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Chapter 2

Fundamentals of Medicinal Application of Titanium Dioxide Nanoparticles

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Abstract

Titanium dioxide (TiO$_2$), a semiconducting material, is a well-known photocatalyst. A nanoparticle (NP) of TiO$_2$ also demonstrates photocatalytic activity. Photo-irradiated TiO$_2$ NPs induce the formation of various reactive species, leading to the damage of biomacromolecules. These reactive species include $\cdot$OH, either free or trapped hydroxyl radicals (OH$^-$), superoxide (O$_2^-$), hydrogen peroxide (H$_2$O$_2$), and singlet oxygen (O$_2^\text{1}$), among others. TiO$_2$ NPs photocatalyze DNA oxidation. A relatively small concentration of TiO$_2$ NPs frequently induces tandem base oxidation at guanine and thymine residues through H$_2$O$_2$ generation in the presence of a copper(II) ion. A copper-peroxo complex is considered to be an important reactive species responsible for this DNA damage. In the case of a high concentration of TiO$_2$ NPs, OH$^-$ contributes to DNA damage without sequence specificity. In the presence of sugars, TiO$_2$ NPs indirectly induce DNA damage by the secondary H$_2$O$_2$, which is produced through an autoxidation process of the product of sugar photooxidized by TiO$_2$ NPs. Furthermore, O$_2^\text{1}$ is also produced by photo-irradiated TiO$_2$ NPs. The photocatalyzed formation of O$_2^\text{1}$ might contribute to the oxidation of the membrane protein. These mechanisms of photocatalytic formation of the reactive species may be involved in the photocytotoxicity of TiO$_2$ NPs.

Keywords: Titanium dioxide, Photocatalyst, Reactive oxygen species, Photomedicine, DNA damage

1. Introduction

Titanium dioxide (TiO$_2$), a semiconducting material, is a well-known photocatalyst [1-5]. Examples of previous studies about TiO$_2$ photocatalytic reactions are listed in Table 1. A nanoparticle (NP) of TiO$_2$ also demonstrates photocatalytic activity. Important applications of TiO$_2$ photocatalysts are bactericidal activity [2-4, 6-12] and degradation of chemical pollutants.
Related physical and chemical mechanisms have been also investigated [2-5, 14-17]. Photo-irradiated TiO$_2$ NPs induce the formation of various reactive species, leading to the damage of biomacromolecules. These reactive species include hole (h$^+$), either free or trapped hydroxyl radicals (OH$^-$), superoxide (O$_2^-$), hydrogen peroxide (H$_2$O$_2$), and singlet oxygen ($^1$O$_2$), among others. Hydroxyl radicals, O$_2^-$, H$_2$O$_2$, and $^1$O$_2$ are the typical reactive oxygen species. TiO$_2$ photocatalysts have been found to kill cancer cells [18-21] other than bacteria, viruses, and algae under ultraviolet-A (wavelength: 315–400 nm) illumination [2-4, 6-12]. Therefore, one of the potential applications of the TiO$_2$ NP photocatalyst is photodynamic therapy (PDT), which is a promising treatment for cancer and some nonmalignant conditions [22-25]. In general, the mechanism of cytotoxicity by the photocatalysis of TiO$_2$ is based on cell membrane damage via the generation of the aforementioned reactive oxygen species. Furthermore, DNA damage in human cells [26-28], mouse lymphoma cells [29], and phage [30] by the TiO$_2$ NP photocatalyst has been reported. Direct damage of isolated DNA by TiO$_2$ photocatalyst in vitro has been also studied [31, 32]. However, the DNA-damaging mechanism in vivo is not well-understood, because the incorporation of the TiO$_2$ NPs in the nucleus is difficult [18]. A previous study has shown that H$_2$O$_2$ formation through the photocatalytic reaction of TiO$_2$ may contribute to cellular DNA damage [2, 19]. Hydrogen peroxide, a long-lived reactive oxygen species, can penetrate the nucleus membrane and induce oxidation of the nucleobase and strand breakage through enhancement by metal ions. Iron or copper ions can enhance the activity of H$_2$O$_2$ to produce OH$^-$ [33] and copper-peroxide [34-36]. Furthermore, secondary generation of reactive oxygen species may contribute to cytotoxicity of TiO$_2$ NPs photocatalyst [37]. Since the photocatalytic reaction will occur in a complex biological environment, an interaction between TiO$_2$ NPs and biomaterials should participate in the generation of reactive species to induce DNA damage. For example, sugars photocatalyzed by TiO$_2$ NPs may secondarily generate H$_2$O$_2$ through their further oxidation process by molecular oxygen in the presence of a metal ion [37]. In addition, the possibility of $^1$O$_2$-mediated cytotoxicity by TiO$_2$ NPs has been proposed [38]. Actually, $^1$O$_2$ generation by photo-irradiated TiO$_2$ NPs was demonstrated by a near-infrared spectroscopy [39, 40]. In this chapter, recent studies about photocatalytic biomacromolecule damage by TiO$_2$ NPs are briefly reviewed.

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Table 1. Summary of the examples of previous studies on TiO$_2$ photocatalyst
1.1. General mechanism of photocatalysis of TiO$_2$ NP

The crystal of TiO$_2$ is a semiconductor, and the two crystalline forms, anatase and rutile, are well-known (Figure 1) [2-5]. The values of the band gap energy of these crystal forms are 3.26 and 3.06 eV for anatase and rutile, respectively. Photo-irradiation to a TiO$_2$ crystal induces the formation of an excited electron (e$^-$) in the conduction band and an h$^+$ in the valence band, leading to the redox reaction of materials adsorbing on the TiO$_2$ surface, including water and/or molecular oxygen. The photocatalytic reactions with its surface water and oxygen cause the formation of various reactive oxygen species such as free or trapped OH$^-$, O$_2$$^-$, H$_2$O$_2$, and ‘O$_2$’ [2-5].

![Diagram](insert diagram)

**Figure 1.** Band gap energy of the two crystalline forms of TiO$_2$.

An excited electron in the conductive band reduces the oxygen molecule adsorbed on the surface of TiO$_2$ NPs, leading to the generation of various reactive oxygen species as follows (Figure 2):

\[
O_2 + e^- \rightarrow O_2^- \quad (1)
\]

\[
2O_2^- + 2H^+ \rightarrow H_2O_2 + O_2 \quad (2)
\]

\[
H_2O_2 \rightarrow 2OH \quad (3)
\]

The reaction (3) is mediated by ultraviolet radiation (hv, wavelength <355 nm), metal ions (M$^{n+}$) such as Fe$^{2+}$, and O$_2$$^-$, as follows [33]:

\[
H_2O_2 + hv \rightarrow 2OH \quad (4)
\]

\[
H_2O_2 + M^{n+} \rightarrow OH^- + OH + M^{(n+1)+} \quad (5)
\]
On the other hand, the formed \( h^+ \) in the valence band can oxidize water to form \( \text{OH}^- \) as follows:

\[
\text{H}_2\text{O} + h^+ \rightarrow \text{OH}^- + \text{H}^+
\]

Furthermore, \( \text{OH}^- \) can produce \( \text{H}_2\text{O}_2 \) as follows:

\[
2\text{OH} \rightarrow \text{H}_2\text{O}_2
\]

These reactive oxygen species should contribute to the mechanism of the phototoxicity induced by \( \text{TiO}_2 \) NPs.
1.2. Sterilization effect by TiO₂

One of the most important medicinal applications of TiO₂ NPs is to kill bacteria on its surfaces. TiO₂ NPs under ultraviolet radiation produce a strong oxidative effect through the formation of above-mentioned reactive oxygen species and can be used as a photocatalytic disinfectant without other chemical reagents. Fujishima and coworkers reported the bactericidal effect of TiO₂ photocatalysts against Escherichia coli under ultraviolet-A irradiation using black light [6]. This is the first report of the application of phototoxicity of TiO₂ NPs. It was speculated that H₂O₂ was a reactive species responsible for this phototoxic effect [7]. Relevantly, the photocatalytic effect of TiO₂ against methicillin-resistant Staphylococcus aureus (MRSA) and Clostridium difficile in hospitals has been reported [10]. The bactericidal effect of TiO₂ NPs could be enhanced by metal doping [9]. Furthermore, visible-light-induced TiO₂ photocatalysts were developed and utilized in antibacterial applications. For example, sulfur-doped TiO₂ demonstrates the killing effect on Escherichia coli under white-light irradiation commonly used in hospitals [11].

1.3. Photodynamic therapy

Photodynamic therapy, which is a promising and less-invasive treatment for cancer, employs a photosensitizer and visible light to produce oxidative stress in cells and ablate cancerous tumors [22-25]. Photodynamic therapy is also used for treating some nonmalignant conditions that are generally characterized by the overgrowth of unwanted or abnormal cells. In general, porphyrins are used as photosensitizers under visible-light irradiation, since the human tissue has relatively high transparency for visible light, especially red light, and visible light has hardly any side effects. In the case of visible light PDT, ¹O₂ is considered an important reactive species for PDT because ¹O₂ can be easily generated by visible light [41-44]. Critical targets of the generated ¹O₂ include mitochondria and enzyme proteins. Moreover, DNA is also an important target biomolecule of photosensitized reactions [45-49]. Relevantly, photocatalytic ¹O₂ generation by TiO₂ has been reported [38-40].


TiO₂, a nontoxic material, is chemically stable, and demonstrates a phototoxic effect. Therefore, an application of TiO₂ for PDT has been investigated [2]. The cytotoxicity of an illuminated TiO₂ film electrode for HeLa cells [18,19] and T-24 human bladder cancer cells [21] has been reported. Animal experiments also demonstrated the antitumor effect of TiO₂ NPs [20]. This report showed an antineoplastic effect on skin cancer in mouse models.

2. Photocatalytic DNA damage by TiO₂ NPs

Cellular DNA damage photocatalyzed by TiO₂ NPs was demonstrated by the experiment using cancer cells [18,19,21]. TiO₂ NPs can be taken into the cancer cell [27]; however, incorporation into the cell nucleus is difficult [18]. Therefore, it is speculated that the indirect mechanism contributes to DNA damage induced by photo-irradiated TiO₂ NPs. Hence, model experiments using isolated DNA were performed [31, 32]. In this section, an example of photocatalytic DNA damage by TiO₂ NPs was introduced.

2.1. Isolated DNA damage photocatalyzed by TiO₂ NPs and its sequence specificity

Photo-irradiated TiO₂ NPs catalyze DNA damage in the presence of copper(II) ion [31]. Relevantly, copper-aided photosterilization of microbial cells on TiO₂ was reported [8]. DNA damage by anatase NPs is more severe than that by rutile NPs. The DNA damage is enhanced by piperidine treatment, because photo-irradiated TiO₂ NPs cause not only DNA strand breakage but also base oxidation. In general, hot piperidine cleaves DNA strand at modified base. Photo-irradiated TiO₂ NPs induce the formation of piperidine-labile products at the bolded site of 5’-TG, 5’-TG, and 5’-TC (Figure 4). Furthermore, TiO₂ NPs photocatalyze DNA strand cleavage at the bolded guanines of 5’-TG and 5’-TC in a DNA fragment treated with *E coli* formamidopyrimidine-DNA glycosylase (Fpg protein), which can catalyze the excision of piperidine-resistant 8-oxo-7,8-dihydro-2’deoxyguanine (8-oxo-G) [50,51]. The formation of 8-oxo-G was confirmed by an analysis with a high-performance liquid chromatography (Figure 5). In addition, Fpg protein can cleave the oxidized cytosine, such as 5-hydroxy cytosine [52]. These results suggest that photo-irradiated TiO₂ NPs induce 8-oxo-G formation adjacent to piperidine-labile thymine lesions. Such double-base lesions should be generated from one radical hit that leads through a secondary reaction to a tandem base damage at pyrimidine and adjacent residues [53-56]. Actually, it has been reported that H₂O₂ induces tandem mutations in human cells via vicinal or cross-linked base modification in the presence of copper(II) ion [57]. Since repairing of cluster DNA damage in living cells is difficult [58], such clustered base damage, including double-base lesions, appears to play an important role in the phototoxicity of TiO₂ NPs.

2.2. Mechanism of DNA damage photocatalyzed by TiO₂ NPs

Catalase, a well-known scavenger of H₂O₂, and bathocuproines, a copper(I) ion chelator, inhibit DNA damage photocatalyzed by TiO₂ NPs, whereas, typical OH⁻ scavenger cannot inhibit the DNA damage. These results suggest that H₂O₂ and copper(I) ion participate in DNA damage
by photo-irradiated TiO$_2$ NPs. It has been reported that OH$^\cdot$ is not the main reactive species involved in DNA damage by H$_2$O$_2$ and copper(I) ions [34-36, 59]. DNA-associated copper(I) ions may generate other oxidants, including a copper–peroxo intermediate, such as Cu(I)-OOH, which is generated from the reaction of H$_2$O$_2$ and copper(I) ions [34-36, 59]. Indeed,
methional, which can scavenge Cu(I)-OOH [36, 59], shows inhibitory effect on DNA damage photocatalyzed by TiO$_2$ NPs. The generation of these reactive species may be responsible for the formation of piperidine-labile products and 8-oxo-G.

On the other hand, a high concentration of anatase NPs can catalyze DNA photodamage without copper(II) ions. Typical OH$^\cdot$ scavengers, ethanol and sugars, effectively inhibit the DNA photodamage by a high concentration of anatase NPs. The DNA damage induced by photo-irradiated anatase NPs without copper(II) ions is observed at every nucleobases without site specificity. Such DNA damage without sequence-specificity is the typical pattern of OH$^\cdot$-mediated DNA damage [34].

A proposed mechanism of DNA damage photocatalyzed by TiO$_2$ NPs is shown in Figure 6. The crystalline forms of TiO$_2$, anatase and rutile, are semiconductors with band gap energies of 3.26 and 3.06 eV, which correspond to the following wavelengths of light: 385 and 400 nm, respectively. When a TiO$_2$ semiconductor NPs absorbs photon with energy greater than their band gap, electrons in the valence band are excited to the conduction band, creating electron-h$^+$ pairs and causing various chemical reactions [2-5]. The electron acts as a reductant, whereas the h$^+$ is a powerful oxidant. In aqueous environments, oxygen molecule can be reduced by the electron into O$_2^\cdot$-, and water molecule can be oxidized by the h$^+$ into OH$^\cdot$. In general, formed O$_2^\cdot$- can be dismutated into H$_2$O$_2$ by proton. The oxygen reduction may precede the reduction of copper(II) ions under aerobic condition, since the concentration of dissolved oxygen is higher (~250 μM) than that of the copper(II) ion used in this study (20 μM). The copper(II) reduction may be mediated by O$_2^\cdot$-. Hydrogen peroxide reacts with copper(I) ions to generate other oxidants, including a copper–peroxo intermediate, resulting in the oxidation of DNA bases. Copper ions, which are essential components of chromatin [60,61], are found to bind DNA with high affinity [62,63]. Therefore, copper ions may play an important role in reactive oxygen generation in vivo, although mammals have evolved means of minimizing the levels of free copper ions and most copper ions bind to protein caries and transporters [64]. Hydroxyl radicals formed by the reaction of water with an h$^+$ in the valence band of TiO$_2$ NPs also slightly participate in DNA damage photocatalyzed by anatase NPs. Because OH$^-$ is a strong oxidative agent, OH$^-$ can damage every nucleobase [34]. The present results suggest that H$_2$O$_2$ mainly participate in the phototoxicity of TiO$_2$ NPs and the contribution of OH$^-$ is relatively small. Fujishima et al. also reported the involvement of H$_2$O$_2$ generated from O$_2^\cdot$- in the cytotoxicity of illuminated TiO$_2$ NPs [2-4, 8-13].

TiO$_2$ NPs might be a potential agent for PDT [22-25]. TiO$_2$ NPs can be incorporated into cancer cells and demonstrate cytotoxicity under photo-irradiation [2-4, 26-28]. Photocatalytic reaction by TiO$_2$ NPs induces a number of functional changes in cell including altered permeability of cellular membranes to potassium and calcium ions, release of RNA and proteins and cytotoxicity [2,18-21]. It has been reported that DNA can be a target biomolecule of the photocatalytic reaction of TiO$_2$ NPs [26-30]. Although incorporation of TiO$_2$ NPs into cell nucleus is difficult [18], the generated H$_2$O$_2$ by a photocatalytic reaction of TiO$_2$ NPs can be easily diffused and incorporated in a cell nucleus, leading to DNA photodamage with metal ions. Relevantly, several studies demonstrated that DNA is a potential target of PDT [47,65,66]. Therefore, the
Metal-mediated DNA damage through the photocatalysis of TiO$_2$ NPs may participate in cytotoxicity by photo-irradiated TiO$_2$ NPs.

3. Secondary production of reactive oxygen species from photocatalyzed materials by TiO$_2$ NPs

As mentioned above, DNA damage in human cells by TiO$_2$ NPs has also been reported [26-28]. The direct DNA damage by TiO$_2$ NPs photocatalyst in vitro has been also studied [31, 32]. However, the DNA-damaging mechanism in vivo is not well-understood because the incorporation of the TiO$_2$ NPs in the cell nucleus is difficult [18]. Since the TiO$_2$ photocatalytic reaction occurs in a complex biological environment, an interaction between TiO$_2$ NPs and biomaterials may participate in the generation of reactive species to induce DNA damage. Hence, the effect of sugars, which are ubiquitous biomaterials, on DNA damage photocatalyzed by TiO$_2$ NPs was examined [37].

In the case of anatase, a high concentration of TiO$_2$ NPs can damage DNA at every nucleobase by OH$^-$ generation in the absence of copper(II) ions. Typical free OH$^-$ scavengers inhibited this copper(II)-independent DNA damage. These results indicate that free OH$^-$ partly contributes to DNA damage photocatalyzed by TiO$_2$. On the other hand, scavengers of OH$^-$, such as a sugar (mannitol), ethanol, and formate, enhanced the copper(II)-dependent DNA damage [31]. These scavengers themselves did not induce DNA damage. Since OH$^-$ can oxidize most biomaterials, the oxidized products of biomaterials by the TiO$_2$ photocatalyst may damage DNA via the generation of secondary reactive oxygen species. The addition of sugars, glucose and galactose, which are ubiquitous biomolecules, enhanced the DNA damage photocatalyzed by TiO$_2$ NPs. Enhancement of DNA damage by sugars has seldom been reported, and these sugars themselves could not induce DNA damage. Therefore, the products of the photocatalytic reaction of these sugars by TiO$_2$ NPs is responsible for the copper(II)-dependent damage to DNA. Indeed, the glucose and galactose oxidized by the TiO$_2$ photocatalytic reaction caused...
DNA damage in the presence of copper(II) ion [37]. The inhibitory effect of various scavengers for DNA damage by the photo-oxidized products of sugars by TiO\textsubscript{2} was examined. Catalase inhibited DNA damage by the photocatalyzed glucose, indicating the involvement of H\textsubscript{2}O\textsubscript{2}. Bathocuproine, which is a chelator of copper(I) ion, also inhibited DNA damage by the photocatalyzed glucose, suggesting the involvement of copper(I) ion. The free OH\textsuperscript{•} scavengers had no or little inhibitory effect on DNA damage. The inhibitory effect of superoxide dismutase (SOD) was weak, suggesting that O\textsubscript{2}\textsuperscript{•} itself is not the main reactive species for DNA damage. Similar results were observed in the case of galactose. Fluorometry using folic acid [67] demonstrated the formation of H\textsubscript{2}O\textsubscript{2} from the photocatalyzed sugars (Figure 7). The amount of H\textsubscript{2}O\textsubscript{2} generation was comparable with that of other H\textsubscript{2}O\textsubscript{2}-mediated DNA-damaging drugs [68]. H\textsubscript{2}O\textsubscript{2} generation was not observed in the absence of copper(II) ions. These results showed that the oxidized products of sugars generate H\textsubscript{2}O\textsubscript{2} during the reaction with copper(II) ions, resulting in secondary DNA damage.

![Figure 7](image_url)

**Figure 7.** Hydrogen peroxide generation from photo-oxidized glucose and galactose by TiO\textsubscript{2} NPs. The buffer solution with 10 mM sugars was previously irradiated (365 nm, 6 J cm\textsuperscript{-2}) with 100 μg mL\textsuperscript{-1} anatase NPs. The TiO\textsubscript{2} NPs were removed by centrifugation, and the solution containing the oxidized sugars was used. One mL of solution containing the treated sugars and 10 μM of folic acid was incubated (60 min, 37 °C) in the presence of 20 μM copper(II) chloride, and the fluorescence intensity was measured (excitation: 360 nm, detection: 450 nm). The concentration of the generated H\textsubscript{2}O\textsubscript{2} was determined by the calibration curve method.

These sugars act as an electron donor for the photocatalytic reaction [15,37]. Partially oxidized sugars, such as aldehyde compounds, are possibly produced through this photocatalytic oxidation. The mechanism of DNA damage by the photocatalyzed product of sugars is proposed in Figure 8. Aldehydes can generate H\textsubscript{2}O\textsubscript{2} via its further oxidation [69], though these sugars themselves are stable compounds. Many studies have reported DNA damage by H\textsubscript{2}O\textsubscript{2} and copper(II) ions [34-36, 70]. Various chemical compounds, including aldehydes, easily produce O\textsubscript{2}\textsuperscript{•} through their autoxidation process. The autoxidation is markedly enhanced by copper(II) ion, which is an essential component of chromatin [60, 61]. The formed O\textsubscript{2}\textsuperscript{•} is rapidly dismutated into H\textsubscript{2}O\textsubscript{2}. Although the generated H\textsubscript{2}O\textsubscript{2} itself cannot damage DNA, H\textsubscript{2}O\textsubscript{2} reduces copper(II) into copper(I), leading to the activation of H\textsubscript{2}O\textsubscript{2} through the formation of reactive
species, such as Cu(I)-OOH [34-36, 59]. Indeed, methional, a scavenger of Cu(I)-OOH, inhibited the DNA damage. This reactive species cannot be scavenged by the free OH− scavengers; however, it can effectively oxidize the nucleobases [34-36, 59].

Figure 8. Proposed mechanism of secondary DNA damage by photocatalyzed sugars.

Although TiO₂ is not likely to be incorporated in a cell nucleus [18], H₂O₂ generated via a photocatalytic reaction can be easily diffused and incorporated in a cell nucleus. This DNA-damaging mechanism via H₂O₂ generation may participate in the phototoxicity of TiO₂. In vivo, the cell membrane is an important reaction field for the TiO₂ photocatalyst because TiO₂ NPs show affinity with a cell membrane [18]. Further, a part of the TiO₂ NPs can become incorporated into the cell. Sugars on the cell membrane and cytoplasm may be oxidized by the TiO₂ photocatalytic reaction. The generated h+ and OH− can oxidize these sugars, leading to the formation of secondary H₂O₂ from their photo-oxidized products.

In summary, sugars enhance the DNA damage photocatalyzed by TiO₂ NPs. This enhancement of DNA damage is due to the secondary generation of a reactive oxygen species, H₂O₂, which can diffuse in the cell and damage cellular DNA. These findings suggest that the secondary H₂O₂ generation contributes to the phototoxicity of TiO₂ more than the direct formation of reactive oxygen species does.

4. Singlet oxygen formation through photocatalytic reaction of TiO₂ NPs

A contribution of ¹O₂ in the TiO₂ photocatalytic reaction was reported [38]. Singlet oxygen generation by TiO₂ photocatalysis has been demonstrated by the emission measurement of ¹O₂, which is assigned to the transition from ¹O₂(¹A₂) to ³O₂(³Σg−) [39, 40]. Because ¹O₂ is considered to be an important reactive species in PDT process [22-25], the clarification of the
contribution of $^{1}\text{O}_2$ generated by TiO$_2$ photocatalysis is closely related to a design of photocatalyst for medicinal application. Thus, $^{1}\text{O}_2$ generation in the TiO$_2$ photocatalysis and its importance on biomolecular damage was examined [40].

The typical emission of $^{1}\text{O}_2$ at around 1270 nm was observed during irradiation of TiO$_2$ NPs. Relatively strong emission of $^{1}\text{O}_2$ was observed in nonpolar organic solvents such as dichloromethane. The quantum yield ($\Phi_\lambda$) of $^{1}\text{O}_2$ generation by TiO$_2$ photocatalysis in ethanol was estimated from the comparison of $^{1}\text{O}_2$ emission intensities by TiO$_2$ NPs and methylene blue ($\Phi_\lambda$= 0.52) [71] and the absorbance of the TiO$_2$ NP dispersions. Because the scattering by suspended TiO$_2$ NPs makes the calculation of absorbed light intensity complex, the precise estimation of the $\Phi_\lambda$ is difficult. Thus, the $\Phi_\lambda$ was estimated using the apparent absorbance of TiO$_2$ NPs. The calculated value indicates the lowest limit of the $\Phi_\lambda$ by TiO$_2$ photocatalysis in ethanol. The reported lifetime of $^{1}\text{O}_2$ generated via TiO$_2$ photocatalytic reaction is 5 μs [39]. This value is shorter than that by the photosensitized reaction of methylene blue (12 μs) [72].

Since the emission intensity of $^{1}\text{O}_2$ is proportional to its lifetime, the $\Phi_\lambda$ was corrected by the lifetime of $^{1}\text{O}_2$. The estimated value of $\Phi_\lambda$ by both types of TiO$_2$, anatase and rutile, was about 0.02 in ethanol. This value of $\Phi_\lambda$ is enough large to induce oxidative damage to biomolecules. The $^{1}\text{O}_2$ emission in D$_2$O was completely quenched by the addition of SOD, which is the enzyme to dismutate O$_2$- into H$_2$O$_2$. These results can be explained by the fact that $^{1}\text{O}_2$ is formed by the reoxidation of O$_2$-, generated from the photoreduction of oxygen molecules by TiO$_2$ NPs (Figure 3). The intensity of $^{1}\text{O}_2$ emission observed in the case of rutile was significantly larger than that by anatase in D$_2$O. The difference of the $^{1}\text{O}_2$ generation by these two types of TiO$_2$ crystalline forms can be reasonably explained by that in aqueous solution. H$_2$O$_2$ generation proceeds in the photocatalysis of anatase rather than O$_2$- generation, whereas O$_2$- is the main product from oxygen photoreduction mediated by rutile [17]. These results support the mechanism of $^{1}\text{O}_2$ generation via O$_2$- by TiO$_2$ photocatalysis.

The emission spectrum of $^{1}\text{O}_2$ by TiO$_2$ (in both, anatase and rutile type cases) slightly blue-shifted (~4 nm) compared with that by methylene blue. These results suggest that the surroundings of the $^{1}\text{O}_2$ generated on the TiO$_2$ surface are different from that by methylene blue in solution. In the case of the photosensitization of methylene blue, the generated $^{1}\text{O}_2$ deactivates in the homogeneous media of solvents. A possible explanation of the blue-shift is that most of the $^{1}\text{O}_2$ generated by TiO$_2$ NPs deactivates on the TiO$_2$ surface.

The intensity of $^{1}\text{O}_2$ emission by TiO$_2$ photocatalysis in liposome was significantly larger than that in an aqueous solution in both, anatase and rutile type cases. The enhancement of the $^{1}\text{O}_2$ emission can be explained by the elongation of the lifetime of $^{1}\text{O}_2$ or the acceleration of the photocatalytic reaction. This result shows that phospholipids membrane is an important environment of the phototoxic reaction mediated by $^{1}\text{O}_2$ in the photocatalytic reactions of TiO$_2$ NPs. Indeed, high affinity of TiO$_2$ NPs with a cell membrane was reported [18]. Consequently, an environmental effect of a cell membrane is important for the photocatalytic reaction of TiO$_2$ NPs. Since amino acid residues in proteins can be oxidized by $^{1}\text{O}_2$ [42], a membrane protein should be the target biomolecule in cell membrane. Indeed, $^{1}\text{O}_2$ emission was quenched by the addition of bovine serum albumin, a typical water soluble protein, suggesting scavenging of the $^{1}\text{O}_2$ generated by TiO$_2$ photocatalysis through oxidation of protein.
In vivo, nicotinamide adenine dinucleotide (NADH) is one of the most important target biomolecule oxidized by $^{1}\text{O}_2$ [73, 74]. NADH demonstrates the typical absorption peak at around 340 nm in an ultraviolet absorption spectral measurement, and this absorption band is diminished by the oxidation. It has been reported that TiO$_2$ NPs hardly induce the oxidation of NADH in aqueous solution during ultraviolet irradiation. Since NADH hardly adsorbed on a surface of TiO$_2$ NPs, the $^{1}\text{O}_2$ could not effectively oxidize NADH in solution. As mentioned above, it has been reported that photo-irradiated TiO$_2$ NPs can induce DNA damage mainly through $\text{H}_2\text{O}_2$ and $\text{OH}^\cdot$, and the $^{1}\text{O}_2$-mediated DNA damage could not be observed [31]. These reports concluded that the photocatalytic $^{1}\text{O}_2$ generation on the surface of TiO$_2$ NPs is not important in the damaging mechanism of the biomolecules such as DNA and NADH, of which the affinity with TiO$_2$ surface is small.

In conclusion, photo-irradiated TiO$_2$ NPs can produce $^{1}\text{O}_2$ through reoxidation of $\text{O}_2^{\cdot-}$, which is formed by photocatalytic reduction of oxygen molecule on the surface of TiO$_2$ NPs. Since most of the $^{1}\text{O}_2$ deactivated on TiO$_2$ surface, the $^{1}\text{O}_2$ on TiO$_2$ surface cannot induce the oxidation of DNA and NADH. However, the $^{1}\text{O}_2$ generation by TiO$_2$ photocatalysis could be enhanced in the microenvironment of phospholipids membrane. These findings suggest that $^{1}\text{O}_2$ may contribute to phototoxicity of TiO$_2$ NPs through oxidation of membrane protein.

5. Conclusions

TiO$_2$ NPs photocatalyze DNA oxidation. A relatively small concentration of TiO$_2$ NPs frequently induces tandem base oxidation at guanine and thymine residues through $\text{H}_2\text{O}_2$ generation in the presence of a copper(II) ion. A copper–peroxo complex is considered to be an important reactive species responsible for this DNA damage. In addition, cytosine residues are also photooxidized by TiO$_2$ NPs. In the case of a high concentration of TiO$_2$ NPs, OH$^\cdot$ contributes to DNA damage without sequence specificity. In the presence of sugars, TiO$_2$ NPs indirectly induce DNA damage by the secondary $\text{H}_2\text{O}_2$, which is produced through an autoxidation process of the photo-oxidized products of sugars by TiO$_2$ NPs. Furthermore, $^{1}\text{O}_2$ is also produced by photo-irradiated TiO$_2$ NPs. The $^{1}\text{O}_2$ generation is explained by the reoxidation of $\text{O}_2^{\cdot-}$, which is produced by photocatalytic reduction of the oxygen molecule adsorbed on the surface of TiO$_2$ NPs. The photocatalyzed formation of $^{1}\text{O}_2$ might contribute to the oxidation of the membrane protein. These mechanisms of photocatalytic reactive oxygen formation should be involved in the photocytotoxicity of TiO$_2$ NPs. Because TiO$_2$ is a chemically stable and nontoxic material, the bactericidal activity and cytotoxicity against cancer cells will play more important roles in the field of medical applications of nanomaterials.

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