and risk of Parkinson disease." *Neurology* 76(10): 863-9.
- Gauss, K. A., L. K. Nelson-Overton, et al. (2007). "Role of NF-kappaB in transcriptional regulation of the phagocyte NADPH oxidase by tumor necrosis factor-alpha." *J Leukoc Biol* 82(3): 729-41.
- Ghosh, A., A. Roy, et al. (2007). "Selective inhibition of NF-kappaB activation prevents dopaminergic neuronal loss in a mouse model of Parkinson's disease." *Proc Natl Acad Sci U S A* 104(47): 18754-9.

- Godbout, J. P. and R. W. Johnson (2009). "Age and Neuroinflammation: A Lifetime of Psychoneuroimmune Consequences." *Immunology and Allergy Clinics of North America* 29(2): 321-337.
- Goldberg, M. P., J. H. Weiss, et al. (1987). "N-methyl-D-aspartate receptors mediate hypoxic neuronal injury in cortical culture." *J Pharmacol Exp Ther* 243(2): 784-91.
- Greenamyre, J. T. and C. F. O'Brien (1991). "N-methyl-D-aspartate antagonists in the treatment of Parkinson's disease." *Arch Neurol* 48(9): 977-81.
- Groemping, Y. and K. Rittinger (2005). "Activation and assembly of the NADPH oxidase: a structural perspective." *Biochem J* 386(Pt 3): 401-16.
- Gutierrez, E. G., W. A. Banks, et al. (1993). "Murine tumor necrosis factor alpha is transported from blood to brain in the mouse." *J Neuroimmunol* 47(2): 169-76.
- Halliwel, B. (2006). "Oxidative stress and neurodegeneration: where are we now?" *J Neurochem* 97(6): 1634-58.
- Henrich-Noack, P., J. H. Prehn, et al. (1996). "TGF-beta 1 protects hippocampal neurons against degeneration caused by transient global ischemia. Dose-response relationship and potential neuroprotective mechanisms." *Stroke* 27(9): 1609-14; discussion 1615.
- Herrera, A. J., M. Tomas-Camardiel, et al. (2005). "Inflammatory process as a determinant factor for the degeneration of substantia nigra dopaminergic neurons." *J Neural Transm* 112(1): 111-9.
- Hirsch, E. C. (2000). "Nigrostriatal system plasticity in Parkinson's disease: effect of dopaminergic denervation and treatment." *Ann Neurol* 47(4 Suppl 1): S115-20; discussion S120-1.
- Hirsch, E. C. and S. Hunot (2009). "Neuroinflammation in Parkinson's disease: a target for neuroprotection?" *Lancet Neurol* 8(4): 382-97.
- Hisanaga, K., M. Asagi, et al. (2001). "Increase in peripheral CD4 bright+ CD8 dull+ T cells in Parkinson disease." *Arch Neurol* 58(10): 1580-3.
- Hu, X., D. Zhang, et al. (2008). "Macrophage antigen complex-1 mediates reactive microgliosis and progressive dopaminergic neurodegeneration in the MPTP model of Parkinson's disease." *J Immunol* 181(10): 7194-204.
- Huusko, J., M. Merentie, et al. (2009). "The effects of VEGF-R1 and VEGF-R2 ligands on angiogenic responses and left ventricular function in mice." *Cardiovasc Res*.
- Huxford, T. and G. Ghosh (2009). "A structural guide to proteins of the NF-kappaB signaling module." *Cold Spring Harb Perspect Biol* 1(3): a000075.
- Imamura, K., N. Hishikawa, et al. (2003). "Distribution of major histocompatibility complex 1-positive microglia and cytokine profile of Parkinson's disease brains." *Acta Neuropathol* 106: 518-526.
- Jackson-Lewis, V. and S. Przedborski (2007). "Protocol for the MPTP mouse model of Parkinson's disease." *Nat Protoc* 2(1): 141-51.
- Jackson-Lewis, V. and R. J. Smeyne (2005). "MPTP and SNpc DA neuronal vulnerability: role of dopamine, superoxide and nitric oxide in neurotoxicity. Minireview." *Neurotox Res* 7(3): 193-202.
- Jellinger, K. A. (2001). "The pathology of Parkinson's disease." *Adv Neurol* 86: 55-72.

- Jenner, P. and C. W. Olanow (1996). "Oxidative stress and the pathogenesis of Parkinson's disease." *Neurology* 47(6 Suppl 3): S161-70.
- Jenner, P. and C. W. Olanow (1998). "Understanding cell death in Parkinson's disease." *Ann Neurol* 44(3 Suppl 1): S72-84.
- Jenner, P. and C. W. Olanow (2006). "The pathogenesis of cell death in Parkinson's disease." *Neurology* 66(10 Suppl 4): S24-36.
- Ji, S., G. Kronenberg, et al. (2009). "Acute neuroprotection by pioglitazone after mild brain ischemia without effect on long-term outcome." *Exp Neurol* 216(2): 321-8.
- Jiang, H., Y. C. Wu, et al. (2007). "Parkinson's disease genetic mutations increase cell susceptibility to stress: mutant alpha-synuclein enhances H₂O₂- and Sin-1-induced cell death." *Neurobiol Aging* 28(11): 1709-17.
- Johnston, L. C., X. Su, et al. (2008). "Human interleukin-10 gene transfer is protective in a rat model of Parkinson's disease." *Mol Ther* 16(8): 1392-9.
- Kang, S. S., J. Y. Lee, et al. (2004). "Neuroprotective effects of flavones on hydrogen peroxide-induced apoptosis in SH-SY5Y neuroblastoma cells." *Bioorg Med Chem Lett* 14(9): 2261-4.
- Keeney, P. M., J. Xie, et al. (2006). "Parkinson's disease brain mitochondrial complex I has oxidatively damaged subunits and is functionally impaired and misassembled." *J Neurosci* 26(19): 5256-64.
- Kim, W. G., R. P. Mohny, et al. (2000). "Regional difference in susceptibility to lipopolysaccharide-induced neurotoxicity in the rat brain: role of microglia." *J Neurosci* 20(16): 6309-16.
- Kin, N. W. and V. M. Sanders (2006). "It takes nerve to tell T and B cells what to do." *J Leukoc Biol* 79(6): 1093-104.
- Kipnis, J. and M. Schwartz (2002). "Dual action of glatiramer acetate (Cop-1) in the treatment of CNS autoimmune and neurodegenerative disorders." *Trends Mol Med* 8(7): 319-23.
- Kirk, S. L. and S. J. Karlik (2003). "VEGF and vascular changes in chronic neuroinflammation." *Journal of Autoimmunity* 21(4): 353-363.
- Klockgether, T. (2004). "Parkinson's disease: clinical aspects." *Cell Tissue Res* 318(1): 115-20.
- Knott, C., G. Stern, et al. (2000). "Inflammatory regulators in Parkinson's disease: iNOS, lipocortin-1, and cyclooxygenases-1 and -2." *Mol Cell Neurosci* 16(6): 724-39.
- Koff, W. C., A. V. Fann, et al. (1986). "Catecholamine-induced suppression of interleukin-1 production." *Lymphokine Res* 5(4): 239-47.
- Koh, J. Y. and D. W. Choi (1987). "Effect of anticonvulsant drugs on glutamate neurotoxicity in cortical cell culture." *Neurology* 37(2): 319-22.
- Kohutnicka, M., E. Lewandowska, et al. (1998). "Microglial and astrocytic involvement in a murine model of Parkinson's disease induced by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)." *Immunopharmacology* 39(3): 167-180.
- Kortekaas, R., K. L. Leenders, et al. (2005). "Blood-brain barrier dysfunction in parkinsonian midbrain in vivo." *Ann Neurol* 57(2): 176-9.
- Koto, H., J. C. Mak, et al. (1996). "Mechanisms of impaired beta-adrenoceptor-induced airway relaxation by interleukin-1beta in vivo in the rat." *J Clin Invest* 98(8): 1780-7.

- Krige, D., M. T. Carroll, et al. (1992). "Platelet mitochondrial function in Parkinson's disease. The Royal Kings and Queens Parkinson Disease Research Group." *Ann Neurol* 32(6): 782-8.
- Kurkowska-Jastrzebska, I., T. Litwin, et al. (2004). "Dexamethasone protects against dopaminergic neurons damage in a mouse model of Parkinson's disease." *Int Immunopharmacol* 4(10-11): 1307-18.
- Kurkowska-Jastrzebska, I., A. Wronska, et al. (1999). "The inflammatory reaction following 1-methyl-4-phenyl-1,2,3, 6-tetrahydropyridine intoxication in mouse." *Exp Neurol* 156(1): 50-61.
- Langston, J. W., L. S. Forno, et al. (1999). "Evidence of active nerve cell degeneration in the substantia nigra of humans years after 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine exposure." *Ann Neurol* 46(4): 598-605.
- Lappas, M., M. Permezel, et al. (2002). "Nuclear factor kappa B regulation of proinflammatory cytokines in human gestational tissues in vitro." *Biol Reprod* 67(2): 668-73.
- Latanich, C. A. and L. H. Toledo-Pereyra (2009). "Searching for NF-kappaB-based treatments of ischemia reperfusion injury." *J Invest Surg* 22(4): 301-15.
- Lawrence, T. (2009). "The nuclear factor NF-kappaB pathway in inflammation." *Cold Spring Harb Perspect Biol* 1(6): a001651.
- Lawrence, T. and C. Fong "The resolution of inflammation: anti-inflammatory roles for NF-kappaB." *Int J Biochem Cell Biol* 42(4): 519-23.
- Lee, M., T. Cho, et al. (2010). "Depletion of GSH in glial cells induces neurotoxicity: relevance to aging and degenerative neurological diseases." *The FASEB Journal* 24(7): 2533-2545.
- Lehnardt, S., L. Massillon, et al. (2003). "Activation of innate immunity in the CNS triggers neurodegeneration through a Toll-like receptor 4-dependent pathway." *Proc Natl Acad Sci U S A* 100(14): 8514-9.
- Lin, M. T. and M. F. Beal (2006). "Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases." *Nature* 443(7113): 787-95.
- Litteljohn, D., E. Mangano, et al. (2011). "Inflammatory mechanisms of neurodegeneration in toxin-based models of Parkinson's disease." *Parkinsons Dis* 2011: 713517.
- Liu, B. and J. S. Hong (2003). "Neuroprotective effect of naloxone in inflammation-mediated dopaminergic neurodegeneration. Dissociation from the involvement of opioid receptors." *Methods Mol Med* 79: 43-54.
- Liu, J., N. Gong, et al. (2009). "Neuromodulatory activities of CD4+CD25+ regulatory T cells in a murine model of HIV-1-associated neurodegeneration." *J Immunol* 182(6): 3855-65.
- Liu, L., E. Buchner, et al. (1996). "Amelioration of rat experimental arthritides by treatment with the alkaloid sinomenine." *Int J Immunopharmacol* 18(10): 529-43.
- Liu, L., K. Resch, et al. (1994). "Inhibition of lymphocyte proliferation by the anti-arthritic drug sinomenine." *Int J Immunopharmacol* 16(8): 685-91.
- Liu, Y., L. Qin, et al. (2003). "Dextromethorphan protects dopaminergic neurons against inflammation-mediated degeneration through inhibition of microglial activation." *J Pharmacol Exp Ther* 305(1): 212-8.

- Loeffler, D. A., D. M. Camp, et al. (2006). "Complement activation in the Parkinson's disease substantia nigra: an immunocytochemical study." *J Neuroinflammation* 3: 29.
- Lotan, M. and M. Schwartz (1994). "Cross-talk between the immune system and the nervous system in response to injury: implications for regeneration." *FASEB J* 8(13): 1026-1033.
- Lucking, C. B., A. Durr, et al. (2000). "Association between early-onset Parkinson's disease and mutations in the parkin gene." *N Engl J Med* 342(21): 1560-7.
- Marshall, K. A., M. Reist, et al. (1999). "The neuronal toxicity of sulfite plus peroxynitrite is enhanced by glutathione depletion: implications for Parkinson's disease." *Free Radic Biol Med* 27(5-6): 515-20.
- Matarese, G., V. De Rosa, et al. (2008). "Regulatory CD4 T cells: sensing the environment." *Trends Immunol* 29(1): 12-7.
- May, M. J., R. B. Marienfeld, et al. (2002). "Characterization of the Ikappa B-kinase NEMO binding domain." *J Biol Chem* 277(48): 45992-6000.
- McGeachy, M. J., K. S. Bak-Jensen, et al. (2007). "TGF-beta and IL-6 drive the production of IL-17 and IL-10 by T cells and restrain T(H)-17 cell-mediated pathology." *Nat Immunol* 8(12): 1390-7.
- McGeer, P. L., S. Itagaki, et al. (1988). "Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains." *Neurology* 38(8): 1285-91.
- McGeer, P. L. and E. G. McGeer (2004). "Inflammation and the degenerative diseases of aging." *Ann N Y Acad Sci* 1035: 104-16.
- McGeer, P. L., C. Schwab, et al. (2003). "Presence of reactive microglia in monkey substantia nigra years after 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine administration." *Ann Neurol* 54(5): 599-604.
- McGeer, P. L., K. Yasojima, et al. (2001). "Inflammation in Parkinson's disease." *Adv Neurol* 86: 83-9.
- McKeage, K. and S. J. Keam (2009). "Salmeterol/fluticasone propionate: a review of its use in asthma." *Drugs* 69(13): 1799-828.
- Mizuno, Y., N. Hattori, et al. (2001). "Familial Parkinson's disease. Alpha-synuclein and parkin." *Adv Neurol* 86: 13-21.
- Mogi, M., M. Harada, et al. (1994). "Interleukin-1 beta, interleukin-6, epidermal growth factor and transforming growth factor-alpha are elevated in the brain from parkinsonian patients." *Neurosci Lett* 180(2): 147-50.
- Mogi, M., M. Harada, et al. (1996). "Interleukin (IL)-1 beta, IL-2, IL-4, IL-6 and transforming growth factor-alpha levels are elevated in ventricular cerebrospinal fluid in juvenile parkinsonism and Parkinson's disease." *Neurosci Lett* 211(1): 13-6.
- Mogi, M., M. Harada, et al. (1994). "Tumor necrosis factor-alpha (TNF-alpha) increases both in the brain and in the cerebrospinal fluid from parkinsonian patients." *Neurosci Lett* 165(1-2): 208-10.
- Monahan, A. J., M. Warren, et al. (2008). "Neuroinflammation and peripheral immune infiltration in Parkinson's disease: an autoimmune hypothesis." *Cell Transplant* 17(4): 363-72.

- Montastruc, J. L., N. Fabre, et al. (1994). "N-methyl-D-aspartate (NMDA) antagonist and Parkinson's disease: a pilot study with dextromethorphan." *Mov Disord* 9(2): 242-3.
- Moore, P. E., T. Lahiri, et al. (2001). "Selected contribution: synergism between TNF-alpha and IL-1 beta in airway smooth muscle cells: implications for beta-adrenergic responsiveness." *J Appl Physiol* 91(3): 1467-74.
- Mosley, R. L., E. J. Benner, et al. (2006). "Neuroinflammation, Oxidative Stress and the Pathogenesis of Parkinson's Disease." *Clin Neurosci Res* 6(5): 261-281.
- Moss, N. C., W. E. Stansfield, et al. (2007). "IKKbeta inhibition attenuates myocardial injury and dysfunction following acute ischemia-reperfusion injury." *Am J Physiol Heart Circ Physiol* 293(4): H2248-2253.
- Nagatsu, T., M. Mogi, et al. (2000). "Changes in cytokines and neurotrophins in Parkinson's disease." *J Neural Transm Suppl*(60): 277-90.
- Nagatsu, T., M. Mogi, et al. (2000). "Cytokines in Parkinson's disease." *J Neural Transm Suppl*(58): 143-51.
- Nagatsu, T. and M. Sawada (2006). "Cellular and molecular mechanisms of Parkinson's disease: neurotoxins, causative genes, and inflammatory cytokines." *Cell Mol Neurobiol* 26(4-6): 781-802.
- Naik, E. and V. M. Dixit (2011). "Mitochondrial reactive oxygen species drive proinflammatory cytokine production." *The Journal of Experimental Medicine* 208(3): 417-420.
- Natarajan, C. and J. J. Bright (2002). "Curcumin inhibits experimental allergic encephalomyelitis by blocking IL-12 signaling through Janus kinase-STAT pathway in T lymphocytes." *J Immunol* 168(12): 6506-13.
- Negi, G., A. Kumar, et al. (2011). "Melatonin modulates neuroinflammation and oxidative stress in experimental diabetic neuropathy: effects on NF- κ B and Nrf2 cascades." *Journal of Pineal Research* 50(2): 124-131.
- Neuwelt, E. A. and W. K. Clark (1978). "The immune system and the nervous system." *Neurosurgery* 3(3): 419-430.
- Nguyen, M. D., T. D'Aigle, et al. (2004). "Exacerbation of motor neuron disease by chronic stimulation of innate immunity in a mouse model of amyotrophic lateral sclerosis." *J Neurosci* 24(6): 1340-9.
- Nguyen, M. D., J. P. Julien, et al. (2002). "Innate immunity: the missing link in neuroprotection and neurodegeneration?" *Nat Rev Neurosci* 3(3): 216-27.
- Nguyen, Q. D., S. Tatlipinar, et al. (2006). "Vascular endothelial growth factor is a critical stimulus for diabetic macular edema." *Am J Ophthalmol* 142(6): 961-9.
- Niehaus, I. and J. H. Lange (2003). "Endotoxin: is it an environmental factor in the cause of Parkinson's disease?" *Occup Environ Med* 60(5): 378.
- Oeckinghaus, A. and S. Ghosh (2009). "The NF-kappaB family of transcription factors and its regulation." *Cold Spring Harb Perspect Biol* 1(4): a000034.
- Orr, C. F., D. B. Rowe, et al. (2002). "An inflammatory review of Parkinson's disease." *Prog Neurobiol* 68(5): 325-40.
- Orr, C. F., D. B. Rowe, et al. (2005). "A possible role for humoral immunity in the pathogenesis of Parkinson's disease." *Brain* 128(Pt 11): 2665-74.

- Owen, A. D., A. H. Schapira, et al. (1996). "Oxidative stress and Parkinson's disease." *Ann N Y Acad Sci* 786: 217-23.
- Pachter, J. S., H. E. de Vries, et al. (2003). "The blood-brain barrier and its role in immune privilege in the central nervous system." *J Neuropathol Exp Neurol* 62(6): 593-604.
- Pan, W. and A. J. Kastin (2002). "TNF[alpha] Transport across the Blood-Brain Barrier Is Abolished in Receptor Knockout Mice." *Experimental Neurology* 174(2): 193-200.
- Panina-Bordignon, P., D. Mazzeo, et al. (1997). "Beta2-agonists prevent Th1 development by selective inhibition of interleukin 12." *J Clin Invest* 100(6): 1513-9.
- Park, K. M. and W. J. Bowers (2010). "Tumor necrosis factor-alpha mediated signaling in neuronal homeostasis and dysfunction." *Cell Signal* 22(7): 977-83.
- Pearce, R. K., A. Owen, et al. (1997). "Alterations in the distribution of glutathione in the substantia nigra in Parkinson's disease." *J Neural Transm* 104(6-7): 661-77.
- Pei, Z., H. Pang, et al. (2007). "MAC1 mediates LPS-induced production of superoxide by microglia: the role of pattern recognition receptors in dopaminergic neurotoxicity." *Glia* 55(13): 1362-73.
- Pereira, S. G. and F. Oakley (2008). "Nuclear factor-kappaB1: regulation and function." *Int J Biochem Cell Biol* 40(8): 1425-30.
- Pisani, V., V. Moschella, et al. (2010). "Dynamic changes of anandamide in the cerebrospinal fluid of Parkinson's disease patients." *Movement Disorders* 25(7): 920-924.
- Polymeropoulos, M. H., C. Lavedan, et al. (1997). "Mutation in the alpha-synuclein gene identified in families with Parkinson's disease." *Science* 276(5321): 2045-7.
- Pope, L. E., M. H. Khalil, et al. (2004). "Pharmacokinetics of dextromethorphan after single or multiple dosing in combination with quinidine in extensive and poor metabolizers." *J Clin Pharmacol* 44(10): 1132-42.
- Prehn, J. H. and J. Krieglstein (1994). "Opposing effects of transforming growth factor-beta 1 on glutamate neurotoxicity." *Neuroscience* 60(1): 7-10.
- Qian, L., M. L. Block, et al. (2006). "Interleukin-10 protects lipopolysaccharide-induced neurotoxicity in primary midbrain cultures by inhibiting the function of NADPH oxidase." *J Pharmacol Exp Ther* 319(1): 44-52.
- Qian, L. and P. M. Flood (2008). "Microglial cells and Parkinson's disease." *Immunol Res* 41(3): 155-64.
- Qian, L., P. M. Flood, et al. (2010). "Neuroinflammation is a key player in Parkinson's disease and a prime target for therapy." *J Neural Transm* 117(8): 971-9.
- Qian, L., X. Gao, et al. (2007). "NADPH oxidase inhibitor DPI is neuroprotective at femtomolar concentrations through inhibition of microglia over-activation." *Parkinsonism Relat Disord* 13 Suppl 3: S316-20.
- Qian, L., K. S. Tan, et al. (2007). "Microglia-mediated neurotoxicity is inhibited by morphine through an opioid receptor-independent reduction of NADPH oxidase activity." *J Immunol* 179(2): 1198-209.
- Qian, L., S. J. Wei, et al. (2008). "Potent anti-inflammatory and neuroprotective effects of TGF-beta1 are mediated through the inhibition of ERK and p47phox-Ser345 phosphorylation and translocation in microglia." *J Immunol* 181(1): 660-8.

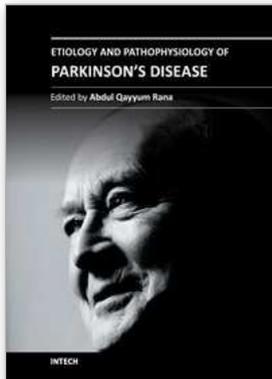
- Qian, L., H. M. Wu, et al. (2011). " β 2-Adrenergic Receptor Activation Prevents Rodent Dopaminergic Neurotoxicity by Inhibiting Microglia via a Novel Signaling Pathway." *J Immunol.* 186: 4443-4454.
- Qian, L., Z. Xu, et al. (2007). "Sinomenine, a natural dextrorotatory morphinan analog, is anti-inflammatory and neuroprotective through inhibition of microglial NADPH oxidase." *J Neuroinflammation* 4: 23.
- Qin, L., Y. Liu, et al. (2004). "NADPH oxidase mediates lipopolysaccharide-induced neurotoxicity and proinflammatory gene expression in activated microglia." *J Biol Chem* 279(2): 1415-21.
- Qin, L., X. Wu, et al. (2007). "Systemic LPS causes chronic neuroinflammation and progressive neurodegeneration." *Glia* 55(5): 453-62.
- Qureshi, G. A., S. Baig, et al. (1995). "Increased cerebrospinal fluid concentration of nitrite in Parkinson's disease." *Neuroreport* 6(12): 1642-4.
- Ransohoff, R. M. and V. H. Perry (2009). "Microglial physiology: unique stimuli, specialized responses." *Annu Rev Immunol* 27: 119-45.
- Reale, M., N. H. Greig, et al. (2009). "Peripheral chemo-cytokine profiles in Alzheimer's and Parkinson's diseases." *Mini Rev Med Chem* 9(10): 1229-41.
- Reuther-Madrid, J. Y., D. Kashatus, et al. (2002). "The p65/RelA subunit of NF-kappaB suppresses the sustained, antiapoptotic activity of Jun kinase induced by tumor necrosis factor." *Mol Cell Biol* 22(23): 8175-83.
- Reynolds, A. D., R. Banerjee, et al. (2007). "Neuroprotective activities of CD4+CD25+ regulatory T cells in an animal model of Parkinson's disease." *J Leukoc Biol* 82(5): 1083-94.
- Reynolds, A. D., D. K. Stone, et al. (2010). "Regulatory T cells attenuate Th17 cell-mediated nigrostriatal dopaminergic neurodegeneration in a model of Parkinson's disease." *J Immunol* 184(5): 2261-71.
- Reynolds, A. D., D. K. Stone, et al. (2009). "Nitrated α -synuclein-induced alterations in microglial immunity are regulated by CD4+ T cell subsets." *J Immunol* 182(7): 4137-49.
- Rite, I., A. Machado, et al. (2007). "Blood-brain barrier disruption induces in vivo degeneration of nigral dopaminergic neurons." *J Neurochem* 101(6): 1567-82.
- Roebuck, K. A. (1999). "Regulation of interleukin-8 gene expression." *J Interferon Cytokine Res* 19(5): 429-38.
- Roebuck, K. A., L. R. Carpenter, et al. (1999). "Stimulus-specific regulation of chemokine expression involves differential activation of the redox-responsive transcription factors AP-1 and NF-kappaB." *J Leukoc Biol* 65(3): 291-8.
- Rudolph, D., W. C. Yeh, et al. (2000). "Severe liver degeneration and lack of NF-kappaB activation in NEMO/IKKgamma-deficient mice." *Genes Dev* 14(7): 854-62.
- Ruocco, A., O. Nicole, et al. (1999). "A transforming growth factor-beta antagonist unmasks the neuroprotective role of this endogenous cytokine in excitotoxic and ischemic brain injury." *J Cereb Blood Flow Metab* 19(12): 1345-53.
- Ryu, J. K. and J. G. McLarnon (2008). "VEGF receptor antagonist Cyclo-VEGI reduces inflammatory reactivity and vascular leakiness and is neuroprotective against acute excitotoxic striatal insult." *J Neuroinflammation* 5: 18.

- Sairam, K., K. S. Saravanan, et al. (2003). "Non-steroidal anti-inflammatory drug sodium salicylate, but not diclofenac or celecoxib, protects against 1-methyl-4-phenyl pyridinium-induced dopaminergic neurotoxicity in rats." *Brain Res* 966(2): 245-52.
- Sawada, M., K. Imamura, et al. (2006). "Role of cytokines in inflammatory process in Parkinson's disease." *J Neural Transm Suppl*(70): 373-81.
- Schapira, A. H. (1994). "Mitochondrial function and neurotoxicity." *Curr Opin Neurol* 7(6): 531-4.
- Schapira, A. H. (2006). "Etiology of Parkinson's disease." *Neurology* 66(10 Suppl 4): S10-23.
- Schapira, A. H., J. M. Cooper, et al. (1989). "Mitochondrial complex I deficiency in Parkinson's disease." *Lancet* 1(8649): 1269.
- Schwartz, M. (2004). "Protective autoimmunity and prospects for therapeutic vaccination against self-perpetuating neurodegeneration." *Ernst Schering Res Found Workshop*(47): 133-54.
- Sekut, L., B. R. Champion, et al. (1995). "Anti-inflammatory activity of salmeterol: down-regulation of cytokine production." *Clin Exp Immunol* 99(3): 461-6.
- Sen, R. and D. Baltimore (1986). "Inducibility of kappa immunoglobulin enhancer-binding protein Nf-kappa B by a posttranslational mechanism." *Cell* 47(6): 921-8.
- Sen, R. and D. Baltimore (1986). "Multiple nuclear factors interact with the immunoglobulin enhancer sequences." *Cell* 46(5): 705-16.
- Severn, A., N. T. Rapson, et al. (1992). "Regulation of tumor necrosis factor production by adrenaline and beta-adrenergic agonists." *J Immunol* 148(11): 3441-5.
- Sherer, T. B., R. Betarbet, et al. (2002). "Environment, mitochondria, and Parkinson's disease." *Neuroscientist* 8(3): 192-7.
- Singh, S., K. Singh, et al. (2008). "Nicotine and caffeine-mediated modulation in the expression of toxicant responsive genes and vesicular monoamine transporter-2 in 1-methyl 4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinson's disease phenotype in mouse." *Brain Res* 1207: 193-206.
- Sizemore, N., A. Agarwal, et al. (2004). "Inhibitor of kappaB kinase is required to activate a subset of interferon gamma-stimulated genes." *Proc Natl Acad Sci U S A* 101(21): 7994-8.
- Sizemore, N., N. Lerner, et al. (2002). "Distinct roles of the Ikappa B kinase alpha and beta subunits in liberating nuclear factor kappa B (NF-kappa B) from Ikappa B and in phosphorylating the p65 subunit of NF-kappa B." *J Biol Chem* 277(6): 3863-9.
- Smeyne, M., Y. Jiao, et al. (2005). "Glia cell number modulates sensitivity to MPTP in mice." *Glia* 52(2): 144-52.
- Smith, A. D. and M. J. Zigmond (2003). "Can the brain be protected through exercise? Lessons from an animal model of parkinsonism." *Exp Neurol* 184(1): 31-9.
- Smith, T. W., U. DeGirolami, et al. (1992). "Neuropathology of immunosuppression." *Brain Pathol* 2(3): 183-194.
- Sofic, E., K. W. Lange, et al. (1992). "Reduced and oxidized glutathione in the substantia nigra of patients with Parkinson's disease." *Neurosci Lett* 142(2): 128-30.
- Spencer, J. P., P. Jenner, et al. (1998). "Conjugates of catecholamines with cysteine and GSH in Parkinson's disease: possible mechanisms of formation involving reactive oxygen species." *J Neurochem* 71(5): 2112-22.

- Stamatovic, S. M., R. F. Keep, et al. (2008). "Brain endothelial cell-cell junctions: how to "open" the blood brain barrier." *Curr Neuropharmacol* 6(3): 179-92.
- Strle, K., J. H. Zhou, et al. (2001). "Interleukin-10 in the brain." *Crit Rev Immunol* 21(5): 427-49.
- Suidan, G. L., J. W. Dickerson, et al. (2010). "CD8 T cell-initiated vascular endothelial growth factor expression promotes central nervous system vascular permeability under neuroinflammatory conditions." *J Immunol* 184(2): 1031-40.
- Sun, M., J. C. Latourelle, et al. (2006). "Influence of heterozygosity for parkin mutation on onset age in familial Parkinson disease: the GenePD study." *Arch Neurol* 63(6): 826-32.
- Szczepanik, M., M. Tutaj, et al. (2005). "Epicutaneously induced TGF-beta-dependent tolerance inhibits experimental autoimmune encephalomyelitis." *J Neuroimmunol* 164(1-2): 105-14.
- Tak, P. P. and G. S. Firestein (2001). "NF-kappaB: a key role in inflammatory diseases." *J Clin Invest* 107(1): 7-11.
- Tanaka, K. F., H. Kashima, et al. (2002). "Existence of functional beta1- and beta2-adrenergic receptors on microglia." *J Neurosci Res* 70(2): 232-7.
- Tanner, C. M. (2003). "Is the cause of Parkinson's disease environmental or hereditary? Evidence from twin studies." *Adv Neurol* 91: 133-42.
- Tansey, M. G., M. K. McCoy, et al. (2007). "Neuroinflammatory mechanisms in Parkinson's disease: potential environmental triggers, pathways, and targets for early therapeutic intervention." *Exp Neurol* 208(1): 1-25.
- Tashkin, D. P. and C. B. Cooper (2004). "The role of long-acting bronchodilators in the management of stable COPD." *Chest* 125(1): 249-59.
- Taylor, D. L., F. Jones, et al. (2005). "Stimulation of microglial metabotropic glutamate receptor mGlu2 triggers tumor necrosis factor alpha-induced neurotoxicity in concert with microglial-derived Fas ligand." *J Neurosci* 25(11): 2952-64.
- Teismann, P. and J. B. Schulz (2004). "Cellular pathology of Parkinson's disease: astrocytes, microglia and inflammation." *Cell Tissue Res* 318(1): 149-61.
- Tetrud, J. W., J. W. Langston, et al. (1989). "Mild parkinsonism in persons exposed to 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)." *Neurology* 39(11): 1483-7.
- Thomas, M. P., K. Chartrand, et al. (2007). "Ion channel blockade attenuates aggregated alpha synuclein induction of microglial reactive oxygen species: relevance for the pathogenesis of Parkinson's disease." *J Neurochem* 100(2): 503-19.
- Tortella, F. C., J. W. Ferkany, et al. (1988). "Anticonvulsant effects of dextrorphan in rats: possible involvement in dextromethorphan-induced seizure protection." *Life Sci* 42(24): 2509-14.
- Tsoufas, G. and D. A. Geller (2001). "NF-kappaB in transplantation: friend or foe?" *Transpl Infect Dis* 3(4): 212-9.
- Unsicker, K. and K. Kriegelstein (2002). "TGF-betas and their roles in the regulation of neuron survival." *Adv Exp Med Biol* 513: 353-74.
- Van der Heiden, K., S. Cuhlmann, et al. "Role of nuclear factor kappaB in cardiovascular health and disease." *Clin Sci (Lond)* 118(10): 593-605.

- van der Poll, T., J. Jansen, et al. (1994). "Noradrenaline inhibits lipopolysaccharide-induced tumor necrosis factor and interleukin 6 production in human whole blood." *Infect Immun* 62(5): 2046-50.
- Van Snick, J. (1990). "Interleukin-6: an overview." *Annu Rev Immunol* 8: 253-78.
- Vandenbroeck, K., I. Alloza, et al. (2004). "Inhibiting cytokines of the interleukin-12 family: recent advances and novel challenges." *J Pharm Pharmacol* 56(2): 145-60.
- Verhagen Metman, L., P. J. Blanchet, et al. (1998). "A trial of dextromethorphan in parkinsonian patients with motor response complications." *Mov Disord* 13(3): 414-7.
- Verhagen Metman, L., P. Del Dotto, et al. (1998). "Dextromethorphan improves levodopa-induced dyskinesias in Parkinson's disease." *Neurology* 51(1): 203-6.
- Vilar, R., H. Coelho, et al. (2007). "Association of A313 G polymorphism (GSTP1*B) in the glutathione-S-transferase P1 gene with sporadic Parkinson's disease." *Eur J Neurol* 14(2): 156-61.
- Wada, K., H. Arai, et al. (2006). "Expression levels of vascular endothelial growth factor and its receptors in Parkinson's disease." *Neuroreport* 17(7): 705-9.
- Wang, M. S., S. Boddapati, et al. "Curcumin reduces alpha-synuclein induced cytotoxicity in Parkinson's disease cell model." *BMC Neurosci* 11: 57.
- Webster, S. D., M. D. Galvan, et al. (2001). "Antibody-mediated phagocytosis of the amyloid beta-peptide in microglia is differentially modulated by C1q." *J Immunol* 166(12): 7496-503.
- Weng, Y. H., Y. H. Chou, et al. (2007). "PINK1 mutation in Taiwanese early-onset parkinsonism : clinical, genetic, and dopamine transporter studies." *J Neurol* 254(10): 1347-55.
- Werling, L. L., E. C. Lauterbach, et al. (2007). "Dextromethorphan as a potential neuroprotective agent with unique mechanisms of action." *Neurologist* 13(5): 272-93.
- Whitton, P. S. (2007). "Inflammation as a causative factor in the aetiology of Parkinson's disease." *Br J Pharmacol* 150(8): 963-76.
- Wu, D. C., V. Jackson-Lewis, et al. (2002). "Blockade of microglial activation is neuroprotective in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson disease." *J Neurosci* 22(5): 1763-71.
- Wyss-Coray, T. and L. Mucke (2002). "Inflammation in neurodegenerative disease--a double-edged sword." *Neuron* 35(3): 419-32.
- Xia, Y., M. E. Pauza, et al. (1997). "RelB regulation of chemokine expression modulates local inflammation." *Am J Pathol* 151(2): 375-87.
- Yamada, T., J. K. Chong, et al. (1993). "Concentration of neural thread protein in cerebrospinal fluid from progressive supranuclear palsy and Parkinson's disease." *Jpn J Psychiatry Neurol* 47(3): 631-5.
- Yamada, T., P. L. McGeer, et al. (1992). "Lewy bodies in Parkinson's disease are recognized by antibodies to complement proteins." *Acta Neuropathol* 84(1): 100-4.
- Yang, S., D. Zhang, et al. (2008). "Curcumin protects dopaminergic neuron against LPS induced neurotoxicity in primary rat neuron/glia culture." *Neurochem Res* 33(10): 2044-53.

- Yazdani, U., D. C. German, et al. (2006). "Rat model of Parkinson's disease: chronic central delivery of 1-methyl-4-phenylpyridinium (MPP+)." *Exp Neurol* 200(1): 172-83.
- Zhang, F., L. Qian, et al. (2010). "Inhibition of IkappaB kinase-beta protects dopamine neurons against lipopolysaccharide-induced neurotoxicity." *J Pharmacol Exp Ther* 333(3): 822-33.
- Zhang, S. X., J. J. Wang, et al. (2006). "Pigment epithelium-derived factor (PEDF) is an endogenous antiinflammatory factor." *FASEB J* 20(2): 323-5.
- Zhang, W., S. Dallas, et al. (2007). "Microglial PHOX and Mac-1 are essential to the enhanced dopaminergic neurodegeneration elicited by A30P and A53T mutant alpha-synuclein." *Glia* 55(11): 1178-88.
- Zhang, W., L. Qin, et al. (2005). "3-hydroxymorphinan is neurotrophic to dopaminergic neurons and is also neuroprotective against LPS-induced neurotoxicity." *FASEB J* 19(3): 395-7.
- Zhang, W., E. J. Shin, et al. (2006). "3-Hydroxymorphinan, a metabolite of dextromethorphan, protects nigrostriatal pathway against MPTP-elicited damage both in vivo and in vitro." *FASEB J* 20(14): 2496-511.
- Zhang, W., T. Wang, et al. (2004). "Neuroprotective effect of dextromethorphan in the MPTP Parkinson's disease model: role of NADPH oxidase." *FASEB J* 18(3): 589-91.
- Zhang, Z. G., L. Zhang, et al. (2000). "VEGF enhances angiogenesis and promotes blood-brain barrier leakage in the ischemic brain." *J Clin Invest* 106(7): 829-38.
- Zhou, Y., Y. Wang, et al. (2005). "Microglial activation induced by neurodegeneration: a proteomic analysis." *Mol Cell Proteomics* 4(10): 1471-9.
- Zhu, Y., G. Y. Yang, et al. (2002). "Transforming growth factor-beta 1 increases bad phosphorylation and protects neurons against damage." *J Neurosci* 22(10): 3898-909.
- Zlokovic, B. V. (2008). "The blood-brain barrier in health and chronic neurodegenerative disorders." *Neuron* 57(2): 178-201.



Etiology and Pathophysiology of Parkinson's Disease

Edited by Prof. Abdul Qayyum Rana

ISBN 978-953-307-462-7

Hard cover, 542 pages

Publisher InTech

Published online 12, October, 2011

Published in print edition October, 2011

This book about Parkinson's disease provides a detailed account of etiology and pathophysiology of Parkinson's disease, a complicated neurological condition. Environmental and genetic factors involved in the causation of Parkinson's disease have been discussed in detail. This book can be used by basic scientists as well as researchers. Neuroscience fellows and life science readers can also obtain sufficient information. Beside genetic factors, other pathophysiological aspects of Parkinson's disease have been discussed in detail. Up to date information about the changes in various neurotransmitters, inflammatory responses, oxidative pathways and biomarkers has been described at length. Each section has been written by one or more faculty members of well known academic institutions. Thus, this book brings forth both clinical and basic science aspects of Parkinson's disease.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Lynda J. Peterson and Patrick M. Flood (2011). Inflammatory Responses and Regulation in Parkinson's Disease, Etiology and Pathophysiology of Parkinson's Disease, Prof. Abdul Qayyum Rana (Ed.), ISBN: 978-953-307-462-7, InTech, Available from: <http://www.intechopen.com/books/etiology-and-pathophysiology-of-parkinson-s-disease/inflammatory-responses-and-regulation-in-parkinson-s-disease>

INTECH

open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.