Pediatric Cranial Ultrasound:  
Techniques, Variants and Pitfalls  

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1. Introduction  
Recent advances in sonographic technology have made ultrasound an increasingly accurate and adequate means to detect cranial morphology and intracranial pathology in infants. When modern equipment is combined with thorough imaging technique, ultrasound delivers similar results to MRI in terms of sensitivity and ability to direct initial management (Daneman et. al, 2006; Epelman et. al., 2010). Furthermore, ultrasound offers certain advantages over other imaging modalities such as MRI and CT. Cranial sonography is less expensive, spares the patient from radiation, does not require sedation, and its portability allows for bedside evaluation in gravely ill infants who cannot be transported to radiology for imaging. The goal of this chapter is to provide an updated approach to modern cranial ultrasound in order to improve diagnostic accuracy and increase recognition of normal anatomical structures, anatomical variants, pathologic processes, and imaging pitfalls that may simulate disease. The discussion will begin with an overview of the approach and interpretation of gray-scale imaging. The chapter will continue with a review of Doppler imaging, the use of additional fontanels, and linear imaging. The chapter will conclude by reviewing pitfalls and normal variants which may simulate pathology.  

2. Ultrasound techniques and interpretation  
2.1 Gray-scale  
The cranial ultrasound examination is performed with a linear-array transducer. Six-eight coronal plane images are taken through the anterior fontanel, beginning in the frontal lobes anterior to the frontal horns and progressing posteriorly to the occipital lobes past the trigones of the lateral ventricles. (Figure 1) The first coronal image acquired at the level of the frontal lobes allows the observer to examine the frontal lobes, orbital cones, and the hyperechoic falx cerebri located within the interhemispheric fissure (Slovis et. al., 1981). The second coronal image is taken through the frontal horns of the lateral ventricles, which allows multiple structures to be imaged at once. The frontal horns and cavum septum pellucidum appear as
anechoic, CSF-filled spaces. The corpus callosum is a linear, hypoechoic structure crossing the hemispheres that is contained within echogenic superior and inferior borders. Finally, the globus pallidus, putamen, caudate nucleus, and thalamus of each side can be visualized in their respective locations with their characteristic gray matter appearance (Seigel, 2002). The third image is obtained at the level of the foramen of Monro and further illustrates many of the structures mentioned above. The third ventricle can be visualized as an anechoic structure beneath the septum pellucidum. Brainstem structures such as the pons and medulla can also be seen (Seigel, 2002). Moving posteriorly, the fourth image is taken at the level of the cerebral peduncles and shows hyperechoic choroid plexus along the floor of the lateral ventricles and the roof of the third ventricle. The tentorium cerebelli and fourth ventricle are also demonstrated (Seigel, 2002). The quadrigeminal plate cistern serves as a marker for the fifth image, which enables the viewer to see both temporal horns of the lateral ventricles as well as cerebellar structures including the hemispheres and vermis. Below the vermis, the cisterna magna is visible. Occasionally, the mammillary bodies can be discerned on this image (Seigel, 2002). The sixth image is taken through the lateral ventral trigones and shows their innate hyperechoic choroid plexus (Seigel, 2002). The periventricular white matter halo is also seen and should be less echogenic than the choroid plexus (Grant et al., 1981; Seigel, 2002). The final image is obtained through the cerebral convexities and displays the interhemispheric fissure and occipital lobes (Seigel, 2002).
Fig. 1. (a-g). Coronal images of a normal neonatal brain. C (caudate) t (thalamus) f (Foramen of Monro) m (midbrain) 3 (third ventricle) Q (quadrigeminal plate cistern) CB (cerebellar hemispheres)

Next, the transducer is turned 90 degrees on the anterior fontanel, and five more images are acquired in the sagittal and parasagittal planes. (Figure 2) The first image is a midline sagittal view which includes the corpus callosum and cerebellar vermis. Additional parasagittal views are obtained laterally on either side, allowing for the inspection of the lateral horns and choroid plexus (Epelman et. al., 2010; Seigel 2002; Rumack et. al. 2005). The first midline sagittal image displays a number of important structures. The C-shaped corpus callosum appears as a hypoechoic structure bound by echogenic superior and inferior borders (Seigel, 2002). The cingulate gyrus is seen above the corpus callosum, while the septum cavum pellucidum is seen below. The third and fourth ventricles along with the cisterna magna appear anechoic at their respective positions (Seigel, 2002). The white matter containing pons and cerebellar vermis are also seen (Grant et. al., 1981 Seigel, 2002). Additional views are obtained in the parasagittal plane by cutting through the lateral ventricles. The intraventricular echogenic choroid plexus is well evaluated in this plane. The temporal lobe and callothalamic groove, which separates the caudate nucleus from the thalamus, are also visualized. Progressing laterally, parasagittal images are obtained through the temporal horns of the lateral ventricles further demonstrating
the hyperechoic choroid plexus (Seigel, 2002). A final view can be made on either side through the lateral thalamus.

Fig. 2. (a-d). Sagittal images of a normal neonatal brain. 3 (third ventricle) mi (mass intermedia) p (belly of the pons) 4 (fourth ventricle) cv (cerebellar vermis) c (caudate nucleus) t (thalamus)

Interpretation of cranial gray-scale imaging relies not only on neuro-anatomical knowledge but also on recognition of the normal echogenic patterns of cranial structures. The following concepts are important to keep in mind when examining cranial sonographic images. Gray matter is normally hypoechoic while white matter is hyperechoic. Therefore, an anomaly is indicated when the appearance of the two is switched (North & Lowe, 2009). Secondly, though the normal brain is always symmetric, symmetry is not always normal. A prime example is seen in bilaterally symmetric hyperechoic thalami which may signal thalamic edema, ischemia, or infarct (Huang & Castillo, 2008; North & Lowe, 2009). Thirdly, it is important to visualize the pia mater, cortical gray matter, and white matter to detect focal hemorrhages and infarcts (Grant et. al. 1981). From superficial to deep, a hyperechoic, hypoechoic, hyperechoic pattern should be seen in these layers, respectively (Figure 3) (Slovis & Kuhns, 1981). Finally, periventricular white matter should look homogenous with
an echogenicity less than or equal to the neighboring choroid plexus (North & Lowe, 2009; Seigel, 2002). A change from this appearance including asymmetry, heterogeneity, or hyperechogenicity greater than that of adjacent choroid could be indicative of periventricular leukomalacia (Figure 4).

Fig. 3. High resolution linear image of the cerebral cortex. Note the hyperechoic pia matter, hypoechoic gray matter, and hyperechoic white matter. This image displays normal gray-white matter differentiation in a term neonate.

2.2 Doppler imaging

The next step in pediatric cranial sonography is Doppler screening of the vasculature (North & Lowe, 2009). First, the Circle of Willis is investigated with color Doppler imaging using either the anterior or temporal fontanels. Spectral tracings of the internal cerebral arteries are obtained and used to calculate the peak systolic velocity (PSV), end diastolic velocity (EDV), and resistive index (RI). Color Doppler images are then performed in the sagittal plane to evaluate the sagittal sinus and vein of Galen. Power Doppler imaging can be utilized to detect signs of ischemia which may present as areas of hyper- or hypovascularity (Epelman et al., 2010).

Doppler imaging, whether color, spectral, or power, can be achieved in either the anterior or temporal fontanels according to ease or vessels to be examined (Figure 5a &b) (Lowe & Bulas, 2005). While either fontanel enables the inspection of the circle of Willis, use of the anterior fontanel better displays the internal cerebral arteries (Figure 5c) (Rumack et al. 2005; Seigel, 2002). Color Doppler imaging is used to screen the vasculature for patency and resistance to flow. Arteries are usually evaluated in the coronal plane, while venous structures are best imaged in sagittal. Indices, including the resistive index (RI), systolic, and diastolic velocities, typically of the middle or internal cerebral arteries are obtained from spectral tracings. The RI is calculated from the peak systolic velocity and end diastolic velocity by the following formula:

\[ RI = \frac{PSV - EDV}{PSV} \]
Fig. 4. (a) Normal echogenicity in the periventricular white matter is symmetric and homogenous. It is less echogenic than the neighboring choroid. (b and c) Periventricular leukomalacia is asymmetric, heterogeneous, and more echogenic than the adjacent choroid plexus.

The RI may be impacted by flow velocity, blood volume, congenital cardiac anomalies, and peripheral vascular resistance (Bulas & Vezina, 1999; Soetaert et. al., 2009). The RI tends to decline with age as diastolic flow increases with the progression from fetal to newborn circulation. Premature infants exhibit a normal RI of .62-.92 that typically drops to .43-.58 by the time the child is older than 2 years (Allison et. al., 2000; Lowe & Bulas, 2005). Therefore, a value ranging from .6-.9 is used to estimate a normal RI in premature and full-term infants. Lower values may indicate acute hypoxia or ischemia, which may trigger increased diastolic flow through cerebral vasodilation. Higher values may suggest cerebral swelling where intracranial pressures rise higher than systemic pressures leading to decreased diastolic flow (North & Lowe, 2009).
As mentioned above, the RI is not always reliable. This is especially true in children with extra cardiac shunts or congenital cardiac malformations with left to right intracardiac shunt, particularly a patent ductus arteriosus (Lipman et. al., 1982). In addition, vessel velocity fluctuations are common leading to the need to acquire multiple measurements.

2.3 Additional fontanels and linear imaging
A state of the art approach to head sonography requires utilization of additional fontanels and high-resolution linear imaging. The mastoid fontanels enable inspection of the cerebellar hemispheres and aid in detection of posterior fossa hemorrhages and congenital anomalies (Figure 6) (Di Salvo, 2001). Mastoid fontanels are also used to evaluate the size and appearance of the fourth ventricle and cisterna magna. Due to improved posterior fossa hemorrhage discovery, these views are now part of the standard of care for hindbrain
imaging (Seigel, 2002). The transverse sinuses can be examined through posterior fontanels or foramen magnum. Color Doppler can aid in evaluating vascular flow and patency (Brennan et. al., 2005; Epelman et. al, 2010; Lowe & Bulas, 2005, Sudakoff et. al., 1993). Linear imaging should be used to assess the subarachnoid space and superficial cortex via the anterior fontanel (Figure 7) (Epelman et. al., 2010). Supplementary fontanels may be explored to gain accessory views.

Fig. 6. Normal posterior fossa image obtained using the mastoid fontanel. cb (cerebellar hemispheres) m (midbrain)

Fig. 7. High resolution linear image demonstrating prominent extra axial CSF spaces. Doppler interrogation demonstrates a vessel crossing the space, confirming subarachnoid fluid rather than subdural.
3. Pitfalls and normal variants

3.1 Immature sulcation in premature infants
The infant brain becomes increasingly sulcated during development. Cerebral cortices in neonates born prior to the 24th week gestational age contain only Sylvian fissures in an otherwise smooth cerebral cortex making lissencephaly in this age group an avoidable diagnosis (Figure 8). At 24 weeks gestational age, the parietooccipital fissure is seen. This is followed by the appearance of the cingulate sulci at 28 weeks gestational age with additional branching occurring into full term (North & Lowe, 2009; Rumack et. al., 2005).

![Figure 8. Coronal image of a 24 week gestation preterm infant. Note the lack of cerebral sulci. Only the Sylvian Fissures are clearly visible.](image)

3.2 Persistent fetal fluid-filled spaces
Healthy infants commonly exhibit persistent fetal fluid-filled spaces. Typical findings consist of cavum septum pellucidum (CSP), cavum vergae, and cavum veli interpositi. Usually, these spaces start to close around the 6th month of gestational age, and most (85%) are completely closed by 3-6 months of chronological age, although some may remain present into adulthood (Epelman, 2006; Faruggia & Babcock, 1981; Needleman et. al., 2007; Osborn, 1991). The CSP is a midline fluid-filled space located anteriorly and between the frontal horns of the lateral ventricles (Figure 9). It is formed from failed septal laminae fusion (Epelman, 2006). If the space extends posterior to the fornices, it is termed cavum vergae. (Bronshtein & Weiner, 1992; Epelman et. al., 2006). A cavum veli interpositi is located even further posteriorly as a separate fluid-filled space found in the pineal region. Due to the location of the cavum veli interpositi. It must be distinguished from other commonly occurring manifestations in this region, specifically, congenital pineal cysts and vein of Galen malformations (Chen et. al., 1998; Epelman et. al., 2006).

3.3 Mega cisterna magna
A mega cisterna magna is defined as a cisterna magna that measures greater than 8mm in either sagittal or axial planes. It is present in approximately 1% of infant brains and is a benign finding of no clinical significance. It is, therefore, key not to mistake a mega cisterna magna for other malformations that can occur in the posterior fossa (Bernard et. al., 2001; Epelman et.al., 2006; Nelson et. al., 2004). Unlike arachnoid cysts, a mega cisterna magna will not exhibit mass effect on adjacent structures. It can also be differentiated from Dandy-
Walker malformations by visualization of the cerebellar vermis (Epelman et al., 2006; Nelson et al., 2004).

Fig. 9. Coronal image of a term neonate demonstrating a midline fluid-filled space between the frontal horns of the lateral ventricles consistent with a cavum septum pellucidum (csp).

3.4 Asymmetric ventricular size
20-40% of infants may exhibit asymmetrical ventricular size (Middleton et al., 2009; Seigel, 2002; Winchester et al., 1986). This can appear as a discrepancy between the frontal and occipital horn size termed colpocephaly or as variations in sizes of the left and right ventricles. The former may co-occur with agenesis of the corpus callosum or Chiari II malformations (Achiron et al., 1997; Horbar et al., 1983; Seigel, 2002). The latter is a common finding in which the less dependent ventricle is the largest. The left ventricle is most commonly the bigger of the two with sizes varying depending on patient position (Enriquez et al., 2003; Koeda et al., 1988; Seigel, 2002).

3.5 Choroid plexus variants
The choroid plexus is located at the roof of the third ventricle and travels through the foramen of Monro into the lateral ventricles (North & Lowe, 2009; Seigel, 2002). It does not extend into the frontal or occipital horns, so hyperechoic material in these areas should suggest pathology. Choroid appearing substance anterior to the caudothalamic groove is likely a germinal matrix hemorrhage, while interventricular hemorrhage may be indicated by hyperechoic material in the dependent portions of the occipital horns (Grant et al., 1983; Seigel, 2002). Choroid plexus morphology, however, is variable and should not be confused with hemorrhage. This is especially true in the glomus of the lateral ventricles and ventricular atria where lobular and bulbous choroid is commonly seen (Figure 10). The occipital horns may also display bulbous and drumstick-shaped choroid variations, particularly in conjunction with Chiari II malformations (Di Salvo, 2001; Seigel, 2002).

In addition, choroid cysts are commonly identified (Figure 11). When seen in isolation and measuring less than 1cm, they are considered a benign finding (DeRoo et al., 1988; Epelman et al., 2006; Ostlere et al., 1990; van Baalen & Versmold, 2004). When choroid cysts are found to be multiple, bilateral, or greater than 1 cm in size, this may suggest a chromosomal abnormality (Gupta et al., 1995; Seigel, 2002).
Fig. 10. Sagittal images of the (a) right and (b) left lateral ventricles demonstrating asymmetry in the choroid plexus (arrows) with the right being larger and more lobulated than the left. Both are normal.

Fig. 11. Coronal (a) and Sagittal (b) images demonstrate an anechoic, cystic structure (arrows) arising from the anterior choroid consistent with a choroid plexus cyst.

3.6 Periventricular cystic lesions

Connatal cysts, subependymal cysts, germinolytic cysts, and white matter cysts can all appear in the periventricular area (Epelman et. al., 2006; North & Lowe, 2009). Connatal cysts are always found in immediate proximity with the frontal horns where they are found in multiples and exhibit a string of pearls appearance (Figure 12). They are a normal variant believed to be caused by deficient coaptation of the ventricles (Chang et. al., 2006; Enriquez et. al., 2003; Epelman et. al, 2006; Pal et. al., 2001; Rosenfeld et. al., 1997). These cysts are often discovered shortly after birth and usually undergo spontaneous regression (Chang et. al., 2006; Rosenfeld et. al., 1997). Subependymal cysts and germinolytic cysts both occur at
the caudothalamic groove. Subependymal cysts are the sequela of germinal matrix hemorrhages and geminolytic cysts are commonly seen in metabolic disorders. Despite the differing pathophysiology, the imaging characteristics are identical and the two cysts cannot be distinguished from one another. (Epelman et. al., 2006; Herini et. al., 2003). Lastly, hypoxia and ischemic injury in preterm infants can lead to white matter cysts located near the lateral ventricles in cases of cystic periventricular leukomalacia (Figure 13) (Epelman et. al., 2006; North & Lowe, 2009).

Fig. 12. Cystic lesions in the bilateral frontal periventricular white matter (arrows) consistent with connatal cysts. Connatal cysts are considered to be a normal variant.

Fig. 13. (a) Coronal and (b) Sagittal images demonstrating multiple cystic lesions in the posterior periventricular white matter consistent with cystic periventricular leukomalacia.
3.7 Periventricular halos
Periventricular halos are hyperechoic white matter pseudolesions found adjacent to ventricles (Figure 14). They are discovered more often in preterm than term infants and are caused by anisotropic effect. These artifacts tend to disappear on tangential imaging and should have less echogenicity than neighboring choroid plexus (Enriquez et. al., 2003; Seigel, 2002). Linear imaging may serve a complimentary role in evaluation, helping distinguish between normal and abnormal processes. Pathological findings such as periventricular hemorrhage, leukomalacia, or other abnormalities, are seen in two planes (DiPietro et. al., 1986; Schafer et. al., 2009).

Fig. 14. Coronal image in a premature infant (note the lack of sulci) demonstrating hyperechoic pseudolesions in the periventricular white matter consistent with normal “periventricular halo”. This should not be confused with the more echogenic, less homogeneous findings of periventricular leukomalacia.

3.8 Lenticulostriate vasculopathy
Lenticulostriate vasculopathy appears as linear, branching or punctate increased echogenicity within the thalami (Figure 15). It can be unilateral or bilateral (Cabanas et. al., 1994; Coley et. al., 2000; Makhoul et. al., 2003). It is associated with congenital infections (TORCH infections), metabolic syndromes (peroxisomal biogenesis disorders), severe congenital heart disease, and chromosomal anomalies (Patau’s syndrome) which are all believed to cause lenticulostriate artery wall thickening (Cabanas et. al. 1994; Leijser et. al., 2007; Makhoul et. al., 2003; Te Pas et. al., 2005; Teele et. al., 1988). Sometimes, however, no specific cause can be ascertained making lenticulostriate vasculopathy a non-specific finding on sonography(Cabanas et. al., 1994; Coley et. al., 2000; Makhoul et. al., 2003).
4. Conclusion

Technical advances in ultrasound examination have made this a highly sensitive means to detect and guide treatment for pediatric cranial pathologies. When performing a head ultrasound, it is important to use modern modalities to full advantage. Doppler and linear sonography as well as the use of additional fontanels play a key role in the evaluation of the pediatric brain. Further knowledge of normal anatomy, normal variants, and imaging pitfalls increases diagnostic accuracy and helps to limit misdiagnoses.

5. References


Grant EG, Schellinger D, Richardson JD, Coffey ML, Smirniotopoulous JG. Echogenic periventricular halo: normal sonographic finding or neonatal cerebral hemorrhage. *AJR* 1983; 140:793–796


Winchester P, Brill PW, Cooper R, Krauss AN, Peterson HD. Prevalence of “compressed” and asymmetric lateral ventricles in healthy full-term neonates: sonographic study. AJR 1986; 146
Neuroimaging methodologies continue to develop at a remarkable rate, providing ever more sophisticated techniques for investigating brain structure and function. The scope of this book is not to provide a comprehensive overview of methods and applications but to provide a 'snapshot' of current approaches using well-established and newly emerging techniques. Taken together, these chapters provide a broad sense of how the limits of what is achievable with neuroimaging methods are being stretched.

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