

A Functional Role for Microglia in Epilepsy

Martine M. Mirrione^{1,2,3} and Stella E. Tsirka¹

¹*Department of Pharmacological Sciences, Molecular and Cellular Pharmacology, Stony Brook University, Stony Brook, New York*

²*Medical Department, Brookhaven National Laboratory, Upton, New York*

³*Cold Spring Harbor Laboratory, Cold Spring Harbor, New York
USA*

1. Introduction

Microglia are the immune competent cells of the CNS and comprise the major mechanism of self-defense against brain injury, infections and disease. Activation of microglia occur as a response to these insults, and both neurotoxic and neuroprotective factors can be released (Streit et al., 1999, Streit, 2002, Schwartz, 2003, Schwartz et al., 2006). There is a great deal of evidence suggesting that microglia have a role in neurodegenerative diseases, either to promote the pathology, or to counter it. However in the case of epilepsy, specific questions still remain including how, why, and when microglia are activated. In this first section, we will provide a general introduction to the role of activated microglia in the central nervous system (CNS). In subsequent sections, we will specifically discuss evidence of a functional role for microglia in epilepsy.

In the normal brain, the majority of microglia are in the resting, or quiescent ramified state. This shape with their long processes, allows them to quickly assess and respond to CNS injury or pathogens. During excitotoxic insult or inflammation, microglia become activated. This activation is manifested morphologically by retraction of their processes and changing to a rounded amoeboid morphology. Microglia also become proliferative and migratory. Their electrophysiological characteristics are altered and the expression of potassium, proton, sodium, calcium and chloride currents changes (Eder, 1998, Ducharme et al., 2007, Averaimo et al., 2010, Skaper, 2011). Once activated, the primary function of microglia is to return injured tissue to homeostasis (Streit and Xue, 2009), but this is a double edged sword, as their presence can be both 'good' and 'bad' for neurons. These effects are complex and overlapping, and not necessarily mutually exclusive.

In an effort to protect surviving cells during pathological conditions, microglia have been shown to 'execute' damaged or dying neurons injured from excitotoxicity in order to protect nearby cells from lytic release of toxic intracellular contents. Thus, microglia will facilitate local tissue repair by phagocytosing these cells and cell debris. This occurs through initial mobilization of cells near the site of injury and recruitment of distant microglia into the damaged area, which release proinflammatory mediators. At appropriately minimal and transient microglial activation, this process is ultimately neuroprotective (Vilhardt, 2005). However unregulated hyperactivation and release of toxic factors, such as nitric oxide (NO),

leading to over production of excess peroxynitrite, and reactive oxygen and nitrogen species (ROS, RNS), may result in an unmanageable level of oxidative stress causing degeneration in nearby 'bystander' cells. Oxidative stress can cause further neuroinflammation as well by recruitment of peripheral immune cells into the damaged brain. This could occur through a compromised blood-brain-barrier (BBB) as can be the case in stroke, Alzheimer's disease, amyotrophic lateral sclerosis, and epilepsy (del Zoppo et al., 2000, Mhatre et al., 2004, van Vliet et al., 2007).

Microglia can also be modulatory by secreting a host of inflammatory mediators upon activation, including cyclooxygenase-2 (Cox-2), interleukin-3 (IL-3), interleukin-6 (IL-6), interleukin-1beta (IL-1 β), tumor necrosis factor-alpha (TNF- α), prostaglandins (PGs), tissue plasminogen activator (tPA), monocyte chemoattractant protein-1 (MCP-1), vascular endothelial growth factor (VEGF), lymph toxin, matrix metalloproteinases (MMPs), and macrophage inflammatory protein-1alpha (MIP-1 α). The identity, quantity and duration of release of these factors can vary widely based on the specific injury. Some of these molecules can be toxic in high amounts, and certainly can have a direct impact on neuronal function. For example, we have studied the protease tissue plasminogen activator (tPA), which along with other trophic factors released by microglia, has been demonstrated to be critical in the sprouting of mossy fibers emanating from the dentate gyrus (DG) (Wu et al., 2000, Ferrer, 2002, Zhang et al., 2004) and may potentially facilitate seizure recruitment and the chronic maintenance of convulsions (Sloviter et al., 1996, Buckmaster et al., 2002, Shibley and Smith, 2002, Winokur et al., 2004). Indeed a number of studies including our own, have suggested that tPA from neurons, and potentially also microglia, plays a role in mediating seizure development (Qian et al., 1993, Schmoll et al., 2003, Yepes and Lawrence, 2004, Pawlak et al., 2005, Mirrione et al., 2007).

There is as well, a good side to this double-edged 'microglia' sword. They can also release several neuroprotective factors, such as neurotrophins including brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF), which can in fact promote neuronal stability (Elkabes et al., 1996, Miwa et al., 1997, Elkabes et al., 1998, Heese et al., 1998, Nakajima et al., 2001a). Maximizing this neuroprotective function can become particularly important in the aging brain as microglia cells enter a phase of senescence, and aged neurons require greater neuroprotection (Streit et al., 2008). Microglia can also positively influence pathological conditions by facilitating the recovery of injured neurons through release of trophic factors as well as extracellular matrix molecules, such as thrombospondin, important for sustaining neuronal function (Chamak et al., 1994). Additionally, application of microglia to the site of spinal cord injury has been shown to improve regenerative neurite outgrowth (Rabchevsky and Streit, 1997).

During CNS injury and glutamate excitotoxicity, microglia can act as both antigen presenting cells (Neumann et al., 1996, O'Keefe et al., 2001), and as cells that can remove harmful materials, including glutamate. Activated microglia express the high affinity glutamate transporter GLT-1, and thus can contribute approximately 10% of glutamate recycling (Nakajima et al., 2001b, Shaked et al., 2005, Persson et al., 2006), which may become more important under pathological conditions especially if astrocytes are overburdened or impaired. In addition, the cerebellum bergman glia have been shown to be involved in tonic GABA release through the ion channel Best1 (Lee et al., 2010), which may also be expressed and function in microglia in other brain regions (Ducharme et al., 2007).

While this remains to be explored, tonic GABA release could also be a mechanism for microglia to modulate seizures.

Other ways in which microglia may be beneficial involves the emerging notion of protective autoimmunity (Shaked et al., 2005). During CNS injury such as glutamate toxicity, a proposed mechanism to minimize the destruction would involve building a tolerance to autoimmune self-antigens, and could involve priming or preconditioning of microglia cells (Schwartz et al., 2003). This may be accomplished in several ways, but one experimental way is to expose microglia to lipopolysaccharide (LPS) prior to the injury or pathology. There are a few examples in the literature where preconditioning microglia with LPS modulates seizures including our own, and this will be discussed in more detail in the next section (Sayyah et al., 2003, Akarsu et al., 2006, Arican et al., 2006, Dmowska et al., 2010, Mirrione et al., 2010, Yang et al., 2010a). One mechanism that may contribute to the positive effects attributed to LPS preconditioning is through enhanced neurotrophin release (Nakajima et al., 2001a). While activated rat microglia have indeed been shown to release BDNF, this release is significantly enhanced when they are stimulated with LPS. Furthermore, these activated microglia also released NGF, suggesting that in the context of LPS preconditioning, microglia could be neuroprotective through release of neurotrophins. A careful balance needs to be maintained in dosage and timing of LPS application, as of course it also activates release of potentially toxic factors. Therefore, new strategies to precondition microglia in more refined ways to promote neuroprotective functions, and minimize neurotoxic factor release, may prove to be therapeutically beneficial. In fact one study showed that application of ceramide increased the release of protective factors, but not other potentially dangerous molecules, through specific signaling mechanisms involving protein kinase C (PKC) (Nakajima et al., 2002). As well, interferon-gamma (INF-gamma) administration causes microglia to significantly increase their ability to remove glutamate, also without causing an inflammatory response (Shaked et al., 2005). These strategies should be explored further in the context of seizures and potentially exploited as neuroprotective modulators.

2. A functional role for microglia in seizures: The ‘bad’ and the ‘good’

A growing body of evidence suggests that microglia have a functional role in the pathology and symptoms of CNS diseases including ischemia and Alzheimer’s disease. In this section, we discuss the evidence linking microglia to seizures and epilepsy. We will focus on examples from the literature suggesting both ‘bad’ neurotoxic, and ‘good’ neuroprotective, contributions. However, while it is clear from these data that a functional role for microglia exists, these effects are certainly complex and even contradictory in different literature reports. Are microglia activated concurrent with seizure pathology, and if so what is the timing, duration, location and extent of this activation? There is growing evidence suggesting that yes, extensive microglial activation known as ‘microgliosis’ occurs in the brain parenchyma of patients with recurrent seizure episodes (as well as reactive astrogliosis), and in animal models of epilepsy particularly in the hippocampus (Beach et al., 1995, Drage et al., 2002). Activated microglia are observed in the same hippocampal regions associated with seizure induced neuronal death clinically and in animal models, owing them a reputation for facilitating neuronal malfunction (Beach et al., 1995, Taniwaki et al., 1996, Tooyama et al., 2002, Borges et al., 2003). In patients, microglia activation was found in the sclerotic hippocampus, suggesting that neuronal degeneration continues to occur as a result of

ongoing seizure activity (Beach et al., 1995). A recent study confirmed the presence both of activated microglial and also immunoreactive leukocytes in tissue resected from patients with intractable medial temporal lobe epilepsy, and in kainic acid (KA) treated mice, either associated with blood vessels or distributed intraparenchymally in the CA1-CA3, hilus, and to a lesser extent, dentate gyrus (Zattoni et al., 2011). Following pilocarpine induced seizures in mice, microgliosis persists for at least 3-31 days in regions of neuronal loss such as the hippocampus and amygdala (Borges et al., 2003, Yang et al., 2010a). However, we have shown that during KA excitotoxicity, microglia activation is associated with tPA release from injured neurons, and thus is a consequence rather than a cause of neurodegeneration, but overall this microglia activation can further exacerbate the injury (Siao et al., 2003). Gliomas have also been associated with epilepsy, and histological examination of patients' tissue following surgical removal has demonstrated that the numbers of activated microglia correlated with the duration of epilepsy, as well as with the frequency of seizures prior to surgical resection (Aronica et al., 2005). Overall, this evidence would suggest that microglia activation is 'bad' for the epileptic brain and is associated with neurodegenerative pathology.

Knowing that microglial activation is associated with seizures and epilepsy is the first step, but what exactly are these cells doing? Given the plethora of molecules microglia release, one can easily imagine numerous potential outcomes. For instance, it has been proposed that microglia activation may contribute to spontaneous recurrent seizures (SRS) by facilitating aberrant migration of newborn neurons in the DG (Yang et al., 2010a). This study showed that LPS injected directly into the DG promoted the development of ectopic hilar basal dendrites in the hippocampus, while addition of minocycline that blocked microglia activation, prevented it. Neurotrophin and cytokine release can contribute to this aberrant granule cell neurogenesis (Scharfman, 2005). The expression of pro-inflammatory transcripts changes after pilocarpine-induced seizures in mice, and includes upregulation of toll-like receptor type 2 (TLR2, a microglia/macrophage marker) and I kappa B alpha (IkB α , index of NF-kappa B activation), particularly in areas undergoing neurodegeneration or demyelination (Turrin and Rivest, 2004). Increased microglial activation is also found in adult animals given KA when they have already been exposed to it during adolescence (Somera-Molina et al., 2009). In this model, early-life seizures created an increased susceptibility to seizures later in life ('two-hit seizure' model), when KA is given at postnatal day 5 (P5), and then again at P45. The expression of proinflammatory cytokines IL-1 beta, TNF-alpha, S100B, and the chemokine CCL2 was found to be enhanced, corresponding to increased susceptibility to seizures in the adults.

Recent evidence strengthens a strong implication of cytokine involvement in modulating acute seizures. mRNA expression of TNF-alpha and IL-6 correlated with seizure development in a viral infection model, and knockout mice lacking the receptors for these cytokines showed reduced seizure frequency (Kirkman et al., 2010). These results suggest the innate immune response to viral infection contributes to seizures through cytokine expression, and can potentially occur through modulation of glutamate receptors/transporters on astrocytes. Such modulation can eventually lead to excessive extrasynaptic/extracellular glutamate levels, which could also be detrimental (Choi and Koh, 2008). Another study supports the role of cytokines showing that in brain regions associated with seizure damage from the nerve agent soman, there was an increased production of IL-1alpha and IL-1beta (neurotoxic cytokines by activated microglia) as well as IL-6 (neuroprotective factor released from neurons and astrocytes) at 72 hrs, in the

piriform cortex, hippocampus, and thalamus (Johnson and Kan, 2010). However in this same model, neurotoxic COX-2 expression, which mediates the production of prostaglandins, was found expressed in neurons, but not in microglia or astrocytes (Angoa-Perez et al., 2010). In the mouse kindling model, COX-1 from microglia was enhanced in the hippocampus during progression of seizures and the administration of SC-560 (a selective COX-1 inhibitor) or indomethacine (a non-selective COX inhibitor) reduced the progress of seizures (Tanaka et al., 2009). These two studies suggest that in addition to cytokines, cyclooxygenase enzymes derived from neurons and microglia may also play an important role in mediating seizures.

In terms of electrophysiological changes, one study showed that the cortical innate immune response to LPS application actually increased local neuronal excitability. Furthermore, in a subset of animals, this also produced motor seizures (Rodgers et al., 2009). Based on their experimental evidence, the authors suggest that microglial activation may therefore be a potential precursor to seizures, and *not* a consequence of them. Specifically, they showed that when LPS is applied to the cortex *in vivo*, the evoked field potential amplitudes were acutely enhanced (as measured within 5min-1hr) and produced focal epileptiform discharges, which were prevented by pre-application of an interleukin-1 receptor antagonist. They suggested that this rapid response may result in increased glutamate and noradrenaline release within 10 min, potentially causing the rapid increases of neuronal excitability (Wang and White, 1999). While this study showed that LPS induced microglial activation produces increases in neuronal excitability, they only measured short time points, and so further experiments are necessary to determine whether this effect is long lasting and can truly facilitate seizure symptoms.

Although the aforementioned data suggest a strict neurotoxic role of microglia, it must be kept in mind however, that this activation is a response to ongoing injury and abnormal circuitry rewiring. Little affirmative evidence is available suggesting that microglia are causing neuronal malfunction directly (with the exception of the acute study by Rogers et al. described above) especially since this evidence comes from rodent models where chemoconvulsants are typically used to trigger the symptoms. Therefore it is important to establish whether microgliosis is a consequence of recurrent seizure episodes or a direct early contributor to symptoms. This is inherently complex however, based on their multiple functions as described in the first section.

The most obvious way to probe these questions is to pharmacologically activate and inhibit microglial activation before, during, and after experimental seizure induction and monitor differences in acute seizure symptoms and pathology. Another way to study this is to examine seizures in knockout mice missing integral components of the inflammatory response. Zattoni et al (Zattoni et al., 2011) showed that selective pharmacological ablation of peripheral macrophages prior to kainate injection actually increased dentate granule cell degeneration. This result suggested that the F4/80 (microglia/macrophage marker) positive cells, specifically of peripheral origin, appear to be required for long-term survival of granule cells. The same investigators also showed enhanced neurodegeneration in mutant mice lacking B or T cells, also suggesting a strong impact of immune-mediated responses on network excitability. Overall, these findings support the idea that lymphocytes and macrophages infiltrating the epileptic focal area may have a neuroprotective role.

Several other groups including ours have used LPS to activate microglia at different time points prior to seizures. An interesting result that may be counterintuitive given all the studies described above, is that preconditioning microglia before seizures has shown to be

potentially protective. This is akin to reports that sublethal stress stimuli induce tolerance in ischemia (Marsh et al., 2009). In one study, LPS administration prior to pentylenetetrazole (PTZ)-induced seizures was beneficial by increasing plasma levels of NO and IL-6, which reduced blood brain barrier permeability (Arican et al., 2006). In kindling, LPS was inhibitory but only when it was administered daily for 16 days, as it blunted the acquisition of kindled behavioral seizures (Sayyah et al., 2003). Neuronal protection from cell death in hippocampal CA1, CA3 and DG was observed with LPS preconditioning 72 hours prior to seizure induction, although seizure behavior was not affected at this low dose of LPS (Dmowska et al., 2010). Importantly, one study addressed the time course of LPS delivery prior to seizure induction and demonstrated complex effects (Akarsu et al., 2006). This group showed that LPS given 4 hr before PTZ-induced seizures was pro-epileptic, but when given 18 hr before, it conferred anticonvulsant effects attributable to the expression of COX-1 and -2. It is possible that LPS stimulated release of cytokines concurrent with PTZ drives the pro-convulsant effect, but how protection is conferred by delaying seizure induction for hours to days after LPS administration is less obvious. Tolerance to excitatory input, depletion of intracellular stores of proinflammatory mediators, and upregulation of glutamate recycling in anticipation of the next insult, are all plausible explanations which need further study.

Important questions that remain include what are the circumstances that cause microglia activation during seizures specifically, and are microglia strictly necessary for seizure development? We were surprised recently in studying these questions to find that the activation state of microglia in the hippocampus has a direct impact on the sensitivity of acutely induced seizures. In our experiment, we asked whether conditional microglia/macrophage ablation could affect epileptogenesis using genetically modified mice (Mirrione et al., 2010). These mice express the herpes simplex virus thymidine kinase (HSVTK) gene under the CD11b macrophage/microglia promoter (CD11b-HSVTK+/-). As shown in the schematic of Figure 1a, activated microglia are present throughout hippocampal subfields associated with seizures. Ganciclovir (GCV) is administered intra-hippocampally through an implanted osmotic mini-pump. Over time, GCV can be taken up by cells in the hippocampus near the infusion site, which extends throughout the ipsilateral hippocampus. Subsequently, LPS and pilocarpine can be administered i.p. during the experiment. Figure 1b describes the effect of the HSVTK transgene which is under the control of the Cd11b promoter, and thus expressed exclusively in microglia and macrophages. Cd11b is expressed in resting, ramified microglial cells at low baseline levels, which results in some moderate HSVTK expression. However, cells that are activated (by LPS or pilocarpine) begin to upregulate Cd11b, and thus will express more of the HSVTK suicide gene. HSVTK by itself does not harm the cell, similar to GCV, which by itself is harmless. However, cells that contain both HSVTK and GCV will be susceptible to ablation. HSVTK will phosphorylate GCV, which is further phosphorylated by intracellular kinases. This toxic triphosphate then competes with thymine for DNA synthesis, and causes DNA replication failure killing the cell (Heppner et al., 2005). Therefore, one can selectively target microglia/macrophages for ablation as the expression of Cd11b is restricted to these cells, and increased in activated microglia. This drives increased expression of the HSVTK gene, which causes apoptosis. Microglia/macrophage cells that are in S phase of cellular division are particularly susceptible to ablation using this method, however non-dividing cells are also susceptible, albeit at reduced levels, which may be due to interference with mitochondrial DNA synthesis (Herraiz et al., 2003).

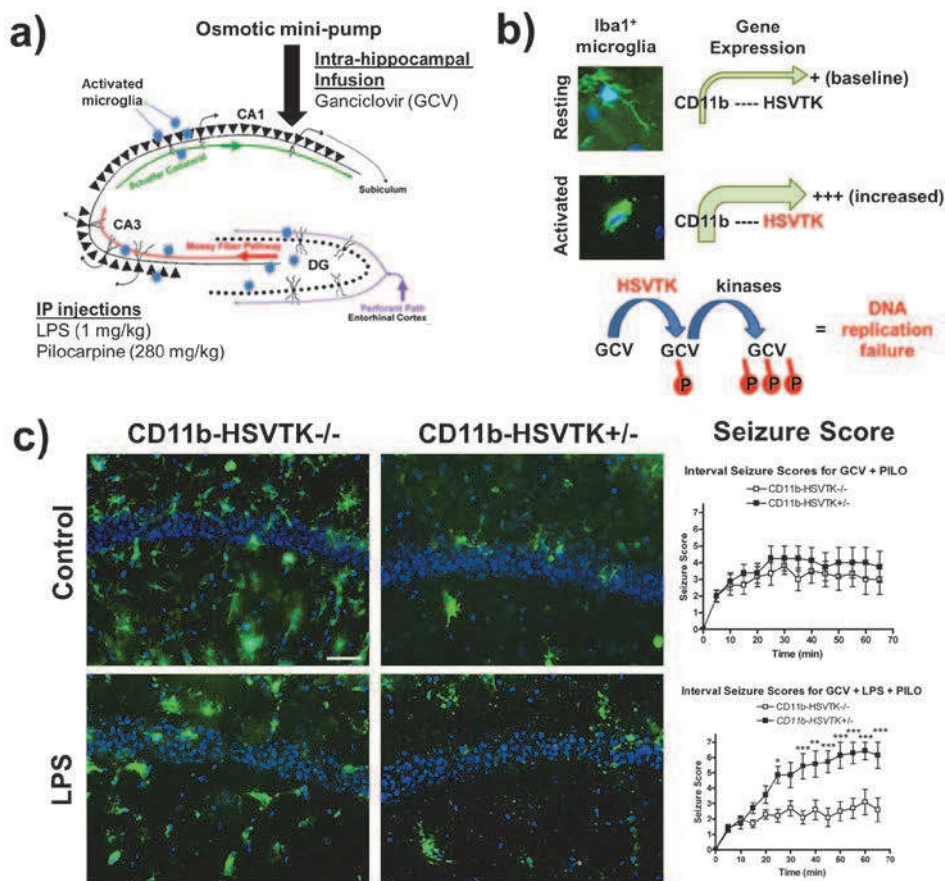


Fig. 1. Microglial ablation and lipopolysaccharide preconditioning modulates pilocarpine-induced seizures in mice. A) Activated microglia are present in hippocampal subfields following LPS and/or pilocarpine induced seizures, Neuronal circuitry in the hippocampus consists of a mainly unidirectional loop of excitatory and inhibitory signals known as the ‘trisynaptic circuit’, although additional connections between subfields also exist. These neurons are vulnerable to neurodegeneration in seizure models, and DG neurons specifically undergo circuitry rewiring propagating seizure activity. We have shown that the activation state of microglia in the hippocampus can have a direct impact on the sensitivity of acutely induced seizures. In the experiment, the hippocampus is targeted with intra-hippocampal infusion of ganciclovir (GCV) through an osmotic mini-pump. LPS and pilocarpine are given intraperitoneally (i.p.) and seizure symptoms are recorded. B) Mechanism of cell ablation via GCV and HSVTK. The morphology of resting and activated microglia are shown. The microglia/macrophage specific integrin protein, Cd11b (the alpha chain of the Mac-1 integrin), has low expression in resting cells, and is increased in activated cells. The HSVTK suicide gene is placed under the control of the promoter CD11b. When microglia are activated, CD11b expression increases and HSVTK is highly expressed. In a cell which has taken up GCV, and is expressing HSVTK, GCV is phosphorylated by

HSVTK and endogenous kinases (phosphorylation represented by red circle's labeled 'P'). This turns GCV into a toxic triphosphate which competes with endogenous thymine for DNA synthesis. Ultimately, DNA replication is disrupted and the cell undergoes apoptosis. C) Data showing CD11b-HSVTK^{-/-} (wild type, left) mice compared to CD11b-HSVTK^{-/-} (transgenic mice, right) under control and LPS preconditioning (CA1 subfield, DAPI (blue) and Iba1 (green), scale bar 50 μ m) conditions. There were significantly fewer Iba1 positive cells (microglia/macrophages) in the transgenic mice following GCV in both conditions; however seizure symptoms (right panels) only showed a significant divergence, when LPS was administered 24 hr before the pilocarpine injection. In this group, seizure scores were dramatically increased over the first 60 min following pilocarpine injection. Thus in the absence of LPS activated microglia/macrophages, seizure symptoms were significantly elevated. Additionally, LPS preconditioning in CD11b-HSVTK^{-/-} (wild type) mice also showed a trend toward reducing seizure scores. (Part C, adapted and partially reproduced from (Mirrione et al., 2010)).

Our results showed that unilateral ablation of quiescent (non-activated) microglia from the dorsal hippocampus did not necessarily alter acute seizure sensitivity (Figure 1 c, top panel), but ablation of activated microglia (by LPS given 24 hrs prior to pilocarpine) did in fact significantly enhance seizure symptoms (Figure 1c, bottom panel, and summarized in Table 1). Furthermore, we observed a trend toward reduced seizure activity in LPS preconditioned mice, similarly to what other groups have reported (Sayyah et al., 2003, Akarsu et al., 2006, Arican et al., 2006, Dmowska et al., 2010). These data would suggest that LPS activated microglia were providing some protection to the hippocampus, perhaps by controlling the spread of seizure activity. Additional studies are required to identify the chemical mediators released by microglia that may underlie this protective effect, and whether bilateral microglia/macrophage ablation would be further protective. As well, it would be interesting to further evaluate whether neurodegeneration patterns are altered at chronic time points. In regards to how LPS activated microglia upregulate Cd11b, one group showed this was mediated by nitric oxide (NO) (Roy et al., 2006, Roy et al., 2008). Therefore, LPS may be enhancing inducible nitric-oxide synthase (iNOS) in microglia, which produces NO, upregulating Cd11b, and concurrently the HSVTK gene. This would increase the percentage of cells ablated by GCV over time, potentiating the response to pilocarpine (Table 1). Furthermore, there is evidence as well that ablation of macrophages, and neutrophils which also express CD11b, may certainly play an important role, and shouldn't be discounted as potential mediators (Zattoni et al., 2011). In the future, these transgenic mice can be extremely useful for in depth examination of conditional microglial/macrophage ablation in other brain regions such as the amygdala, cortex, or more specifically targeted subfield regions of the hippocampus.

Microglia in transgenic mice (CD11b-HSVTK +/-)	Expression level		# in S phase (dividing)	GCV	Cell death	Response to PILO (compared to CD11b-HSVTK ^{-/-})
	CD11b	HSVTK				
Naïve (resting or in S phase)	+	+	+	No	no effect	n.s.
				Yes	+	n.s.
LPS Activated	+++	+++	+++	No	no effect	n.s.
				Yes	+++	potentiated

Note: plus signs (+) represent relative amounts; (PILO) pilocarpine; (n.s.) not significantly different (data from Mirrione et al., 2010)

Table 1. Expression of transgene based on activation state of microglia.

In our study, we also explored whether changes in glial function were a component of metabolic changes in these mice following pilocarpine using small animal imaging with 2-deoxy-2-[(18)F]fluoro-d-glucose (¹⁸FDG) and positron emission tomography (PET). Our goal was to find the brain regions involved in the behavioral divergence we observed in CD11b-HSVTK+/- mice, compared to wild type mice, using a systems approach. In pilocarpine treated animals of both genotypes, we found increased ¹⁸FDG uptake during seizures in the septum, thalamus, hippocampus, midbrain, and cerebellum, along with decreases in the striatum, which was consistent with our previous findings (Mirrione et al., 2007). In addition, we observed reduced metabolic activation in the ipsilateral hippocampus corresponding to the location of unilateral microglial ablation. While we interpreted this result with caution, we concluded that activated microglia may contribute a small, but potentially important, proportion of the metabolic signal. It is possible then, that changes in glial function could be a component of metabolic abnormalities which are observed in epileptic patients (Lamusuo et al., 2001, Goffin et al., 2008b) and rodent seizure models (Kornblum et al., 2000, Mirrione et al., 2006, Mirrione et al., 2007, Goffin et al., 2009). Therefore, in the next section we will discuss how neuroimaging approaches using specific radiotracers which target microglia, can facilitate our understanding of the pathology underlying epileptogenesis.

3. Neuroimaging as a potential tool for exploring *in vivo* function of microglia during epileptogenesis

We and others have taken advantage of the unique systems level, *in vivo* neuroimaging modalities to probe questions related to seizure circuitry and malfunction. These techniques have become increasingly utilized both clinically and in basic research to understand epileptogenesis. Metabolic neuroimaging with positron emission tomography (PET) is the most widely used diagnostic, (for review (Mirrione and Tsirka, 2011)). A handful of recent studies in animal models have also begun to utilize specific radioligands studying GABA, serotonin, dopamine, and the cannabinoid systems (Werhahn et al., 2006, Goffin et al., 2008a, Liefwaard et al., 2009, Liew et al., 2009, Assem-Hilger et al., 2010). There are also a few pilot studies in the literature examining the acetylcholine (Mohamed et al., 2005) and opioid systems (Theodore et al., 1992) in epilepsy patients. While the availability of these radioligands is still somewhat limited in clinical settings, and requires further validation, they may prove to be valuable tools for future research and diagnosis.

PET imaging utilizes a radiotracer, a radiolabeled molecule, that when injected intravenously into the subject will bind specifically to the location of interest. For microglia, the radiotracer should be a ligand which binds to a receptor or transporter expressed specifically on activated microglia. The ligand needs to be chosen carefully, as it would need to exhibit the necessary pharmacokinetic properties to make it a good radiotracer. Specifically, it should effectively enter the central nervous system through the blood stream by crossing the blood brain barrier (lipophilicity is expressed as log D, or log P), and it should bind to activated microglia (known as the affinity, or dissociation constant K_d) with enough strength to be injected in very low amounts, but still located by the PET camera. Other helpful properties of these radiotracers (radioligands) are that they have low non-specific binding (that they do not bind randomly to other molecules), and are they are not converted into 'active' metabolites, which can bind to some other unwanted target in the brain and perturb the analysis.

The first major radioligand that has been used to study activated microglia is carbon-11 labeled PK11195, a compound that binds to the peripheral benzodiazepine receptor (PBR). The PBR is a mitochondrial protein expressed on both microglia and macrophages. Its expression is increased after brain injury or neuroinflammation. [¹¹C]PK11195 was used in a pilot study comparing 4 normal human subjects, 3 patients with clinically stable hippocampal sclerosis and low seizure frequency, and 2 patients with histologically confirmed Rasmussen's encephalitis (RE) (Banati et al., 1999). This initial study showed that specific binding of the radiotracer the RE patients showed a focal and diffuse increase in binding throughout the seizure-affected hemisphere. Interestingly, the clinically stable hippocampal sclerosis group was similar to normal controls, suggesting that at this stage for these few patients, activated microglia were no longer contributing to the symptoms or pathology. Furthermore, a case of cerebral vasculitis in refractory epilepsy was cured with [¹¹C]PK11195 imaging (Goerres et al., 2001). This adult female patient showed an area of increased radiotracer signal in the left occipital lobe. Biopsy showed that this was positive for activated microglia, vascular and perivascular accumulations of mononuclear cells. The patient was then treated with prednisolone and azathioprine (immune suppressing drugs) which subsequently reduced her seizures.

Based on these initial studies, a recent case report using [¹¹C]PK11195 of encephalitis facilitated treatment of a 5 year old patient who did not respond to medication (Kumar et al., 2008). Using ¹⁸FDG was not particularly informative, as this patient showed diffuse glucose hypometabolism without any focal abnormality. As the physicians suspected that underlying brain inflammation was the cause, a [¹¹C]PK11195 scan was conducted which showed an area of increased uptake in the left temporal-occipital cortex. This area was subsequently resected and the patient fully recovered. They concluded that focal areas of neuroinflammation play an important role in refractory seizures associated with encephalitis, and using this imaging approach highlighted the region for surgical removal, and led to a successful recovery. This approach was used again and resulted in the treatment of another female patient with intractable epilepsy due to focal cortical dysplasia (Butler et al., 2011).

One can easily imagine from these case reports how examining the time course of microglia activation over the development and progression of epileptogenesis in patients and animal models can truly shed light on the microglial contribution to pathology. While PK1195 has proven to be a successful indicator of microglia activation in seizures, it has limited uptake in the brain and thus has limited quantitative capacity. Much effort has been put into finding alternative ligands for the PBR which have higher affinity's and better pharmacological properties as radiotracers. The list of these experimental compounds is actually quite long, and the available ligands for studying microglia were recently reviewed (Leung, 2004-2010, Schweitzer et al., 2010). One potential tracer [¹¹C]DAA1106 has been shown to have a higher affinity to the PBR compared to PK1195 and may be more suitable for measuring subtle changes in microglia activation during disease (Maeda et al., 2004). Several other tracers have similar enhanced potential including [¹²⁵I]DPA-713 (a ligand for SPECT imaging, which also has a slightly higher affinity) (Wang et al., 2009), and [¹¹C]CLINME (Boutin et al., 2007) which has lower non-specific uptake.

Thus far, the most popular target to image microglia remains as the PBR, however other targets for activated microglia should also be explored. New tracers may be significantly useful for examining the impact and progression of microgliosis during different stages of

epileptogenesis, first in animal models and eventually in patients. We urge the continued development of such radiotracers specifically for this purpose.

4. Optogenetics as a potential tool for exploring *in vivo* function of microglia during epileptogenesis

Optogenetics is an exponentially growing field in neuroscience where light responsive ion channels called opsins are genetically introduced into neurons and astrocytes (Deisseroth, 2011). Thus far, no experimental evidence is available that suggests that microglia could also be targeted with opsins, however this is certainly plausible. Channel rhodopsin-2 can activate neurons with blue light by allowing influx of sodium, but can also allow for influx of calcium, which has been exploited for activating astrocytes (Gradinaru et al., 2009, Figueiredo et al., 2011). Astrocytes in fact, could also be explored in the context of seizures using these methods. Microglia are unique however, in that once activated they express an inward chloride conductance (Averaimo et al., 2010). This could also theoretically be mimicked with expression of halorhodopsin, which when activated with yellow light, allows passage of chloride ions. These swelling-activated chloride channels I (Clswell) are naturally expressed on microglia (Ducharme et al., 2007), and contribute to their phagocytic capacity (Furtner et al., 2007), and to the production of nitric oxide (Kjaer et al., 2009). Similar to experiments on astrocytes, transgenic mouse lines can be created where these channels are expressed under microglia/macrophage specific promoters. Cre-dependent viruses could also be used to target microglia specifically. Implanted optical fibers could then deliver light stimulation to specific brain regions *in vivo* under various experimental conditions.

This method may be advantageous over pharmacological approaches, as these channels respond very quickly to light on and off by opening and closing, thus allowing for a variety of experimental manipulations *in vivo*, in real time. Potentially this could allow for fast activation and deactivation of microglia. However, the real kinetics that returns the cells to a resting state once activated is of course very different in glia than neurons. This could potentially be a limitation of this approach, since once microglia are activated, it takes them time to naturally return to a resting state. Despite this caveat, optogenetics may provide an interesting basic research avenue in the future for studying channel properties of microglia.

5. Microglia as a therapeutic target for epilepsy

Can microglia be therapeutically targeted to benefit epilepsy patients? While this question has yet to be systematically explored, a few case studies and basic research experiments may suggest a tentative yes. However, given the complexity of microglia-neuronal interactions, a systematic analysis of activated microglia function at various stages during epileptogenesis is necessary.

There are several drugs used experimentally in animal models to modulate the activation state of microglia, which are effective as anti-inflammatory agents. One recent example showed that erythropoietin administered to rats reduced a rise in inflammatory genes, edema, and the numbers of activated microglia after febrile seizures, thereby reducing the risk of subsequent spontaneous seizures (Jung et al., 2011). Another report showed that levetiracetam reduced the harmful spread of excitation during seizures by restoring gap junctions in astrocytes co-cultured with microglia (Stienen et al., 2011). The investigators

found that TGFbeta1 expression was increased, which normalized the impaired astrocyte membrane resting potentials via modification of inward and outward rectifier currents. TGFbeta1 has also been shown to be a potent anti-inflammatory and neuroprotective agent by either preventing the classical activation of microglia, or by enhancing the presence of alternatively activated microglia, and reducing oxidative stress (Qian et al., 2008).

Inflammatory blockade has also been accomplished with celecoxib, a selective cyclooxygenase-2 inhibitor (Jung et al., 2006). When given during the latent period, it prevented neuronal death and microglia activation in the hilus and CA1. It also inhibited ectopic granule cell formation in the hilus and new glia infiltrating the CA1. Interestingly, COX-2 was shown to be neuroprotective as SC-58236, another selective COX-2 inhibitor, reversed the anti-seizure activity of LPS given 18 hr before PTZ (Akarsu et al., 2006). In the same study, a COX-1 selective inhibitor, valeryl salicylate, facilitated PTZ-induced seizures. However, another group showed that in the mouse kindling model, the administration of SC-560 (a selective COX-1 inhibitor) or indomethacine (a non-selective COX inhibitor) actually reduced the progress of seizures (Tanaka et al., 2009).

Carbamazepine was also anti-inflammatory as it inhibited the production of NO and prostaglandin E2 production (Matoth et al., 2000). Resveratrol, a naturally occurring nonflavonoid polyphenol found in grapes, has been shown to inhibit microglia activation, proinflammatory factor release, and NADPH oxidase which protected neurons (Zhang et al., 2010a, Zhang et al., 2010b). Ghrelin attenuates KA induced cell death in the hippocampus of mice by preventing the activation of microglia and astrocytes and their expression of cytokines TNF-alpha, IL-1b, and COX-2 (Lee et al., 2010a). Minoxac, another small molecule that reduces proinflammatory cytokine production, was given following early life seizure induction. Minoxac attenuated the enhanced microglial responses that increased vulnerability to seizures in adulthood (Somera-Molina et al., 2009). Similarly, naloxone is also anti-inflammatory, reducing IL-1beta synthesis and microglial activation following early life seizure induction. It also lowered the vulnerability of immature brains to a second seizure episode in adulthood (Yang et al., 2010b).

In terms of cytokines and inflammatory mediators, IL-1 beta has been shown to be proconvulsant, and IL-1 receptor antagonism (via IL-1Ra) has proven to be a powerful inhibitor of seizures (Vezzani et al., 1999, Vezzani et al., 2000). A recent study reported findings that supported this notion, as IL-1 receptor antagonism blocked the effects of LPS induced enhancement of evoked field potential amplitudes and focal epileptiform discharges in the first minutes to hour of exposure (Rodgers et al., 2009). However, in this case, further study at longer time points should be evaluated to determine if this effect is lasting, or whether it changes, as we and others have shown is the case for LPS on seizure symptoms. Other inflammatory mediators, interleukin-18 (IL-18) and interferon-gamma (INF-gamma) have also proven to be protective against neuronal damage following status epilepticus (SE) (Ryu et al., 2010). Valproic acid, a standard clinical therapy for seizures, was shown to inhibit the production of NF-kappaB, which blocked the release of TNF-alpha and IL-6 by microglia (Ichiyama et al., 2000). It has also been shown to cause microglial apoptosis through the p38 MAPK and mitochondrial apoptosis pathways (Xie et al., 2010).

A recent review highlights the potential for targeting ion channels on microglia to modulate microglial activity (Skaper, 2011). For example, chloride intracellular channel 1 translocation to the activated plasma membrane generates ROS which mediates amyloid beta-peptide

clearance (Milton et al., 2008, Paradisi et al., 2008). Also, inhibiting the Ca²⁺/calmoduline-activated K⁺ channel expressed on microglia actually reduced LPS induced toxicity (Li et al., 2008).

Several studies have been published suggesting positive regulation of microglia can also have beneficial effects for seizure therapy. An intriguing concept is to activate or 'program' microglia with certain drugs to harness their neuroprotective capabilities. Ceramide is one such compound, that when applied in culture, increased the release of protective BDNF, but not other dangerous molecules, TNF-alpha, IL-1beta, or NO (Nakajima et al., 2002). Interferon-gamma (INF-gamma) has been shown to drive microglia to significantly increase their ability to remove glutamate, without causing an inflammatory response (Shaked et al., 2005). Also, insulin-like growth factor-1 (IGF-1) from microglia and astrocytes was shown to promote hippocampal CA1 neuronal survival following injury to dentate granule cells (Wine et al., 2009).

While there is overwhelming evidence that microglia should be inhibited as a means to treat epilepsy, it is important to keep in mind the context of this experimental evidence as it mostly originates from chemoconvulsant models and *in vitro* measurements. *In-vivo* situations need to be further explored to fully validate these hypotheses and potential therapeutic approaches. Overall however, the strategies outlined in this section should be assessed in the context of seizures. Of particular interest clinically, is the concept of pharmacologically harnessing the neuroprotective qualities of microglia, while inhibiting their neurotoxic properties.

6. Summary

Overall, a growing body of evidence suggests that microglia have a functional role in the pathology and symptoms of epilepsy. Future studies should focus on elucidating the time course of microglial activation during seizure progression, as well as specific factors released at these various times. Furthermore, novel approaches such as *in vivo* neuroimaging and optogenetics will provide additional avenues to examine these questions. Fully exploiting the therapeutic benefit of modulating microglia for patients with epilepsy has great potential.

7. Acknowledgements

This work was supported by the National Institutes of Health (NIH) with funding to S.E.T. (R01NS042168). The authors are grateful to Frank L. Heppner and Adriano Aguzzi for collaboration and sharing the CD11b-HSVTK transgenic mice. We would also like to thank Dr Fritz Henn for support to M.M.M while writing this manuscript.

8. References

- Akarsu ES, Ozdayi S, Algan E, Ulupinar F (2006) The neuronal excitability time-dependently changes after lipopolysaccharide administration in mice: possible role of cyclooxygenase-2 induction. *Epilepsy Res* 71:181-187.
- Angoa-Perez M, Kreipke CW, Thomas DM, Van Shura KE, Lyman M, McDonough JH, Kuhn DM (2010) Soman increases neuronal COX-2 levels: possible link between seizures and protracted neuronal damage. *Neurotoxicology* 31:738-746.

- Arıcan N, Kaya M, Kalaycı R, Uzun H, Ahışhali B, Bilgic B, Elmas I, Kucuk M, Gurses C, Uzun M (2006) Effects of lipopolysaccharide on blood-brain barrier permeability during pentylentetrazole-induced epileptic seizures in rats. *Life Sci* 79:1-7.
- Aronica E, Gorter JA, Redeker S, Ramkema M, Spliet WG, van Rijen PC, Leenstra S, Troost D (2005) Distribution, characterization and clinical significance of microglia in glioneuronal tumours from patients with chronic intractable epilepsy. *Neuropathol Appl Neurobiol* 31:280-291.
- Assem-Hilger E, Lanzenberger R, Savli M, Wadsak W, Mitterhauser M, Mien LK, Stogmann E, Baumgartner C, Kletter K, Asenbaum S (2010) Central serotonin 1A receptor binding in temporal lobe epilepsy: A [carbonyl-(11)C]WAY-100635 PET study. *Epilepsy Behav.*
- Averaimo S, Milton RH, Duchen MR, Mazzanti M (2010) Chloride intracellular channel 1 (CLIC1): Sensor and effector during oxidative stress. *FEBS Lett* 584:2076-2084.
- Banati RB, Goerres GW, Myers R, Gunn RN, Turkheimer FE, Kreutzberg GW, Brooks DJ, Jones T, Duncan JS (1999) [11C](R)-PK11195 positron emission tomography imaging of activated microglia in vivo in Rasmussen's encephalitis. *Neurology* 53:2199-2203.
- Beach TG, Woodhurst WB, MacDonald DB, Jones MW (1995) Reactive microglia in hippocampal sclerosis associated with human temporal lobe epilepsy. *Neurosci Lett* 191:27-30.
- Borges K, Gearing M, McDermott DL, Smith AB, Almonte AG, Wainer BH, Dingledine R (2003) Neuronal and glial pathological changes during epileptogenesis in the mouse pilocarpine model. *Experimental Neurology* 182:21-34.
- Boutin H, Chauveau F, Thominaux C, Kuhnast B, Gregoire MC, Jan S, Trebossen R, Dolle F, Tavitian B, Mattner F, Katsifis A (2007) In vivo imaging of brain lesions with [(11)C]CLINME, a new PET radioligand of peripheral benzodiazepine receptors. *Glia* 55:1459-1468.
- Buckmaster PS, Zhang GF, Yamawaki R (2002) Axon Sprouting in a Model of Temporal Lobe Epilepsy Creates a Predominantly Excitatory Feedback Circuit. *J Neurosci* 22:6650-6658.
- Butler T, Ichise M, Teich AF, Gerard E, Osborne J, French J, Devinsky O, Kuzniecky R, Gilliam F, Pervez F, Provenzano F, Goldsmith S, Vallabhajosula S, Stern E, Silbersweig D (2011) Imaging Inflammation in a Patient with Epilepsy Due to Focal Cortical Dysplasia. *J Neuroimaging.*
- Chamak B, Morandi V, Mallat M (1994) Brain macrophages stimulate neurite growth and regeneration by secreting thrombospondin. *J Neurosci Res* 38:221-233.
- Choi J, Koh S (2008) Role of brain inflammation in epileptogenesis. *Yonsei Med J* 49:1-18.
- Deisseroth K (2011) Optogenetics. *Nat Methods* 8:26-29.
- del Zoppo G, Ginis I, Hallenbeck JM, Iadecola C, Wang X, Feuerstein GZ (2000) Inflammation and stroke: putative role for cytokines, adhesion molecules and iNOS in brain response to ischemia. *Brain Pathol* 10:95-112.
- Dmowska M, Cybulska R, Schoenborn R, Piersiak T, Jaworska-Adamu J, Gawron A (2010) Behavioural and histological effects of preconditioning with lipopolysaccharide in epileptic rats. *Neurochem Res* 35:262-272.
- Drage MG, Holmes GL, Seyfried TN (2002) Hippocampal neurons and glia in epileptic EL mice. *J Neurocytol* 31:681-692.

- Ducharme G, Newell EW, Pinto C, Schlichter LC (2007) Small-conductance Cl⁻ channels contribute to volume regulation and phagocytosis in microglia. *Eur J Neurosci* 26:2119-2130.
- Eder C (1998) Ion channels in microglia (brain macrophages). *Am J Physiol* 275:C327-342.
- Elkabes S, DiCicco-Bloom EM, Black IB (1996) Brain microglia/macrophages express neurotrophins that selectively regulate microglial proliferation and function. *J Neurosci* 16:2508-2521.
- Elkabes S, Peng L, Black IB (1998) Lipopolysaccharide differentially regulates microglial trk receptor and neurotrophin expression. *J Neurosci Res* 54:117-122.
- Ferrer I (2002) [Cell signaling in the epileptic hippocampus]. *Rev Neurol* 34:544-550.
- Figueiredo M, Lane S, Tang F, Liu BH, Hewinson J, Marina N, Kasymov V, Souslova EA, Chudakov DM, Gourine AV, Teschemacher AG, Kasparov S (2011) Optogenetic experimentation on astrocytes. *Exp Physiol* 96:40-50.
- Furtner T, Zierler S, Kerschbaum HH (2007) Blockade of chloride channels suppresses engulfment of microspheres in the microglial cell line, BV-2. *Brain Res* 1184:1-9.
- Goerres GW, Revesz T, Duncan J, Banati RB (2001) Imaging cerebral vasculitis in refractory epilepsy using [(11)C](R)-PK11195 positron emission tomography. *AJR Am J Roentgenol* 176:1016-1018.
- Goffin K, Bormans G, Casteels C, Bosier B, Lambert DM, Grachev ID, Van Paesschen W, Van Laere K (2008a) An in vivo [18F]MK-9470 microPET study of type 1 cannabinoid receptor binding in Wistar rats after chronic administration of valproate and levetiracetam. *Neuropharmacology* 54:1103-1106.
- Goffin K, Dedeurwaerdere S, Van Laere K, Van Paesschen W (2008b) Neuronuclear assessment of patients with epilepsy. *Semin Nucl Med* 38:227-239.
- Goffin K, Van Paesschen W, Dupont P, Van Laere K (2009) Longitudinal microPET imaging of brain glucose metabolism in rat lithium-pilocarpine model of epilepsy. *Exp Neurol* 217:205-209.
- Gradinaru V, Mogri M, Thompson KR, Henderson JM, Deisseroth K (2009) Optical deconstruction of parkinsonian neural circuitry. *Science* 324:354-359.
- Heese K, Fiebich BL, Bauer J, Otten U (1998) NF-kappaB modulates lipopolysaccharide-induced microglial nerve growth factor expression. *Glia* 22:401-407.
- Heppner FL, Greter M, Marino D, Falsig J, Raivich G, Hovelmeyer N, Waisman A, Rulicke T, Prinz M, Priller J, Becher B, Aguzzi A (2005) Experimental autoimmune encephalomyelitis repressed by microglial paralysis. *Nat Med* 11:146-152.
- Herraiz M, Beraza N, Solano A, Sangro B, Montoya J, Qian C, Prieto J, Bustos M (2003) Liver failure caused by herpes simplex virus thymidine kinase plus ganciclovir therapy is associated with mitochondrial dysfunction and mitochondrial DNA depletion. *Hum Gene Ther* 14:463-472.
- Ichiyama T, Okada K, Lipton JM, Matsubara T, Hayashi T, Furukawa S (2000) Sodium valproate inhibits production of TNF-alpha and IL-6 and activation of NF-kappaB. *Brain Res* 857:246-251.
- Johnson EA, Kan RK (2010) The acute phase response and soman-induced status epilepticus: temporal, regional and cellular changes in rat brain cytokine concentrations. *J Neuroinflammation* 7:40.
- Jung KH, Chu K, Lee ST, Kim J, Sinn DI, Kim JM, Park DK, Lee JJ, Kim SU, Kim M, Lee SK, Roh JK (2006) Cyclooxygenase-2 inhibitor, celecoxib, inhibits the altered

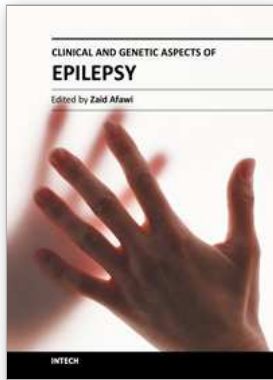
- hippocampal neurogenesis with attenuation of spontaneous recurrent seizures following pilocarpine-induced status epilepticus. *Neurobiol Dis* 23:237-246.
- Jung KH, Chu K, Lee ST, Park KI, Kim JH, Kang KM, Kim S, Jeon D, Kim M, Lee SK, Roh JK (2011) Molecular alterations underlying epileptogenesis after prolonged febrile seizure and modulation by erythropoietin. *Epilepsia* 52:541-550.
- Kirkman NJ, Libbey JE, Wilcox KS, White HS, Fujinami RS (2010) Innate but not adaptive immune responses contribute to behavioral seizures following viral infection. *Epilepsia* 51:454-464.
- Kjaer K, Strobaek D, Christophersen P, Ronn LC (2009) Chloride channel blockers inhibit iNOS expression and NO production in IFN γ -stimulated microglial BV2 cells. *Brain Res* 1281:15-24.
- Kornblum HI, Araujo DM, Annala AJ, Tatsukawa KJ, Phelps ME, Cherry SR (2000) In vivo imaging of neuronal activation and plasticity in the rat brain by high resolution positron emission tomography (microPET). *Nat Biotechnol* 18:655-660.
- Kumar A, Chugani HT, Luat A, Asano E, Sood S (2008) Epilepsy surgery in a case of encephalitis: use of 11C-PK11195 positron emission tomography. *Pediatr Neurol* 38:439-442.
- Lamusuo S, Jutila L, Ylinen A, Kalviainen R, Mervaala E, Haaparanta M, Jaaskelainen S, Partanen K, Vapalahti M, Rinne J (2001) [18F]FDG-PET reveals temporal hypometabolism in patients with temporal lobe epilepsy even when quantitative MRI and histopathological analysis show only mild hippocampal damage. *Arch Neurol* 58:933-939.
- Lee S, Yoon BE, Berglund K, Oh SJ, Park H, Shin HS, Augustine GJ, Lee CJ (2010) Channel-mediated tonic GABA release from glia. *Science* 330:790-796.
- Leung K (2004-2010) N-Benzyl-N-methyl-2-[7,8-dihydro-7-(2-[18F]fluoroethyl)-8-oxo-2-phenyl-9H-purin-9-yl]acetamide. In: *Molecular Imaging and Contrast Agent Database (MICAD)* [Internet] Bethesda (MD): National Center for Biotechnology Information (US); 2004-2010.
- Li F, Lu J, Wu CY, Kaur C, Sivakumar V, Sun J, Li S, Ling EA (2008) Expression of Kv1.2 in microglia and its putative roles in modulating production of proinflammatory cytokines and reactive oxygen species. *J Neurochem* 106:2093-2105.
- Liefaard LC, Ploeger BA, Molthoff CF, de Jong HW, Dijkstra J, van der Weerd L, Lammertsma AA, Danhof M, Voskuyl RA (2009) Changes in GABA A receptor properties in amygdala kindled animals: in vivo studies using [11C]flumazenil and positron emission tomography. *Epilepsia* 50:88-98.
- Liew CJ, Lim YM, Bonwetsch R, Shamim S, Sato S, Reeves-Tyer P, Herscovitch P, Dustin I, Bagic A, Giovacchini G, Theodore WH (2009) 18F-FCWAY and 18F-FDG PET in MRI-negative temporal lobe epilepsy. *Epilepsia* 50:234-239.
- Maeda J, Suhara T, Zhang MR, Okauchi T, Yasuno F, Ikoma Y, Inaji M, Nagai Y, Takano A, Obayashi S, Suzuki K (2004) Novel peripheral benzodiazepine receptor ligand [11C]DAA1106 for PET: an imaging tool for glial cells in the brain. *Synapse* 52:283-291.
- Marsh B, Stevens SL, Packard AE, Gopalan B, Hunter B, Leung PY, Harrington CA, Stenzel-Poore MP (2009) Systemic lipopolysaccharide protects the brain from ischemic injury by reprogramming the response of the brain to stroke: a critical role for IRF3. *J Neurosci* 29:9839-9849.

- Matoth I, Pinto F, Sicsic C, Brenner T (2000) Inhibitory effect of carbamazepine on inflammatory mediators produced by stimulated glial cells. *Neurosci Res* 38:209-212.
- Mhatre M, Floyd RA, Hensley K (2004) Oxidative stress and neuroinflammation in Alzheimer's disease and amyotrophic lateral sclerosis: common links and potential therapeutic targets. *J Alzheimers Dis* 6:147-157.
- Milton RH, Abeti R, Averaimo S, DeBiasi S, Vitellaro L, Jiang L, Curmi PM, Breit SN, Duchen MR, Mazzanti M (2008) CLIC1 function is required for beta-amyloid-induced generation of reactive oxygen species by microglia. *J Neurosci* 28:11488-11499.
- Mirrione M, Tsirka S (2011) Neuroimaging in animal seizure models with 18FDG-PET. *Epilepsy Research and Treatment*.
- Mirrione MM, Konomos DK, Gravanis I, Dewey SL, Aguzzi A, Heppner FL, Tsirka SE (2010) Microglial ablation and lipopolysaccharide preconditioning affects pilocarpine-induced seizures in mice. *Neurobiol Dis* 39:85-97.
- Mirrione MM, Schiffer WK, Fowler JS, Alexoff DL, Dewey SL, Tsirka SE (2007) A novel approach for imaging brain-behavior relationships in mice reveals unexpected metabolic patterns during seizures in the absence of tissue plasminogen activator. *Neuroimage* 38:34-42.
- Mirrione MM, Schiffer WK, Siddiq M, Dewey SL, Tsirka SE (2006) PET imaging of glucose metabolism in a mouse model of temporal lobe epilepsy. *Synapse* 59:119-121.
- Miwa T, Furukawa S, Nakajima K, Furukawa Y, Kohsaka S (1997) Lipopolysaccharide enhances synthesis of brain-derived neurotrophic factor in cultured rat microglia. *J Neurosci Res* 50:1023-1029.
- Mohamed A, Eberl S, Fulham MJ, Kassiou M, Zaman A, Henderson D, Beveridge S, Constable C, Lo SK (2005) Sequential 123I-iododexetimide scans in temporal lobe epilepsy: comparison with neuroimaging scans (MR imaging and 18F-FDG PET imaging). *Eur J Nucl Med Mol Imaging* 32:180-185.
- Nakajima K, Honda S, Tohyama Y, Imai Y, Kohsaka S, Kurihara T (2001a) Neurotrophin secretion from cultured microglia. *J Neurosci Res* 65:322-331.
- Nakajima K, Tohyama Y, Kohsaka S, Kurihara T (2001b) Ability of rat microglia to uptake extracellular glutamate. *Neurosci Lett* 307:171-174.
- Nakajima K, Tohyama Y, Kohsaka S, Kurihara T (2002) Ceramide activates microglia to enhance the production/secretion of brain-derived neurotrophic factor (BDNF) without induction of deleterious factors in vitro. *J Neurochem* 80:697-705.
- Neumann H, Boucraut J, Hahnel C, Misgeld T, Wekerle H (1996) Neuronal control of MHC class II inducibility in rat astrocytes and microglia. *Eur J Neurosci* 8:2582-2590.
- O'Keefe GM, Nguyen VT, Ping Tang LL, Benveniste EN (2001) IFN-gamma regulation of class II transactivator promoter IV in macrophages and microglia: involvement of the suppressors of cytokine signaling-1 protein. *J Immunol* 166:2260-2269.
- Paradisi S, Matteucci A, Fabrizi C, Denti MA, Abeti R, Breit SN, Malchiodi-Albedi F, Mazzanti M (2008) Blockade of chloride intracellular ion channel 1 stimulates Abeta phagocytosis. *J Neurosci Res* 86:2488-2498.
- Pawlak R, Melchor JP, Matys T, Skrzypiec AE, Strickland S (2005) Ethanol-withdrawal seizures are controlled by tissue plasminogen activator via modulation of NR2B-containing NMDA receptors. *Proc Natl Acad Sci U S A* 102:443-448.

- Persson M, Sandberg M, Hansson E, Ronnback L (2006) Microglial glutamate uptake is coupled to glutathione synthesis and glutamate release. *Eur J Neurosci* 24:1063-1070.
- Qian L, Wei SJ, Zhang D, Hu X, Xu Z, Wilson B, El-Benna J, Hong JS, Flood PM (2008) Potent anti-inflammatory and neuroprotective effects of TGF-beta1 are mediated through the inhibition of ERK and p47phox-Ser345 phosphorylation and translocation in microglia. *J Immunol* 181:660-668.
- Qian Z, Gilbert ME, Colicos MA, Kandel ER, Kuhl D (1993) Tissue-plasminogen activator is induced as an immediate-early gene during seizure, kindling and long-term potentiation. *Nature* 361:453-457.
- Rabchevsky AG, Streit WJ (1997) Grafting of cultured microglial cells into the lesioned spinal cord of adult rats enhances neurite outgrowth. *J Neurosci Res* 47:34-48.
- Rodgers KM, Hutchinson MR, Northcutt A, Maier SF, Watkins LR, Barth DS (2009) The cortical innate immune response increases local neuronal excitability leading to seizures. *Brain* 132:2478-2486.
- Roy A, Fung YK, Liu X, Pahan K (2006) Up-regulation of microglial CD11b expression by nitric oxide. *J Biol Chem* 281:14971-14980.
- Roy A, Jana A, Yatish K, Freidt MB, Fung YK, Martinson JA, Pahan K (2008) Reactive oxygen species up-regulate CD11b in microglia via nitric oxide: Implications for neurodegenerative diseases. *Free Radic Biol Med* 45:686-699.
- Sayyah M, Najafabadi IT, Beheshti S, Majzoob S (2003) Lipopolysaccharide retards development of amygdala kindling but does not affect fully-kindled seizures in rats. *Epilepsy Res* 57:175-180.
- Scharfman HE (2005) Seizure-induced neurogenesis and its dependence on growth factors and cytokines. *Growth Factors and Epilepsy Hauppauge: Nova Sciences*.
- Schmoll H, Badan I, Grecksch G, Walker L, Kessler C, Popa-Wagner A (2003) Kindling status in sprague-dawley rats induced by pentylenetetrazole: involvement of a critical development period. *Am J Pathol* 162:1027-1034.
- Schwartz M (2003) Macrophages and microglia in central nervous system injury: are they helpful or harmful? *J Cereb Blood Flow Metab* 23:385-394.
- Schwartz M, Butovsky O, Bruck W, Hanisch UK (2006) Microglial phenotype: is the commitment reversible? *Trends Neurosci* 29:68-74.
- Schwartz M, Shaked I, Fisher J, Mizrahi T, Schori H (2003) Protective autoimmunity against the enemy within: fighting glutamate toxicity. *Trends Neurosci* 26:297-302.
- Schweitzer PJ, Fallon BA, Mann JJ, Kumar JS (2010) PET tracers for the peripheral benzodiazepine receptor and uses thereof. *Drug Discov Today* 15:933-942.
- Shaked I, Tchoresh D, Gersner R, Meiri G, Mordechai S, Xiao X, Hart RP, Schwartz M (2005) Protective autoimmunity: interferon-gamma enables microglia to remove glutamate without evoking inflammatory mediators. *J Neurochem* 92:997-1009.
- Shibley H, Smith BN (2002) Pilocarpine-induced status epilepticus results in mossy fiber sprouting and spontaneous seizures in C57BL/6 and CD-1 mice. *Epilepsy Res* 49:109-120.
- Siao CJ, Fernandez SR, Tsirka SE (2003) Cell type-specific roles for tissue plasminogen activator released by neurons or microglia after excitotoxic injury. *J Neurosci* 23:3234-3242.

- Skaper SD (2011) Ion channels on microglia: therapeutic targets for neuroprotection. *CNS Neurol Disord Drug Targets* 10:44-56.
- Sloviter RS, Dean E, Sollas AL, Goodman JH (1996) Apoptosis and necrosis induced in different hippocampal neuron populations by repetitive perforant path stimulation in the rat. *J Comp Neurol* 366:516-533.
- Somera-Molina KC, Nair S, Van Eldik LJ, Watterson DM, Wainwright MS (2009) Enhanced microglial activation and proinflammatory cytokine upregulation are linked to increased susceptibility to seizures and neurologic injury in a 'two-hit' seizure model. *Brain Res* 1282:162-172.
- Stienen MN, Haghikia A, Dambach H, Thone J, Wiemann M, Gold R, Chan A, Dermietzel R, Faustmann PM, Hinkerohe D, Prochnow N (2011) Anti-inflammatory effects of the anticonvulsant drug levetiracetam on electrophysiological properties of astroglia are mediated via TGFbeta1 regulation. *Br J Pharmacol* 162:491-507.
- Streit WJ (2002) Microglia as neuroprotective, immunocompetent cells of the CNS. *Glia* 40:133-139.
- Streit WJ, Miller KR, Lopes KO, Njie E (2008) Microglial degeneration in the aging brain--bad news for neurons? *Front Biosci* 13:3423-3438.
- Streit WJ, Walter SA, Pennell NA (1999) Reactive microgliosis. *Prog Neurobiol* 57:563-581.
- Streit WJ, Xue QS (2009) Life and death of microglia. *J Neuroimmune Pharmacol* 4:371-379.
- Tanaka S, Nakamura T, Sumitani K, Takahashi F, Konishi R, Itano T, Miyamoto O (2009) Stage- and region-specific cyclooxygenase expression and effects of a selective COX-1 inhibitor in the mouse amygdala kindling model. *Neurosci Res* 65:79-87.
- Taniwaki Y, Kato M, Araki T, Kobayashi T (1996) Microglial activation by epileptic activities through the propagation pathway of kainic acid-induced hippocampal seizures in the rat. *Neurosci Lett* 217:29-32.
- Theodore WH, Carson RE, Andreasen P, Zametkin A, Blasberg R, Leiderman DB, Rice K, Newman A, Channing M, Dunn B, et al. (1992) PET imaging of opiate receptor binding in human epilepsy using [18F]cyclofoxy. *Epilepsy Res* 13:129-139.
- Tooyama I, Bellier JP, Park M, Minnasch P, Uemura S, Hisano T, Iwami M, Aimi Y, Yasuhara O, Kimura H (2002) Morphologic study of neuronal death, glial activation, and progenitor cell division in the hippocampus of rat models of epilepsy. *Epilepsia* 43 Suppl 9:39-43.
- Turrin NP, Rivest S (2004) Innate immune reaction in response to seizures: implications for the neuropathology associated with epilepsy. *Neurobiol Dis* 16:321-334.
- van Vliet EA, da Costa Araujo S, Redeker S, van Schaik R, Aronica E, Gorter JA (2007) Blood-brain barrier leakage may lead to progression of temporal lobe epilepsy. *Brain* 130:521-534.
- Vilhardt F (2005) Microglia: phagocyte and glia cell. *Int J Biochem Cell Biol* 37:17-21.
- Wang H, Pullambhatla M, Guilarte TR, Mease RC, Pomper MG (2009) Synthesis of [(125)I]iodoDPA-713: a new probe for imaging inflammation. *Biochem Biophys Res Commun* 389:80-83.
- Wang YS, White TD (1999) The bacterial endotoxin lipopolysaccharide causes rapid inappropriate excitation in rat cortex. *J Neurochem* 72:652-660.
- Werhahn KJ, Landvogt C, Klimpe S, Buchholz HG, Yakushev I, Siessmeier T, Muller-Forell W, Piel M, Rosch F, Glaser M, Schreckenberger M, Bartenstein P (2006) Decreased

- dopamine D2/D3-receptor binding in temporal lobe epilepsy: an [18F]fallypride PET study. *Epilepsia* 47:1392-1396.
- Wine RN, McPherson CA, Harry GJ (2009) IGF-1 and pAKT signaling promote hippocampal CA1 neuronal survival following injury to dentate granule cells. *Neurotox Res* 16:280-292.
- Winokur RS, Kubal T, Liu D, Davis SF, Smith BN (2004) Recurrent excitation in the dentate gyrus of a murine model of temporal lobe epilepsy. *Epilepsy Res* 58:93-105.
- Wu YP, Siao CJ, Lu W, Sung TC, Frohman MA, Milev P, Bugge TH, Degen JL, Levine JM, Margolis RU, Tsirka SE (2000) The tissue plasminogen activator (tPA)/plasmin extracellular proteolytic system regulates seizure-induced hippocampal mossy fiber outgrowth through a proteoglycan substrate. *J Cell Biol* 148:1295-1304.
- Xie N, Wang C, Lin Y, Li H, Chen L, Zhang T, Sun Y, Zhang Y, Yin D, Chi Z (2010) The role of p38 MAPK in valproic acid induced microglia apoptosis. *Neurosci Lett* 482:51-56.
- Yang F, Liu ZR, Chen J, Zhang SJ, Quan QY, Huang YG, Jiang W (2010a) Roles of astrocytes and microglia in seizure-induced aberrant neurogenesis in the hippocampus of adult rats. *J Neurosci Res* 88:519-529.
- Yang L, Li F, Ge W, Mi C, Wang R, Sun R (2010b) Protective effects of naloxone in two-hit seizure model. *Epilepsia* 51:344-353.
- Yepes M, Lawrence DA (2004) Neuroserpin: a selective inhibitor of tissue-type plasminogen activator in the central nervous system. *Thromb Haemost* 91:457-464.
- Zattoni M, Mura ML, Deprez F, Schwendener RA, Engelhardt B, Frei K, Fritschy JM (2011) Brain infiltration of leukocytes contributes to the pathophysiology of temporal lobe epilepsy. *J Neurosci* 31:4037-4050.
- Zhang F, Liu J, Shi JS (2010a) Anti-inflammatory activities of resveratrol in the brain: role of resveratrol in microglial activation. *Eur J Pharmacol* 636:1-7.
- Zhang F, Shi JS, Zhou H, Wilson B, Hong JS, Gao HM (2010b) Resveratrol protects dopamine neurons against lipopolysaccharide-induced neurotoxicity through its anti-inflammatory actions. *Mol Pharmacol* 78:466-477.
- Zhang Y, Huang P, Du G, Kanaho Y, Frohman MA, Tsirka SE (2004) Increased expression of two phospholipase D isoforms during experimentally induced hippocampal mossy fiber outgrowth. *Glia* 46:74-83.



Clinical and Genetic Aspects of Epilepsy

Edited by Dr. Zaid Afawi

ISBN 978-953-307-700-0

Hard cover, 204 pages

Publisher Intech

Published online 15, September, 2011

Published in print edition September, 2011

This book on Epilepsy was conceived and produced as a source of information on wide range of issues in epilepsy. We hope that it will help health care providers in daily practices and increase their understanding on diagnosis and treatment of epilepsies. The book was designed as an update for neuroscientists who are interested in epilepsy, primary care physicians and students in health care professions.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Martine M. Mirrione and Stella E. Tsirka (2011). A Functional Role for Microglia in Epilepsy, Clinical and Genetic Aspects of Epilepsy, Dr. Zaid Afawi (Ed.), ISBN: 978-953-307-700-0, Intech, Available from: <http://www.intechopen.com/books/clinical-and-genetic-aspects-of-epilepsy/a-functional-role-for-microglia-in-epilepsy>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the [Creative Commons Attribution-NonCommercial-ShareAlike-3.0 License](#), which permits use, distribution and reproduction for non-commercial purposes, provided the original is properly cited and derivative works building on this content are distributed under the same license.