

Microwave Assisted Disinfection Method in Dentistry

Carlos Eduardo Vergani¹, Daniela Garcia Ribeiro², Livia Nordi Dovigo¹,
Paula Volpato Sanitá¹ and Ana Claudia Pavarina¹

¹Araraquara Dental School, UNESP- Univ Estadual Paulista,
Department of Dental Materials and Prosthodontics, Araraquara, SP,
²Department of Dentistry, Ponta Grossa State University, Ponta Grossa, PR,
Brazil

1. Introduction

Control of cross-infection has been a subject of interest to the dentistry area over the last few decades, due to the concern about the transmission of infectious-contagious diseases, such as AIDS, hepatitis, tuberculosis, pneumonia, and herpes, between the dental patients and dental personnel and the dental office and dental prosthesis laboratory (Codino & Marshall, 1976; Infection control in the dental office, Council on Dental Materials and Devices, 1978; Sande et al., 1975). This concern began in the 1970s, when the microorganism *Mycobacterium tuberculosis* was isolated in patients' molds (Ray & Fuller, 1963) and it was found that *Mycoplasma pneumoniae* was transmitted to laboratory technicians when they performed wear on contaminated dentures (Sande et al., 1975). Several items, such as instruments (Tarantino et al., 1997), dental burs (Rizzo, 1993), impressions (Abdelaziz et al., 2004), gypsum casts (Berg et al., 2005; Davis et al., 1989), dental appliances (Rohrer & Bulard, 1985), and others are often heavily contaminated with microorganisms from saliva and blood. Powell et al. (1990) also observed that 67% of all the materials received in the laboratories, among them crowns, molds wax records, and dentures, were contaminated with pathogenic microorganisms. The constant exposure of dental personnel and instruments to saliva and blood in virtually every procedure is an ever present hazard and potential contributor to the transmission of infection. Therefore, the use of an inadequate disinfection procedure in the handling of dental materials not only places the unwary staff at risk, but also results in a high level of avoidable cross-contamination.

Dental practitioner has a legal and ethical responsibility to prevent infections in patients and staff members and an interest in protecting her-himself from contracting a disease from a patient. The Centers for Disease Control and Prevention of the United States (Kohn et al., 2004) have recommended sterilization and disinfection procedures to guarantee the safety of dental treatment in the dental health care settings. Typical methods of sterilization and disinfection include dry heat at 160 to 180°C for two hours, wet steam under pressure at 121°C for at least 15 minutes (autoclaving), immersion in chemical solutions, gamma radiation, and ethylene dioxide gas. However, these methods can be time-consuming and some of them require special and costly equipment, knowledgeable operative personnel, and constant surveillance. In order to overcome these limitations, microwave irradiation has

been recommended as a practical physical sterilization method (Cottone et al., 1991) that is as effective as autoclaving (Tate et al., 1995). The lethal effects of a high frequency electric field on microorganisms was first described in 1925 (Kahler, 1929) and the destructive effects of strong radio-frequency fields on microorganisms were investigated in 1954 (Brown & Morrison, 1954). In fact, there are studies that have pointed out microwaves as a method for disinfecting food (Culkin & Fung, 1975), contact lenses (Hiti et al., 2001; Rohrer et al., 1986), laboratory microbiologic materials (Border & Rice-Spearman, 1999; Latimer, 1977), items of intimate clothing contaminated with *Candida albicans* (Friedrich & Phillips, 1988), hospital garbage (Hoffman & Hanley, 1994), coloring matter used in the cosmetic industry (Jasnow, 1975), and instruments used in medicine (Rosaspina et al., 1994). Hence, interest in this area has been maintained and in 1985 the technology was applied to the sterilization of dental appliances (Rohrer & Bulard, 1985).

In dentistry, microwave irradiation has been used for several purposes, including disinfection of toothbrushes (Nelson-Filho et al., 2011; Spolidorio et al., 2011), tongue scrapers (Spolidorio et al., 2011), instruments (Tarantino, 1997), contaminated gauze (Border & Rice-Spearman, 1999; Cardoso et al., 2007), dental burs (Fais et al., 2009; Rizzo, 1993), composite polishing instruments (Tate et al., 1995, 1996), molds made from elastomers (Abdelaziz et al., 2004), and gypsum casts (Berg et al., 2005; Davis et al., 1989). Furthermore, microwave irradiation has been widely accepted for polymerizing acrylic resin (Ilbay et al., 1994), drying gypsum products and investment materials (Hersek et al., 2002; Luebke & Chan, 1985; Luebke & Schneider, 1985; Tuncer et al., 1993), and as a post-polymerization treatment for reducing the residual monomer contents of polymerized acrylic resins and its cytotoxicity (Jorge et al., 2009; Nunes de Mello et al., 2003). Besides these purposes, some relevant applications of microwave energy in the dentistry field are the disinfection of removable dentures (Dovigo et al., 2009; D.G. Ribeiro et al., 2009; Rohrer & Bulard, 1985; Sanitá et al., 2009; Silva et al., 2006) and the use of this disinfection method to treat patients with oral candidiasis (Banting & Hill, 2001; Neppelenbroek et al., 2008; Webb et al., 2005).

Although the lethal action of microwaves on microorganisms is well established in the literature, the mechanism of destruction of microwaves is not completely understood. While some studies sustain that the effect of microwave irradiation on microorganisms is directly of thermal character (Fitzpatrick et al., 1978; Jeng et al., 1987; Yeo et al., 1999), others claim that the killing of the organisms probably also results from the non-thermal effects of microwaves (Carrol & Lopez, 1969; Culkin & Fung, 1975; Olsen, 1965; Rohrer et al., 1986). In order to attempt these mechanisms of destruction of microwaves, many different microwave regimes have been tested and advocated (Banting & Hill, 2001; Neppelenbroek et al., 2008; Rohrer & Bulard, 2001; Sanitá et al., 2009). Efficacy of microwave irradiation seems to be associated with the vehicle in which the dentures are immersed, the time of exposure, the level of power of the microwave oven, and the type of microorganisms. In addition, when selecting a disinfection procedure, its effect on the physical and mechanical properties of the irradiated materials must be carefully considered. Thus, the establishment of different protocols must be essential to each particular case, with the goal of achieving consistent sterilization without harming dental materials.

Based on the given information, the purposes of this chapter are: 1. to explain and describe the range of applications of microwave irradiation in dentistry field; 2. to review the microbiological effectiveness and recommended protocols of microwave irradiation; 3. to show the mechanisms of action of the microwaves on the microorganisms; and 4. to discuss the effects of microwave irradiation on the properties of dental materials and appliances.

2. Applications in dentistry field

The recognition of the potential for transmission of numerous infectious microorganisms during dental procedures has led to an increased concern for infection control in dental practice. Approaches to the clinical use of microwaves for preventing cross-infection have shown relevant results. Devices and instruments used in dental offices have been identified as a source of cross-contamination among patients and from patients to dental personnel. With this in mind, investigations were undertaken to explore the efficacy of microwave irradiation in disinfecting dental mirrors (Tarantino et al., 1997) and handpieces (Rohrer & Bulard, 1985). In addition, dental burs, which may become heavily contaminated with necrotic tissues, saliva, blood, and potential pathogens during use, can also be sterilized by microwave irradiation (Fais et al., 2009; Rizzo, 1993; Rohrer & Bulard, 1985). In order to prevent cross-infection, microwave energy can also be used to the disinfection of finishing and polishing instruments (Tate et al., 1995, 1996). As any another device used in dental offices, finishing and polishing instruments routinely come into contact with patient's saliva and blood and may also act as a source of cross-contamination. In accordance with the studies of Tate et al. (Tate et al., 1995, 1996), these dental devices can be effectively sterilized by microwave irradiation.

Another common dental procedure that may cause cross-infection, especially between patients and dental laboratory personnel, is the making of impressions. Previous studies showed that the majority of impressions arriving at a dental laboratory were contaminated with bacteria and other microorganisms (Egusa et al., 2008; Powel et al., 1990; Ray & Fuller, 1963; Sande et al., 1975; Sofou et al., 2002), irrespective of whether they had been exposed to a disinfectant procedure or merely rinsed with tap water (Sofou et al., 2002). The study of Egusa et al. (2008) showed an extensive contamination of alginate impressions with oral streptococci, staphylococci, *Candida* spp., methicilin resistant *Staphylococcus aureus* (MRSA), and *Pseudomonas aeruginosa*. These results indicate that a large number of microbes are retained on impression materials and are viably transferred onto the surface of stone casts. In fact, it has been shown that microorganisms can be transferred from impressions to gypsum models (Egusa et al., 2008; Leung & Schonfeld, 1983) and that the dental casts are potential sources of microbial transmission (Egusa et al., 2008; Leung & Schonfeld, 1983; Sofou et al., 2002). Also, even a cast from a properly disinfected impression may subsequently become contaminated by a technician or clinician. Considering all these information, microwave irradiation has been suggested to the disinfection of both impressions (Abdelaziz et al., 2004) and dental casts (Berg et al., 2005; Davis et al., 1989). The clinical relevance of impressions and dental casts microwave disinfection is that this procedure can be performed quickly and repeatedly, without the use of toxic, pungent, or allergenic chemicals. However, the disinfection of impression materials hinders possible cross-contamination only at the time the cast is poured. Because casts become contaminated after the intra-oral adjustments of dental appliances, they must be regarded as the major vehicle for cross-contamination and should be disinfected using microwave energy throughout all phases of the dental treatment.

Another application of microwave irradiation in preventing cross-infection is the disinfection of disposable materials. Although there is little scientific support for this purpose, some reports documented the disinfection of contaminated gauze and swabs (Border & Rice-Spearman, 1999; Cardoso et al., 2007) with the use of microwave irradiation. Cardoso et al. (2007) and Border & Rice-Spearman (1999) demonstrated that a short period

of exposure of contaminated gauze pieces and swabs to microwave energy (30 seconds) inhibit the growth of pathogenic microorganisms. The authors suggested that microwave oven could be used instead of an autoclave in a variety of clinical and research settings because the procedure is rapid and the equipment cost is minimal.

Among the purposes of microwave energy in cross-infection prevention, one of the most important is the disinfection of removable dentures. In addition to its contamination by the oral microorganisms, it has been reported that dentures are contaminated at various stages during their fabrication (Verran et al., 1996; Wakefield, 1980; Williams et al., 1985) and are capable of transmitting microorganisms to other materials, dental equipments, laboratory, and technicians (Kahn et al., 1982). Microscopic studies have also demonstrated that a biofilm similar to that formed on natural teeth is present on dentures (Budtz-Jorgensen & Theilade, 1983). Large quantity of *Candida* spp. (Budtz-Jorgensen & Theilade, 1983) and some bacterial species associated with systemic diseases have been found in removable dentures, with predominance of gram-positive bacteria, as *Staphylococcus* spp., *Streptococcus* spp., and *Actinomyces* spp. (Chau et al., 1995; Glass et al., 2001a; Monsenego, 2000). Gram-negative species, such as *Neisseria perflava*, *P. aeruginosa*, *Klebsiella pneumoniae*, and *Enterobacter cloacae*, have also been identified (Henderson et al., 1987; Latimer, 1977). In fact, in vivo studies (D.G. Ribeiro et al., 2009; Rossi et al., 1996) found *C. albicans*, *S. aureus*, *Streptococcus mutans*, and MRSA on the surfaces of dentures from patients, with *C. albicans* having the highest prevalence in these biofilms. Therefore, the denture can function as a reservoir of microorganisms, enabling the transmission of diseases in the dental office and from it to the prosthetic laboratories. Manipulation of dentures in the different dental procedures may also disseminate the microorganisms throughout the environment in the form of aerosols (Clifford & Burnett, 1995). These pathogens may be inhaled by the dentist, assistants, and laboratory technicians, resulting in cross-infection between patients and dental personnel. In the context of denture microwave disinfection, the first studies were performed in order to demonstrate the effectiveness of microwave irradiation in inactivating microorganisms adhered to complete dentures (Thomas & Webb, 1995; Rohrer & Bulard, 1985; Webb et al., 1998). Using several protocols, positive results were obtained by in vitro and in vivo studies, proving that microwave energy can be an effective method in the disinfection of dentures (Dovigo et al., 2009; Glass et al., 2001b; D.G. Ribeiro et al., 2009; Sanitá et al., 2009; Silva et al., 2006). Thus, performing microwave disinfection of a removable denture before it is transferred to a dental laboratory, and immediately before it is returned to the patient, provides a measure of infection control for all parties.

In the course of time, the fit of dentures progressively declines as a result of time-dependent changes in the supporting tissue. In this context, hard chairside reline acrylic resins or soft denture liners are proposed for permanent or temporary improvement of denture fit. These auto-polymerizing acrylic resins allow the clinician to reline a removable prosthesis directly in the mouth in intimate contact with a large area of oral mucosa. Although such liners improve the denture fit and offer comfort to some edentulous and partially edentulous patients, these materials may present a source of other problems. One of their greatest disadvantages is the ongoing task of hygiene maintenance. Denture reline materials, especially the soft liners, have been found to be more prone to microbial adhesion than acrylic resin base materials because of their surface texture and higher porosity (Nikawa et al., 1992). The rougher surface of a prosthesis with a soft liner exhibits greater colonization by *C. albicans* when compared with a conventional acrylic resin denture (Wright et al., 1985). As a result, the oral mucosa

is more susceptible to infections. A study evaluated the effectiveness of microwave disinfection on three hard chairside reline resins and observed a consistent sterilization after microwave exposure (Neppelenbroek et al., 2003). In other investigations (Dixon et al., 1999; Mima et al., 2008), microwave irradiation resulted in sterilization of a hard chairside reline resin and soft denture liners contaminated by four pathogenic microorganisms. It is also important to emphasize that disinfection by microwaves promotes inactivation of the pathogenic microorganisms present on both the surface and inside the pores of the acrylic materials (Chau et al., 1995; Dixon et al., 1999).

By being a reservoir of pathogens, the tissue surface of the acrylic resin denture enhances the infective potential of microorganisms and favors the appearance of oral infections (Budtz-Jorgensen, 1990). Oral candidiasis, represented by denture stomatitis in denture wearers, is one of the most common manifestations of disease associated with the use of removable dentures. Denture stomatitis is mainly caused by microorganisms of the *Candida* genus and normally affects the palate of approximately 65% of denture wearer patients (Chandra et al., 2001). This infection is characterized by the presence of multiple hyperemic points in the mucosa subjacent to the removable dentures of patients and, in more advanced cases, diffused erythematous areas and papillary hyperplasia of the palate may also be observed (Newton, 1962). Considering that microbial adherence and colonization on dental prostheses favor the appearance of denture stomatitis, the cleansing and disinfection of dentures is fundamental to prevent this disease. Recent studies have suggested denture disinfection by microwave irradiation for the treatment of denture stomatitis. The microwave regimes established in laboratorial studies provided the baseline for subsequent clinical trials. Banting & Hill (2001) conducted the first study that evaluated the effectiveness of microwave energy for denture disinfection as a co-adjuvant in the treatment of denture stomatitis. The authors observed that the method was effective in the reduction of the clinical signs of infection. These findings are in agreement with those found by Webb et al. (2005) a few years later. A more recent study conducted by Neppelenbroek et al. (2008) also evaluated the effectiveness of complete denture disinfection by microwave energy in the treatment of patients with denture stomatitis. In agreement to Banting & Hill (2001) and Webb et al. (2005), it was observed that disinfection of the dentures by microwaves was effective for the treatment of denture stomatitis (Neppelenbroek et al., 2008). Investigations are still being carried out to evaluate the effectiveness of microwave irradiation in the treatment of denture stomatitis (Silva et al., 2008; Vergani et al., 2008). Further tests have been performed in order to evaluate the effectiveness of denture irradiation in the treatment of diabetic denture wearer patients with denture stomatitis, showing promising outcomes (Sanitá et al., 2010, 2011). Based on the above mentioned clinical studies, the use of microwaves has shown important results for the treatment of denture stomatitis. This disease is one of the most frequent opportunistic infections found in denture wearers, including the diabetic patients, and it may extend regionally and result in a systemic infection that is associated with high mortality rates (Colombo et al., 1999; Meunier-Carpentier et al., 1981). Hence, the prevention of colonization of the oropharynx is critically important in preventing systemic infections due to *Candida*.

A recent study has shown that pathogenic microorganisms can adhere in toothbrushes and tongue scrapers made from stainless steel- and polystyrene-based injection-moulded plastic (Spolidorio et al., 2011). Thus, toothbrushes and tongue scrapers become contaminated after use and, if not properly disinfected, may be a reservoir of microorganisms that maintain their viability for a significant amount of time, ranging 24 hours to 7 days (Nelson-Filho et

al., 2006). Microbial survival promotes reintroduction of potential pathogens in the oral cavity or dissemination to other individuals when cleaning devices are stored together or shared (Ankola et al., 2009). Hence, these cleaning devices should be regularly disinfected. With this in mind, investigations have demonstrated the efficacy of microwave irradiation for disinfection of toothbrushes and tongue cleaners (Nelson-Filho et al., 2011; Spolidorio et al., 2011), suggesting that this may be a practical and low-cost alternative method of disinfection that can be easily used in the oral hygiene care practices.

This section of the chapter describes the several applications of microwave energy in dentistry field. The use of microwave irradiation for rapid disinfection of different dental materials and appliances may be an important tool in the context of the prevention of cross-infection in dental practice. In addition, there is sufficient scientific evidence that the use of this physical method of denture disinfection is effective in the treatment of denture stomatitis.

3. Microbiological effectiveness and recommended protocols

The microbiological effectiveness of microwave irradiation has been documented in the literature and the effectiveness seems to be directly related to the protocol adopted. When defining a microwave irradiation protocol, the parameters to be considered are: the time of exposure; the level of power of the microwave oven; the material to be irradiated; the vehicle in which the material is immersed, and the type of microorganisms that colonize the material. Considering these parameters and the several applications of microwave energy in dentistry field, different protocols of microwave irradiation are available and have been tested.

There are some studies that evaluate microwave irradiation for the disinfection of dental air turbine handpieces (Rohrer & Bulard, 1985), mirrors (Tarantino et al., 1997), burs (Fais et al., 2009; Rizzo, 1993; Rohrer & Bulard, 1985), and finishing and polishing instruments (Tate et al., 1995, 1996). In the study of Rohrer & Bulard (1985), handpieces contaminated with a mixture of four aerobic bacteria (*Staphylococcus epidermis*, *S. aureus*, *K. pneumonia*, *Bacillus subtilis*, *Clostridium histolyticum*) and *C. albicans* were consistently sterilized with an exposure of 10 minutes at 720W to microwave irradiation when the materials were attached to a three-dimensional rotating device. These authors also observed that handpieces contaminated by both polio type 1 and herpes simplex type 1 viruses were consistently sterilized after an exposure to microwave irradiation. Another in vitro investigation proved that dental mirrors contaminated with *S. aureus*, *B. subtilis*, and *Bacillus stearothermophilus* can be sterilized by microwave irradiation at 600W for 4 minutes, with the instruments immersed in an aldehyde solution (Tarantino et al., 1997). A microwave regime of 10 minutes at 720W is sufficient to sterilize carbide and diamond burs contaminated with a mixture of *S. aureus*, *K. pneumoniae*, and *B. subtilis* (Rohrer & Bulard, 1985). In another effective protocol, carbide burs are individually placed in a loosely capped test tube with 10 mL of distilled water, transferred to the right lateral position inside the microwave oven, and then exposed to 5 minutes at 600W (Fais et al., 2009). A similar protocol consisting of 6 minutes of irradiation at 750W (Tate et al., 1995, 1996) can be adopted when disinfecting finishing and polishing instruments (Enhance finishing cups, L.D. Caulk Co., Milford, DE, USA and Min-Identoflex fine cups, Centrix Inc., Shelton, CT, USA).

Likewise, 10 minutes of irradiation at high power can sterilize customized impressions made from both vinyl polysiloxane (Cinch Platinum, Parkell, Farmingdale, NY, USA) and

polyether (Impregum F, 3M ESPE AG Dental Products, Seefeld, Germany) rubber impression materials (Abdelaziz et al., 2004). Microwave irradiation can also be used for gypsum casts disinfection (Berg et al., 2005; Davis et al., 1989). In the studies of Davis et al. (1989) and Berg et al. (2005), molds were contaminated with pathogenic microorganisms and the stone casts obtained were submitted to microwave irradiation. While *Serratia marcescens* cells on casts were inactivated by microwave irradiation at 900W for 1, 5 or 20 minutes (Davis et al., 1989), *S. aureus* and *P. aeruginosa* were killed after 5 minutes of irradiation (Berg et al., 2005). Microwave irradiation was ineffective in killing *B. subtilis* transferred to stone casts (Davis et al., 1989). Although a complete inactivation was not obtained, the gypsum casts submitted to microwaves for 20 minutes exhibited less growth than the samples irradiated for shorter times (Davis et al., 1989).

There are also two available protocols for the microwave disinfection of disposable materials, such as swabs and gauze. Cardoso et al. (2007) used stock cultures of *Escherichia coli*, *S. aureus*, *S. epidermidis*, *P. aeruginosa*, and *C. albicans* to contaminate gauze and swabs. Thereafter, the materials were placed into autoclave bags, sealed, and irradiated for 30 seconds at 1000W. This time/power regime was efficient in the sterilization of the disposable materials tested. When a lower power was used (650W), 30 seconds of microwave irradiation were also sufficient to sterilize gauze contaminated with the same pathogenic bacteria and fungi (Border & Rice-Spearman, 1999). With the regard of oral cleaning devices, disinfection of toothbrushes contaminated with a suspension containing *S. mutans* was obtained after exposure to microwaves for 7 minutes at 1100W (Nelson-Filho et al., 2011). However, the results from another study demonstrated that toothbrushes and tongue scrapers contaminated with *C. albicans*, *S. aureus*, and *S. mutans* were effectively disinfected after 1 minute of microwaving at 650W (Spolidorio et al., 2011).

Given the efficacy of microwave disinfection, much attention has been focused on the use of this method for denture decontamination. Various studies have been conducted to determine the most suitable time/power protocol when using microwave irradiation to disinfect dentures. Some studies have used home microwave ovens for the inactivation of pathogenic microorganisms, such as those recommended by the *Handbook of Disinfectants and Antiseptics* (Cole & Robison, 1996). Among these microorganisms, there are those considered indicators of sterilization, such as *S. aureus* (gram-positive bacteria), *P. aeruginosa* (gram-negative bacteria), and *B. subtilis* (sporulated aerobic microorganisms). In this context, Rohrer & Bulard (1985) investigated the possibility of using microwave irradiation to sterilize dentures and concluded that 8 minutes of irradiation at 720W were sufficient to sterilize the dentures contaminated with a mixture of five bacteria and a fungus. To obtain these results, the authors attached the dentures to a three dimensional rotating device. However, such device is not commercially available or practical for use by dentists or health care facilities. Ten years later, Thomas & Webb (1995) observed that an unmodified domestic microwave oven could be used in the disinfection of dentures. They also demonstrated that sterilization of dentures inoculated with *C. albicans* and *Streptococcus gordonii* could be achieved at 2, 4, 6, 8, and 10 minutes exposure times at high setting (650W) (Webb et al., 1998). Likewise, using an unmodified domestic microwave oven with a rotating table, Baysan et al. (1998) observed that microwave irradiation at 650W for 5 minutes promoted a reduction in the quantity of *C. albicans* and *S. aureus* present in resilient relining materials. The study of Meşe & Meşe (2007) also investigated the effect of microwave energy on the growth of *C. albicans* in resilient relining material and verified the reduction in the colony counts of the fungus after dry exposure to microwaves for 5 minutes (650W). The results

obtained with the aforementioned microbiological studies, in which the test specimens or dentures were irradiated in a dry state, are variable with regard to the effectiveness of disinfection by microwaves. Since irradiation in water provides uniform heating of the materials, Dixon et al. (1999) suggested immersing the samples in water during exposure to microwaves. This procedure was considered adequate for eliminating microorganisms, including those located inside the pores. The authors inoculated *C. albicans* in resilient liners and a heat-polymerized resin and showed that the specimens immersed in water during irradiation were completely disinfected after 5 minutes at maximum power, while those not immersed in water were only partially disinfected after the same time of exposure. Bearing in mind these results, Neppelenbroek et al. (2003) evaluated the effectiveness of a home microwave oven for the inactivation of *S. aureus*, *P. aeruginosa*, *B. subtilis*, and *C. albicans* present in three reline resins. The contaminated samples were immersed in 200 mL of sterile distilled water and irradiated for 6 minutes at 650W. It was observed that all the test specimens were sterilized after irradiation, as no microbiological growth was noticed after the irradiated specimens had remained incubated for 7 days. Silva et al. (2006) evaluated the same protocol for disinfecting simulated complete dentures and observed that the lethal action varied according to the microorganisms tested. While complete disinfection was achieved for the dentures contaminated with *S. aureus* and *C. albicans*, those contaminated with *P. aeruginosa* and *B. subtilis* showed little, but detectable, microbial growth. The different results from those of Neppelenbroek et al. (2003) are probably related to the larger surface area of the complete dentures, given that the number of microorganism colonies on the acrylic resin surface is proportional to the total area involved. In addition, this difference could be related to the microorganisms tested. A greater resistance of *B. subtilis* to microwaves has been reported (Davis et al., 1989; Najdovski et al., 1991) and these results are probably related to the sporulation capacity of *B. subtilis*. Bacterial spores are metabolically inactive and particularly resistant to situations of stress, such as heating and radiation. Microwaves promote heating of the test specimens and water in which they are immersed, thus there is the possibility of spore formation during this procedure (Najdovski et al., 1991). A more recent study (Mima et al., 2008) showed that the experimental protocols advocated by Dixon et al. (1999), Neppelenbroek et al. (2003), and Silva et al. (2006) could be used with lower exposure times. Test specimens contaminated with four microorganisms (*S. aureus*, *P. aeruginosa*, *B. subtilis*, and *C. albicans*) were immersed in water and submitted to microwave irradiation (650W) at different exposure times (5, 4, 3, 2, and 1 minutes). It was observed that all the test specimens irradiated for 3, 4, and 5 minutes were completely disinfected after microwave irradiation. When the time of irradiation was reduced to 2 minutes, the samples contaminated with *C. albicans* were completely disinfected while those inoculated with *S. aureus*, *P. aeruginosa*, and *B. subtilis* demonstrated microbial growth. When submitted to sterilization by humid heat, bacterial cells are inactivated at higher temperatures than fungal cells (Pelczar et al., 1993). Therefore, irradiation for at least 2 minutes promoted sufficient water heating to inactivate *C. albicans* but not the bacteria. Moreover, the yeast cells are larger than those of the bacteria (Verran & Maryan, 1997). Therefore, one could suppose that the *C. albicans* cells contained more water in their composition than did the other microorganisms tested, and therefore, they had been more susceptible to microwave irradiation. In spite of this *in vitro* study has evaluated small size test specimens that had surfaces with vitreous characteristics, differently from those observed clinically, its results indicated that the protocol should be evaluated in other experimental conditions. Therefore, laboratorial investigations were performed to evaluate the effectiveness of this protocol in

the disinfection of dentures contaminated by several microorganisms. Sanitá et al. (2009) demonstrated that simulated complete dentures inoculated with different *Candida* spp. (*C. albicans*, *Candida tropicalis*, and the intrinsically resistant *Candida glabrata*, *Candida dubliniensis*, and *Candida krusei*) were completely disinfected by microwave irradiation for 3 minutes at 650W. Similar results have been reported for complete dentures contaminated with *S. aureus* and *P. aeruginosa* (Dovigo et al., 2009).

Given the promising results, some protocols were tested in clinical trials to evaluate the effectiveness of microwave irradiation in disinfecting patients' dentures (Glass et al., 2001b; D.G. Ribeiro et al., 2009). In one study, fragments of dentures that had been worn for periods ranging from 12 days to 48 years were immersed in a chemical solution (MicroDent Sanitizing and Cleaning System®) and exposed to microwaves for 2 minutes (Glass et al., 2001b). This method showed positive results for denture decontamination, considering that no microbial growth was observed on the fragments. In addition, the study of D.G. Ribeiro et al. (2009) showed that 3 minutes of irradiation at 650W resulted in complete inactivation of the biofilm present on dentures of 30 patients. It emerged also from this study that a lower reduction in the count of microorganisms (*C. albicans*, *S. aureus*, and *S. mutans*) was observed when a lower time of exposure was used (2 minutes).

In terms of treating denture stomatitis, Banting & Hill (2001) conducted the first clinical study that evaluated the effectiveness of microwave energy for denture disinfection as a co-adjuvant treatment. This study was performed in 2001, when the effective protocol of 3 minutes at 650W had not yet established. Patients received topical antifungal medication (nystatin/ three times a day) for 14 days and had their dentures irradiated (850W for 1 minute) on three different days (1st, 5th and 10th day). The authors observed that the method was effective in the reduction of the clinical signs of infection and of the invasive forms of *C. albicans* (pseudohyphas) adhered on the surfaces of the dentures. In another clinical study, microwave disinfection of dentures (10 minutes at 350W) in a daily basis during one week reduced the palatal inflammation and the numbers of *Candida* on cultures from the palates and dentures of patients (Webb et al., 2005). A more recent study (Neppelenbroek et al., 2008) also evaluated the effectiveness of denture microwave disinfection in the treatment of patients with denture stomatitis. The microwave treatment protocol adopted was 6 minutes of irradiation of the complete dentures at 650W, three times a week for 30 days. It was verified that disinfection of the dentures by microwaves was effective for the treatment of denture stomatitis and for the reduction of the mycelial forms of *Candida* spp. In agreement to Banting & Hill (2001), these authors also observed that the levels of recurrence of *C. albicans* on the internal surfaces of the dentures and the supporting mucosa were significantly reduced in the patients whose dentures were microwaved (Neppelenbroek et al., 2008). Other clinical investigations have been carried out to evaluate the effectiveness of microwave irradiation in the treatment of denture stomatitis. A modification on the protocol proposed by Neppelenbroek et al. (2008) was evaluated by Vergani et al. (2008) and Silva et al. (2008). These authors demonstrated that denture microwave irradiation for 3 minutes at 650W, three times weekly for 14 days, is able to reduce the clinical signs of denture stomatitis on the palatal mucosa and the *Candida* colonization on complete dentures. Further clinical evaluations have also been conducted in order to evaluate the effectiveness of this protocol in the treatment of diabetic denture wearer patients with denture stomatitis (Sanitá et al., 2010, 2011). It was observed that microwave disinfection of complete dentures, by itself, was as effective as nystatin, the more conventional topical antifungal medication, in reducing the *Candida* counts and the clinical signs of denture stomatitis infection in patients with diabetes mellitus.

Based on the aforementioned studies, it can be seen that several regimes of microwave irradiation in relation to time/power are available. The protocol must be selected in accordance to the specific application of the microwave energy. Regardless these parameters, microwave irradiation is a potentially effective method for inactivating various microbial species present on dental materials, many of which are related to oral pathologies.

4. Mechanisms of action on the microorganisms

While the inhibitory effect of microwave irradiation on microorganisms is being researched extensively, how microwave brings about this effect has been a matter of discussion. Some authors believe that microorganism inactivation by microwave irradiation is explained by a thermal effect (Fitzpatrick et al., 1978; Jeng et al., 1987; Yeo et al., 1999). Nevertheless, several studies suggest that, in addition to the heat generated around the microorganisms, there are other mechanisms resulting directly from the interaction of the electromagnetic field (Carrol & Lopez, 1969; Culkin & Fung, 1975; Olsen, 1965; Rohrer et al., 1986). Various mechanisms have been suggested to explain the nature of the so called non-thermal theory. Depending on the chemical composition of the microorganisms and the surrounding medium, the microbial cells may be selectively heated by the microwaves (Carrol & Lopez, 1969; Hiti et al., 2001; Yeo et al., 1999). Therefore, a certain frequency of microwave energy may be absorbed by certain fundamental biological molecules, such as the nucleic acids (Rohrer et al., 1986). Moreover, the level of molecular response from the biological system to the quantity of thermal energy may also explain the non-thermal effect of microwaves (Carrol & Lopez, 1969). The structural changes in the more peripheral layer around the biological macromolecules may alter their stability and function, and, consequently, these molecules may be denatured in an irreversible manner (Culkin & Fung, 1975). Studies have also demonstrated that the exposition of bacterial suspensions to microwave irradiation caused reduction on viable cell counts and increased the leaching of DNA and protein (Woo et al., 2000). These results suggest that microwaves caused changes in structural integrity and permeability of cell membrane and cell wall that have detrimentally affected the cell metabolism and lead to cell death (Campanha et al., 2007; Carrol & Lopez, 1969; Culkin & Fung, 1975; Olsen, 1965). Campanha et al. (2007) verified that leveduriform suspensions submitted to microwave irradiation at 650W for 6 minutes presented significantly lower cell count values and a larger number of substances released in comparison with the non-irradiated suspensions. The distinction between integral and non-integral cells was made based on the entry of methylene blue coloring into the cells, which is an indirect form of evaluating the cell membrane and wall integrity. Disintegrated cells were found in the irradiated suspension, indicating an alteration in the permeability or integrity of these structures. Moreover, the cells of this suspension lost their characteristic refringence, in spite of preserving their ellipsoidal leveduriform morphology. It was also demonstrated that after irradiation by microwaves, the release of electrolytes (K^+ , Ca^{++}) and nucleic acids was significantly higher in the irradiated suspensions than that from the non-irradiated (Campanha et al., 2007). However, despite of cell inactivation, the optic density and cell concentration were not altered in comparison with those of the control suspensions, indicating that cells were not completely lysed. Irrespective of the mechanism of microwaves on pathogenic microorganisms being thermal or non-thermal, it is known that the effect of inactivation occurs mainly in the presence of water, this being an important factor for sterilization in microwave ovens. Freeze-dried or dry organisms are unlikely to be

affected, even when submitted to prolonged exposures, indicating that humidity plays an important role in microwave energy absorption (Dixon et al., 1999; Watanabe et al., 2000). The water molecules present in the medium or inside the cells, being diploid, interact with the electromagnetic field of the microwaves. Consequently, numerous intermolecular collisions may occur and this vibration produces heating (Najdovski et al., 1991). This increase in temperature may cause protein and DNA denaturation (Ponne & Bartels, 1995). The consistent results from several studies, in which strains were completely inactivated when microwave irradiation was carried out with the specimens immersed in water, confirm this hypothesis.

According to scanning electron microscopic (SEM) studies (Neppelenbroek et al., 2003; Mima et al., 2008), microwave irradiation not only inactivated *C. albicans*, but also removed the nonviable yeast cells from resin surface. In this case, the irradiation was performed with the resin specimens immersed in water and, since water started to boil after approximately 1.5 minutes of irradiation, the movement of the water particles probably removed microbial cells from the resins. Verran & Maryan (1997) reported that the larger yeasts cells are more easily dislodged from acrylic resin surfaces compared with smaller bacteria. Considering the information discussed above, it can be concluded that, the nature of the lethality of the microwave irradiation for microorganisms may be a combination of thermal and non-thermal effects.

5. Effects on physical and mechanical properties of dental materials and appliances

Several investigations have focused on finding the adequate microwave parameters for microbial inactivation and cross-infection control. Different irradiation protocols have proved to be remarkably effective for disinfection of dental prostheses and other materials frequently used in dental practice. However, to enable this method of disinfection to be safely recommended, it is important to clearly demonstrate that it does not exert deleterious effects on the physical and mechanical properties of the materials submitted to microwaves. For this reason, studies have been conducted to examine the effect of microwave disinfection on dental instruments and burs, impressions, gypsum products, acrylic resins, denture lining materials, and artificial teeth.

Rohrer & Bulard (1985) exposed dental air turbine handpieces to microwaves for 2, 4, 6, 8, 10, and 15 minutes (720W). After 25 cycles of 10 minutes, the dental handpiece tested showed no decrease in the pressure reading and no apparent alteration in sound or cutting power. Another study (Tate et al., 1996) evaluated instrument performance of two composite finishing and polishing systems before and after three cycles of microwave irradiation (6 minutes at 750W). The sample surfaces were examined with a profilometer after the finishing procedure and the results demonstrated that the systems tested can be submitted to microwave irradiation at least three times without affecting performance. Questions have also been raised about the effects of microwave regimens on the microscopic characteristics, durability, and strength of dental burs, which can have their sharpness and ability to effectively cut tooth structure altered. The effect of sterilization with microwaves on diamond burs was evaluated by Rizzo (1993). The author evaluated the dental burs by viewing them under stereomicroscope before and after sterilization cycles. It was found that no damage was present after 15 cycles. The possible influence of microwave irradiation on the cutting capacity of carbide burs was also investigated (Fais et al., 2009). The burs were

used to cut glass plates in a cutting machine set for 12 cycles of 2.5 minutes each and, after each cycle, they were exposed to microwave irradiation for 5 minutes at 600W. The cutting capacity of the burs was determined by a weight-loss method. Compared to the control conditions, the microwaved burs showed a statistically significant decrease in their cutting capacity. Thus, microwave irradiation requires further investigations before final recommendations can be made for disinfection of carbide burs.

Microwave disinfection of rubber impressions was also suggested by some authors as an alternative approach of controlling microbial transmission. In this context, the reproducibility of rubber impressions after microwave irradiation (10 minutes/720W) has been evaluated and the results compared with other disinfection methods (Abdelaziz et al., 2004). Microwave sterilization had a small effect on accuracy of impressions and this procedure has been recommended as a suitable technique for sterilizing rubber impressions. Another application of microwaves in dentistry is the disinfection of gypsum casts. Although there are no studies evaluating the disinfection protocols on the properties of gypsum materials, the effect of drying casts by microwaves has been described. In this context, microwave irradiation of gypsum casts has been tested as to its effect on the resistance to fracture (Hersek et al., 2002; Luebke & Schneider, 1985; Tuncer et al., 1993) and hardness (Luebke & Chan, 1985). In general, the results indicated an improvement in these properties. However, there was some concern that a decrease in the compressive strength and the appearance of cracks or porosities in the surface might occur when gypsum casts were exposed to irradiation with a very high power (1450W). Other physical and mechanical properties, such as abrasion resistance and dimensional stability, should be performed to confirm the clinical applicability of this procedure.

One of the main applications of microwaves in dentistry is to disinfect dental prosthesis. A large number of investigations have been published in the past and recent years concerning its effectiveness and limitations. Laboratorial investigations aimed at identifying if microwave exposure affects the surface hardness of the denture base acrylic resins, relining materials, and artificial denture teeth. The hardness of a material is the result of the interaction of several properties, such as ductility, malleability, and resistance to cutting, and, therefore, hardness tests may be used as an indicator of these properties (Anusavice, 1996). Also, the hardness of materials is related to its resistance to local plastic deformation. Two universal types of microhardness test, Vickers and Knoop, are standard methods for measuring hard surfaces, while the Shore A measures hardness in terms of the elasticity of the material. It has been demonstrated that the Vickers hardness of a heat-polymerizable acrylic resin was not changed after different times of exposure to microwaves (6, 5, 4, 3, 2, and 1 minutes) at 650W (Machado et al., 2009; D.G. Ribeiro et al., 2008). The Knoop hardness values of a denture base resin were also not changed after two cycles of microwave disinfection for 6 minutes at 690W (Sartori et al., 2008). In addition, no alterations in the Shore A hardness values were detected after microwave irradiation (3 minutes/500W) of resilient relining materials (Pavan et al., 2007). Other studies showed, however, increased hardness of denture base materials associated with microwave irradiation. Polyzois et al. (1995) evaluated two microwave disinfection protocols on the hardness of test specimens of a heat-polymerizable resin (3 or 15 minutes at 500W). Both protocols provided an increase in microhardness values. Similar findings were described by Machado et al. (2005) for two resilient relining materials submitted to seven irradiation cycles for 6 minutes (650W). D.G. Ribeiro et al. (2008) evaluated the effect of different times of exposure to microwaves (5, 4, 3, 2, and 1 minutes/ 650W) on the hardness of four reline materials and the findings suggested

that the disinfectant method promoted an increase in the hardness of the reline resins. The increase in microhardness values after microwave disinfection may be related to the increase in temperature during the irradiation procedure. Arab et al. (1989) reported an increase in hardness values when heat-polymerizable resins were immersed in water heated to 100°C. Similarly, an increase in Vickers hardness of a reline resin after heat treatment in a water bath at 55°C for 10 minutes has been reported (Seó et al., 2007a). These results may be attributed to the reduction in the level of residual monomer, as a result of the complementary processes of polymerization (Lamb et al., 1983; Sideridou et al., 2004) and diffusion of residual monomer, both favored by the increase in water temperature during microwave irradiation.

Besides the acrylic denture base resins, dentures also comprise artificial teeth. For this reason, the effect of microwave disinfection on the surface hardness of artificial teeth commonly used for denture construction was also evaluated (Campanha et al., 2005). Two microwave disinfection cycles of 6 minutes each (650W), preceded or not by immersion in distilled water for 90 days, were tested. From these experiments, a reduction in surface hardness of the artificial denture teeth was observed after microwave irradiation. It seems that the high temperature associated with the movements of the molecules probably caused an increase in the speed of diffusion of the water molecules into the polymer, facilitating the movement of the polymeric chains during performance of the hardness (Takahashi et al., 1998). Thus, it is feasible that the reduction in hardness after irradiation is related to water sorption rather than to microwave irradiation. In fact, the teeth from the group that was microwaved after 90 days of water saturation presented no significant alteration in hardness after disinfection. Since microwave disinfection involves the exposition of dentures to water at high temperature, it has been hypothesized that this may affect the bond strength between the artificial teeth and the acrylic resin from which dentures are made. Results from a study evaluating the effect of microwave disinfection (6 minutes/650W) on the bond strength of two types of denture teeth to three acrylic resins showed that microwave disinfection did not adversely affect the bond strength of all tested materials, with the exception of one tooth/resin combination (R.C. Ribeiro et al., 2008). In another study microwave irradiation for 3 minutes at 650W promoted a reduction in the impact strength of the tooth/acrylic resin interface, which could be explained by the increase in the degree of conversion of self-polymerizing resin, reducing the cohesion at the interface of the samples (Consani et al., 2008a).

The effects of microwave irradiation on other surface properties, such as roughness and porosity, have also been investigated. According to Allison & Douglas (1973), smoother surface retain a smaller quantity of biofilm, thus avoiding the proliferation of microorganisms on the acrylic surface of dentures. Surface roughness is an important characteristic of dental materials and, therefore, there is a direct correlation between the values of roughness and bacterial adherence. Moreover, according to Yannikakis et al. (2002), the presence of pores may reduce the mechanical properties of acrylic resin, as well as interfere in denture hygiene. It seems that the effect of microwave irradiation on a denture's surface roughness and porosity depends on the time of exposure, number of cycles, and the type of denture resin used. Novais et al. (2009) investigated the occurrence of porosity on the surface of four self-polymerizable acrylic resins and one heat-polymerizable resin, after two or seven cycles of microwave disinfection (6 minutes/ 650W). The number of pores found in two out of five resins remained similar after microwaving, while a reduction in porosity was observed in two resins after seven disinfection cycles. In these cases, it was

suggested that the high temperatures may have been attained during exposure to microwaves, which led to a greater degree of conversion and continuation of polymerization. Seven cycles of microwave disinfection increased the number of pores in one material. According to the authors, this material presented a high level of residual monomer and, therefore, a probable explanation for the increase in the number of pores in this resin was related to monomer vaporization. Another investigation also showed that the use of microwave energy can modify the surface texture of acrylic resins (Sartori et al., 2006). It was reported that two microwave disinfection cycles (6 minutes at 690W) promoted an increase in surface roughness of an acrylic resin. However, only one material was evaluated and the effect of reduced exposure times on surface roughness was not investigated.

From the literature, it seems that microwave disinfection can play a role in promoting changes in denture materials. The flexural strength of five chairside reline resins and one denture base resin were evaluated after two and seven cycles of microwave disinfection at 650W for 6 minutes (Pavarina et al., 2005). The flexure strength of three resins presented significant increase in strength values. In contrast, two reline resins presented reduced flexure values after microwave irradiation. The heating provided by each of the seven cycles of microwave disinfection may have increased the absorption of water of some of the evaluated materials, resulting in a reduction in the flexural strength values. In view of these results, shorter times of exposure to microwaves, and their effects on the flexural strength of acrylic resins, were investigated and the findings showed that the flexural strength of four reline materials and one heat-polymerizable resin was not detrimentally affected after 5, 4, 3, 2, and 1 minutes of microwave irradiation at 650W (D.G. Ribeiro et al., 2008). Indeed, the disinfection method was capable of significantly increase the flexural strength of one reline resin. A similar result was described elsewhere after a disinfection cycle for 3 minutes at 650W (Consani et al., 2008b). In addition, another investigation (Polizois et al., 1995) observed that the flexural properties of a heat-polymerizable resin remained unaltered after the use of a low power (500W) associated with a long exposure time (15 minutes). However, in this study the samples were irradiated in a dry condition, a procedure that has been shown to be less effective for microbiological inactivation. In a more clinically relevant approach, the effect of denture microwave disinfection on the maximum fracture load, deflection at fracture, and fracture energy of intact and relined denture bases was evaluated. After exposed to microwave irradiation for 7 days (6 minutes/650W), the strength of the denture bases was similar to the strength of those immersed in water for 7 days (Seó et al., 2008).

An aspect that has also been investigated is the influence of microwave disinfection on bond strength between resilient liners and denture base acrylic resin. Test specimens made of resilient resins bonded to a denture base resin were submitted to microwave disinfection for two and seven irradiation cycles of 6 minutes at 650W (Machado et al., 2005). Microwave disinfection did not compromise the adhesion of resilient liners to the denture base resin. In a subsequent study, seven microwave disinfection cycles (6 minutes/650W) did not decrease the torsional bond strength between two hard reline resins and a denture base resin (Machado et al., 2006). Therefore, in general, the use of microwaves for denture disinfection does not appear to have any negative effect on the bond strength of reline materials frequently used in dental practice. Recently, a clinical study (R.C. Ribeiro et al., 2009) also reported the color stability of a hard chairside reline resin after microwave disinfection.

Another important aspect that should be considered prior to the selection of a disinfection procedure is the maintenance of adequate adaptation between the denture base and residual

ridge. Clinically, this characteristic is fundamental both for denture retention and the preservation of the supporting tissues. Changes in denture base adaptation could act as one of the causes of alveolar bone loss and be indirectly responsible for decreasing the retention and stability of the denture (De Gee et al., 1979). A denture that exactly reproduces the supporting tissue may assure a uniform distribution of forces on the largest possible area of surface. Thus, analyses of the effect of microwave disinfection on the dimensional stability of denture bases resins have been conducted. Burns et al. (1990) aimed to determine the possible influence of microwaving on the dimensional stability of heat-polymerizable, self-polymerizable, and light-polymerizable resins. Test specimens were fabricated and submitted to measurements of mass and length before and after microwave irradiation (15 minutes/ 650W). The results showed that all the materials maintained dimensional stability after the disinfection procedure. In another study, microwave disinfection (3 or 5 minutes at 650W) promoted small, clinically insignificant dimensional changes on test specimens of a heat-polymerizable resin (Polyzois et al., 1995). Contrasting results were obtained in the study of Gonçalves et al. (2006), who evaluated the effect of two or seven cycles of microwave disinfection (6 minutes/ 650W) on the linear dimensional change of four relined resins and one denture base resin. Three of the evaluated resins presented significant alteration in the linear dimensional after disinfection. In spite of the positive and negative findings found by these different studies, all of the presented results cannot be extrapolated directly to a clinical situation, since the test specimens used did not simulate the dimensions and shape of denture bases. Standardized dentures were fabricated in the study of Thomas & Webb (1995) in order to evaluate the effect of microwaves on their dimensional stability. After 10 minutes of exposure to microwaves at 604W, some of the areas measured in the dentures presented significant shrinkage or expansion. Similarly, Sartori et al. (2008) observed that the disinfection cycles of 6 minutes (690W) could compromise the dimensional stability of denture base resins. In another study, however, a lower power setting (331W) decreased the occurrence of dimensional changes when the dentures were microwaved for 6 minutes (Thomas & Webb, 1995). The protocol of 6 minutes/ 650W of microwave irradiation was also tested for the dimensional stability of denture bases (Seó et al., 2007b). Repeated disinfection cycles were performed (two and seven) and an increase in shrinkage was observed both in intact bases and bases relined with heat-polymerizable resins. As the bases were immersed in water during irradiation, the results could be related to the possible occurrence of complementary polymerization as a result of water heating. In another investigation, the use of the same microwave power (650W), but with irradiation for 3 minutes, promoted no deleterious effects on the adaptation of denture bases (Consani et al., 2007). In fact, the authors observed that microwave irradiation improved the adaptation of bases in some experimental conditions. Pavan et al. (2005) also suggested that use of shorter irradiation times preserves the dimensional stability of dentures. The authors fabricated 30 denture bases that were submitted to irradiation for 3 minutes/500W or 10 minutes/ 604W. No dimensional alteration was observed in the bases irradiated for 3 minutes. Taken together, the results from all the cited studies suggest that short microwave irradiation times should be used, so that the adaptation of denture bases to subjacent tissue is not changed. Recently, Basso et al. (2010) performed a clinical evaluation of the effect of 3 minutes of microwave irradiation at 650W once or three times a week on the linear dimensional stability of complete dentures. Measurements were taken before the first microwave disinfection (baseline) and after each week of disinfection. Furthermore, the dentures were monitored clinically. Three times weekly irradiation showed significantly

higher shrinkage in all evaluated weeks. This result could be attributed to the heating generated by microwave irradiation in an already polymerized material. Even though three microwave disinfections showed statistically significant greater shrinkage, the clinical evaluation did not reveal any change. Therefore, the authors suggested that microwave irradiation can be used clinically for the disinfection of dentures and treatment of denture stomatitis.

Given the information above, it is clear that discordant results have been published regarding the risks of denture microwave disinfection. It seems that the occurrence of negative effects on physical and mechanical properties of dentures depends on the microwave protocol tested and type of material evaluated. It also seems that the use of short exposure times could minimize the occurrence of harmful effects on the dental prostheses. Taking all the results into account it would seem that the microwave regime of 3 minutes at 650W is adequate for denture disinfection without causing significant detrimental effects on the denture materials.

6. Conclusion

According to the information discussed in this chapter, there is scientific evidence to support the efficacy of microwave irradiation in preventing cross-infection and treating denture stomatitis. Several protocols of irradiation were described and discussed, and the microbiology effectiveness of microwave energy was clearly demonstrated. Regardless of all the parameters used, we can conclude that microwave disinfection is an effective, quick, easy, and inexpensive versatile tool that can be performed by dentists, assistants, technicians, patients and/or their caregivers to inactivate microorganisms. In addition, the use of a microwave oven does not require special storage and does not induce resistance for fungi or other microorganisms. Thus this method may have an important potential use in dental offices, dental laboratories, and institutions and hospitals in which patients are treated, especially those wearing removable dentures.

7. References

- Abdelaziz, K.M.; Hassan, A.M. & Hodges JS. (2004). Reproducibility of sterilized rubber impressions. *Brazilian Dental Journal*, Vol.15, No.3, (December 2004), pp. 209-213, ISSN 0103-6440
- Allison, R.J. & Douglas, W.H. (1973). Microcolonization of the denture-fitting surface by *Candida albicans*. *Journal of Dentistry*, Vol.1, No.5, (June 1973), pp. 198-201, ISSN 0300-5712
- Ankola, A.V.; Hebbal, M. & Eshwar, S. (2009). How clean is the toothbrush that cleans your tooth? *International Journal of Dental Hygiene*, Vol.7, No.4, (November 2009), pp. 237-240, ISSN 1601-5029
- Anusavice, K.J. (1996). *Phillips' science of dental materials*. (10th Ed), W.B. Saunders, ISBN 0721657419, Philadelphia, United States
- Arab, J.; Newton, J.P. & Lloyd, C.H. (1989). The effect of an elevated level of residual monomer on the whitening of denture base and its physical properties. *Journal of Dentistry*, Vol.17, No.4, (August 1989), pp. 189-194, ISSN 0300-5712

- Banting, D.W. & Hill, S.A. (2001). Microwave disinfection of dentures for the treatment of oral candidiasis. *Special Care in Dentistry*, Vol.21, No.1, (January 2001), pp. 4-8, ISSN 0275-1879
- Basso, M.F.; Giampaolo, E.T.; Vergani, C.E.; Machado, A.L.; Pavarina, A.C. & Compagnoni, M.A. (2010). Influence of microwave disinfection on the linear dimensional stability of complete dentures: a clinical study. *The International Journal of Prosthodontics*, Vol.23, No.4, (August 2010), pp. 318-320, ISSN 0893-2174
- Baysan, A.; Whiley, R. & Wright, P.S. (1998). Use of microwave energy a long-term soft lining material contaminated with *Candida albicans* or *Staphylococcus aureus*. *The Journal of Prosthetic Dentistry*, Vol.79, No.4, (April 1998), pp. 454-458, ISSN 0022-3913
- Berg, E.; Nielsen, O. & Skaug, N. (2005). High-level microwave disinfection of dental gypsum casts. *The International Journal of Prosthodontics*, Vol.18, No.6, (December 2005), pp. 520-525, ISSN 0893-2174
- Border, B.G. & Rice-Spearman, L. (1999). Microwaves in the laboratory: effective decontamination. *Clinical Laboratory Science*, Vol.12, No.3, (June 1999), pp. 156-160, ISSN 0894-959X
- Brown, G.H & Morrison, W.C. (1954). An exploration of the effects of strong radio-frequency fields on micro-organism in aqueous solutions. *Food Technology*, Vol.8, pp. 361-366, ISSN: 0015-6639
- Budtz-Jørgensen, E. & Theilade, E. (1983). Regional variations in viable bacterial and yeast counts of 1-week-old denture plaque in denture-induced stomatitis. *Scandinavian Journal of Dental Research*, Vol.91, No.4, (August 1983), pp.288-295, ISSN 0029-845X
- Budtz-Jørgensen, E. (1990). Etiology, pathogenesis, therapy, and prophylaxis of oral yeast infections. *Acta odontologica Scandinavica*, Vol.48, No.1, (February 1990), pp. 61-69, ISSN 0001-6357
- Burns, D.R.; Kazanoglu, A.; Moon, P.C. & Gunsolley, J.C. (1990). Dimensional stability of acrylic resin materials after microwave sterilization. *The International Journal of Prosthodontics*, Vol.3, No.5, (October 1990), pp. 489-493, ISSN 0893-2174
- Campanha, N.H.; Pavarina, A.C.; Vergani, C.E. & Machado, AL. (2005). Effect of microwave sterilization and water storage on the Vickers hardness of acrylic resin denture teeth. *The Journal of Prosthetic Dentistry*, Vol.93, No.5, (May 2005), pp. 483-487, ISSN 0022-3913
- Campanha, N.H.; Pavarina, A.C.; Brunetti, I.L.; Vergani, C.E.; Machado, A.L. & Spolidorio, D.M.P. (2007). *Candida albicans* inactivation and cell membrane integrity damage by microwave irradiation. *Mycoses*, Vol.50, No.2, (March 2007), pp. 140-147, ISSN 1439-0507
- Cardoso, V.H.; Goncalves, D.L.; Angioletto, E.; Dal-Pizzol, F. & Streck, E.L. (2007). Microwave disinfection of gauze contaminated with bacteria and fungi. *Indian Journal of Medical Microbiology*, Vol.25, No.4, (October 2007), pp. 428-429, ISSN 0255-0857
- Carrol, D.E. & Lopez, A. (1969). Lethality of radio-frequency energy upon microorganisms in liquid, buffered, and alcoholic food systems. *Journal of Food Science*, Vol.34, No.4, (July 1969), pp. 320-324, ISSN 1750-3841

- Chandra, J.; Mukherjee, P.K.; Leidich, S.D.; Faddoul, F.F.; Hoyer, L.L.; Douglas, L.J. & Ghannoum, M.A. (2001). Antifungal resistance of candidal biofilms formed on denture acrylic in vitro. *Journal of Dental Research*, Vol.80, No.3, (March 2001), pp. 903-908, ISSN 0022-3913
- Chau, V.B.; Saunders, T.R.; Pimsler, M. & Elfring, D.R. (1995). In-depth disinfection of acrylic resins. *The Journal of Prosthetic Dentistry*, Vol.74, No.3, (September 1995), pp. 309-313, ISSN 0022-3913
- Clifford, T.J. & Burnett, C.A. (1995). The practice of consultants in restorative dentistry (UK) in routine infection control for impressions and laboratory work. *The European Journal of Prosthodontics and Restorative Dentistry*, Vol.3, No.4, (June 1995), pp. 175-177, ISSN 0965-7452
- Codino, R.J. & Marshall, W.E. (1976). Control of infection in the dental operator. *Dental Survey*, Vol.52, No.5, (May 1976), pp. 42-50, ISSN 0011-8788
- Cole, E.C. & Robison, R. (1996). Test methodology for evaluation of germicides. In: *Handbook of Disinfectants and Antiseptics*. J.M. Ascenzi, 1-13, Marcel Dekker, ISBN 0-8247-9524-5, New York, United States
- Colombo, A.L.; Nucci, M.; Salomão, R.; Branchini, M.L.; Richtmann, R.; Derossi, A. & Wey, S.B. (1999). High rate of non-*albicans* candidemia in Brazilian tertiary care hospitals. *Diagnostic Microbiology and Infectious Disease*, Vol.34, No.4, (August 1999), pp. 281-286, ISSN 0732-8893
- Consani, R.L.; Mesquita, M.F.; de Arruda Nobilo, M.A. & Henriques, G.E. (2007). Influence of simulated microwave disinfection on complete denture base adaptation using different flask closure methods. *The Journal of Prosthetic Dentistry*, Vol.97, No.3, (March 2007), pp. 173-178, ISSN 0022-3913
- Consani, R.L.; Mesquita, M.F.; Zampieri, M.H.; Mendes, W.B. & Consani, S. (2008). Effect of the simulated disinfection by microwave energy on the impact strength of the tooth/acrylic resin adhesion. *The Open Dentistry Journal*, Vol.2, (January 2008), pp. 3-7, ISSN 1874-2106
- Consani, R.L.; Vieira, E.B.; Mesquita, M.F.; Mendes, W.B. & Arioli-Filho, J.N. (2008). Effect of microwave disinfection on physical and mechanical properties of acrylic resins. *Brazilian Dental Journal*, Vol.19, No.4, pp. 348-353, ISSN 0103-6440
- Cottone, J.A.; Tererhalmy, G.T. & Molinari, J.A. (1991). *Practical Infection Control in Dentistry* (1st Edition), Lea & Febiger, ISBN 0683021389, Malvern, PA
- Culkin, K.A. & Fung, D.Y.C. (1975). Destruction of *Escherichia coli* and *Salmonella typhimurium* in microwave-cooked soups. *Journal of Milk and Food Technology*, Vol.38, No.1, pp. 8-15, ISSN 0022-2747
- Davis, D.R.; Curtis, D.A. & White, J.M. (1989). Microwave irradiation of contaminated dental casts. *Quintessence International*, Vol.20, No.8, (August 1989), pp. 583-585, ISSN 0033-6572
- De Gee, A.J.; Ten Harkel, E.C. & Davidson, C.L. (1979). Measuring procedure for the determination of the three-dimensional shape of dentures. *The Journal of Prosthetic Dentistry*, Vol.42, No.2, (August 1979), pp. 149-53, ISSN 0022-3913

- Dixon, D.L.; Breeding, L.C. & Faler, T.A. (1999). Microwave disinfection of denture base materials colonized with *Candida albicans*. *The Journal of Prosthetic Dentistry*, Vol.81, No.2, (February 1999), pp. 207-214, ISSN 0022-3913
- Dovigo, L.N.; Pavarina, A.C.; Ribeiro, D.G.; de Oliveira, J.A.; Vergani, C.E. & Machado, A.L. (2009). Microwave disinfection of complete dentures contaminated in vitro with selected bacteria. *Journal of Prosthodontics*, Vol.18, No.7, (October 2009), pp. 611-617, ISSN 1059-941X
- Egusa, H.; Watamoto, T.; Abe, K.; Kobayashi, M.; Kaneda, Y.; Ashida, S.; Matsumoto, T. & Yatani, H. (2008). An analysis of the persistent presence of opportunistic pathogens on patient-derived dental impressions and gypsum casts. *The International Journal of Prosthodontics*, Vol.21, No.1, (February 2008), pp. 62-68, ISSN 0893-2174
- Fais, L.M.; Pinelli, L.A.; Adabo, G.L.; Silva, R.H.; Marcelo, C.C. & Guaglianoni, D.G. (2009). Influence of microwave sterilization on the cutting capacity of carbide burs. *Journal of Applied Oral Science*, Vol.17, No.6, (December 2009), pp. 584-589, ISSN 1678-7757
- Fitzpatrick, J.A.; Kwao-Paul, J. & Massey, J. (1978). Sterilization of bacteria by means of microwave heating. *Journal of Clinical Engineering*, Vol.3, No.1, (March 1978), pp. 44-47, ISSN 0363-8855
- Friedrich Jr, E.G. & Phillips, L.E. (1988). Microwave sterilization of *Candida* on underwear fabric. A preliminary report. *The Journal of Reproductive Medicine*, Vol.33, No.5, (May 1988), pp. 421-422, ISSN 0024-7758
- Glass, R.T.; Bullard, J.W.; Hadley, C.S.; Mix, E.W. & Conrad, R.S. (2001a). Partial spectrum of microorganisms found in dentures and possible disease implications. *The Journal of the American Osteopathic Association*, Vol.101, No.2, (February 2001), pp. 92-94, ISSN 1945-1997
- Glass, R.T.; Goodson, L.B.; Bullard, J.W. & Conrad, R.S. (2001b). Comparison of the effectiveness of several denture sanitizing systems: a clinical study. *Compendium of Continuing Education in Dentistry*, Vol.22, No.12, (December 2001), pp. 1093-1096, 1098, 1100-2 passim; quiz 1108, ISSN 1548-8578
- Gonçalves, A.R.; Machado, A.L.; Giampaolo, E.T.; Pavarina, A.C. & Vergani, C.E. (2006). Linear dimensional changes of denture base and hard chair-side relining resins after disinfection. *Journal of Applied Polymer Science*, Vol.102, No.2, (October 2006), pp. 1821-1826, ISSN 0021-8995
- Henderson, C.W.; Schwartz, R.S.; Herbold, E.T. & Mayhew R.B. (1987). Evaluation of the barrier system, an infection control system for the dental laboratory. *The Journal of prosthetic dentistry*, Vol.58, No.4, (October 1987), p. 517-521, ISSN 1097-6841
- Hersek, N.; Canay, S.; Akça, K. & Ciftçi, Y. (2002). Tensile strength of type IV dental stones dried in a microwave oven. *The Journal of Prosthetic Dentistry*, Vol.87, No.5, (May 2002), pp. 499-502, ISSN 1097-6841
- Hiti, K.; Walochnik, J.; Faschinger, C.; Haller-Schober, E.M. & Aspöck, H. (2001). Microwave treatment of contact lens cases contaminated with *Acanthamoeba*. *Cornea*, Vol.20, No.5, (July 2001), pp. 467-470, ISSN 1536-4798
- Hoffman, P.N. & Hanley, M.J. (1994). Assessment of a microwave-based clinical waste decontamination unit. *The Journal of Applied Bacteriology*, Vol.77, No.6, (December 1994), pp. 607-612, ISSN 0021-8847

- Ilbay, S.G.; Güvener, S. & Alkumru, H.N. (1994). Processing dentures using a microwave technique. *Journal of Oral Rehabilitation*, Vol.21, No.1, (January 1994), pp.103-109, ISSN 1365-2842
- Infection control in the dental office. Council on Dental Materials and Devices. Council on Dental Therapeutics. (1978). *Journal of the American Dental Association*, Vol.97, No.4, (October 1978), pp. 673-677, ISSN 0002-8177
- Jasnow, S.B. & Smith, J.L. (1975). Microwave sanitization of color additives used cosmetics: feasibility study. *Applied Microbiology*, Vol.30, No.2, (August 1975), pp. 205-211, ISSN 0003-6919
- Jeng, D.K.; Kaczmarek, K.A.; Woodworth, A.G. & Balasky, G. (1987). Mechanism of microwave sterilization in the dry state. *Applied and Environmental Microbiology*, Vol.53, No.9, (September 1987), pp. 2133-2137, ISSN 1098-5336
- Jorge, J.H.; Giampaolo, E.T.; Vergani, C.E.; Pavarina, A.C.; Machado, A.L. & Carlos, I.Z. (2009) Effect of microwave postpolymerization treatment and of storage time in water on the cytotoxicity of denture base and reline acrylic resins. *Quintessence International*, Vol.40, No.10, (November-December 2009), pp. e93-100, ISSN 0033-6572
- Kahler, H. (1929). The nature of the effect of a high-frequency electric field upon *Paramecium*. *Public Health Reports*, Vol.44, No.7, (February 1929), pp. 339-347, ISSN 0033-3549
- Kahn, R.C.; Lancaster, M.V. & Kate, Jr.W. (1982). The microbiologic crosscontamination of dental prostheses. *The Journal of Prosthetic Dentistry*, Vol.47, No.5, (May 1982), pp. 556-559, ISSN 1097-6841
- Kohn, W.G.; Harte, J.A.; Malvitz, D.M.; Collins, A.S.; Cleveland, J.L. & Eklund, K.J. (2004). Centers for Disease Control and Prevention. Guidelines for infection control in dental health care settings-2003. *The Journal of the American Dental Association*, Vol.135, No.1, (January 2004), pp. 33-47, ISSN 0002-8177
- Lamb, D.J.; Ellis, B. & Priestley, D. (1983). The effects of process variables on levels of residual monomer in autopolymerizing dental acrylic resin. *Journal of Dentistry*, Vol.11, No.1, (March 1983), pp. 80-88, ISSN 0300-5712
- Latimer, J.M. (1977). Microwave oven irradiation as a method for bacterial decontamination in a clinical microbiology laboratory. *Journal of Clinical Microbiology*, Vol.6, No.4, (October 1977), pp. 340-342, ISSN 0095-1137
- Leung, R.L. & Schonfeld, S.E. (1983). Gypsum casts as a potential source of microbial cross-contamination. *The Journal of Prosthetic Dentistry*, Vol.49, No.2, (February 1983), pp. 210-211, ISSN 1097-6841
- Luebke, R.J. & Schneider, R.L. (1985). Microwave oven drying of artificial stone. *The Journal of Prosthetic Dentistry*, Vol.53, No.2, (February 1985), pp. 261-265, ISSN 1097-6841
- Luebke, R.J. & Chan, K.C. (1985). Effect of microwave oven drying on surface hardness of dental gypsum products. *The Journal of Prosthetic Dentistry*, Vol.54, No.3, (September 1985), pp. 431-435, ISSN 1097-6841
- Machado, A.L.; Breeding, L.C. & Puckett, A.D. (2005). Effect of microwave disinfection on the hardness and adhesion of two resilient liners. *The Journal of Prosthetic Dentistry*, Vol.94, No.2, (August 2005), pp. 183-189, ISSN 1097-6841

- Machado, A.L.; Breeding, L.C. & Puckett, A.D. (2006). Effect of microwave disinfection procedures on torsional bond strengths of two hard chairside denture relined materials. *Journal of Prosthodontics*, Vol.15, No.6, (November-December 2006), pp. 337-344, ISSN 1532-849X
- Machado, A.L.; Breeding, L.C.; Vergani, C.E. & da Cruz Perez, L.E. (2009). Hardness and surface roughness of relined and denture base acrylic resins after repeated disinfection procedures. *The Journal of Prosthetic Dentistry*, Vol.102, No.2, (August 2009), pp. 115-122, ISSN 1097-6841
- Meşe, A. & Meşe, S. (2007). Effect of microwave energy on fungal growth of resilient denture liner material. *Biotechnology & Biotechnological Equipment*, Vol.21, No.1, (January 2007), pp. 91-93, ISSN 1310-2818
- Meunier-Carpentier, F.; Kiehn, T.E. & Armstrong, D. (1981). Fungemia in the immunocompromised host. *The American Journal of Medicine*, Vol.71, No.3, (September 1981), pp. 363-370, ISSN 0002-9343
- Mima, E.G.; Pavarina, A.C.; Neppelenbroek, K.H.; Vergani, C.E.; Spolidorio, D.M. & Machado, A.L. (2008). Effect of different exposure times of microwave irradiation on the disinfection of a hard chairside relined resin. *Journal of Prosthodontics*, Vol.17, No.4, (June 2008), pp. 312-317, ISSN 1532-849X
- Monsenego, P. (2000). Presence of microorganisms on the fitting denture complete surface: study in vivo. *Journal of Oral Rehabilitation*, Vol.27, No.8, (August 2000), pp. 708-713, ISSN 1365-2842
- Najdovski, L.; Dragas, A.Z. & Kotnik, V. (1991). The killing activity of microwaves on some non-sporogenic and sporogenic medically important bacterial strains. *The Journal of hospital infection*, Vol.19, No.4, (December 1991), pp. 239-247, ISSN 0195-6701
- Nelson-Filho, P.; Faria, G.; da Silva, R.A.; Rossi, M.A. & Ito IY. (2006). Evaluation of the contamination and disinfection methods of toothbrushes used by 24- to 48-month-old children. *Journal of Dentistry for Children*, Vol.73, No.3, (September-December 2006), pp. 152-158, ISSN 1935-5068
- Nelson-Filho, P.; da Silva, L.A.; da Silva, R.A.; da Silva, L.L.; Ferreira, P.D. & Ito, I.Y. (2011). Efficacy of microwaves and chlorhexidine on the disinfection of pacifiers and toothbrushes: an in vitro study. *Pediatric Dentistry*, Vol.33, No.1, (January-February 2011), pp. 10-13, ISSN 0164-1263
- Neppelenbroek, K.H.; Pavarina, A.C.; Spolidorio, D.M.; Vergani, C.E.; Mima, E.G. & Machado, A.L. (2003). Effectiveness of microwave sterilization on three hard chairside relined resins. *The International Journal of Prosthodontics*, Vol.16, No.6, (November-December 2003), pp. 616-620, ISSN 0893-2174
- Neppelenbroek, K.H.; Pavarina, A.C.; Palomari Spolidorio, D.M.; Sgavioli Massucato, E.M.; Spolidorio, L.C. & Vergani, C.E. (2008). Effectiveness of microwave disinfection of complete dentures on the treatment of *Candida*-related denture stomatitis. *Journal of Oral Rehabilitation*, Vol.35, No.11, (November 2008), pp. 836-846, ISSN 1365-2842
- Newton, A.V. (1962). Denture sore mouth. A possible etiology. *British Dental Journal*, Vol.112, No.1, (May 1962), pp. 357-360, ISSN 0007-0610

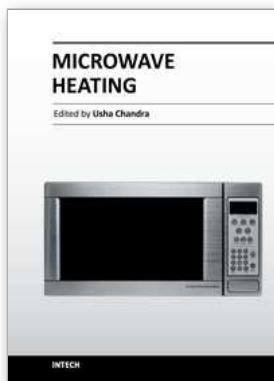
- Nikawa, H.; Iwanaga, H.; Kameda, M. & Hamada T. (1992). In vitro evaluation of *C. albicans* adherence to soft denture-lining materials. *The Journal of Prosthetic Dentistry*, Vol.68, No.5, (November 1992), pp. 804-808, ISSN 1097-6841
- Novais, P.M.; Giampaolo, E.T.; Vergani, C.E.; Machado, A.L.; Pavarina, A.C. & Jorge, J.H. (2009). The occurrence of porosity in reline acrylic resins. Effect of microwave disinfection. *Gerodontology*, Vol.26, No.1, (March 2009), pp. 65-71, ISSN 1741-2358
- Nunes de Mello, J.A.; Braun, K.O.; Rached, R.N. & Del Bel Cury, A.A. (2003). Reducing the negative effects of chemical polishing in acrylic resins by use of an additional cycle of polymerization. *The Journal of Prosthetic Dentistry*, Vol.89, No.6, (June 2003), pp. 598-602, ISSN 1097-6841
- Olsen, C.M. (1965). Microwaves inhibit bread mold. *Food Engineering*, Vol.37, No.7, (1965), pp. 51-53, ISSN 0260-8774
- Pavan, S.; Arioli Filho, J.N.; Dos Santos, P.H. & Mollo, F.deA.Jr. (2005). Effect of microwave treatments on dimensional accuracy of maxillary acrylic resin denture base. *Brazilian Dental Journal*, Vol.16, No.2, (May-August 2005), pp. 119-123, ISSN 1806-4760
- Pavan, S.; Arioli Filho, J.N.; Dos Santos, P.H.; Nogueira, S.S. & Batista A.U. (2007). Effect of disinfection treatments on the hardness of soft denture liner materials. *Journal of prosthodontics*, Vol.16, No.2, (March-April 2007), pp. 101-106, ISSN 1532-849X
- Pavarina, A.C.; Neppelenbroek, K.H.; Guinesi, A.S.; Vergani, C.E.; Machado, A.L. & Giampaolo, E.T. (2005). Effect of microwave disinfection on the flexural strength of hard chairside reline resins. *Journal of Dentistry*, Vol.33, No.9, (October 2005), pp. 741-748, ISSN 1879-176X
- Pelczar, Jr.M.J.; Chan, E.C.S. & Krieg, N.R. (1993). Control of 1 microorganisms: principles 2 and physical agents. In: *Microbiology: concepts and applications*, McGraw-Hill, pp. 200-220, ISBN 0071129146, New York
- Polyzois, G.L.; Zissis, A.J. & Yannikakis, S.A. (1995). The effect of glutaraldehyde and microwave disinfection on some properties of acrylic denture resin. *The International Journal of Prosthodontics*, Vol.8, No.2, (March-April 1995), pp. 150-154, ISSN 0893-2174
- Ponne, C.T. & Bartels, P.V. (1995). Interaction of electromagnetic energy with biological material - relation to food processing. *Radiation Physics and Chemistry*, Vol.45, No.4, (April 1995), pp. 591-607, ISSN 0969-806X
- Powell, G.L.; Runnells, R.D.; Saxon, B.A. & Whisenant, B.K. (1990). The presence and identification of organisms transmitted to dental laboratories. *The Journal of Prosthetic Dentistry*, Vol.64, No.2, (August 1990), pp. 235-237, ISSN 0022-3913
- Ray, K.C. & Fuller, M.L. (1970). Isolation of *Mycobacterium* from dental impression material. *The Journal of Prosthetic Dentistry*, Vol.24, No.3, (September 1970), pp. 335-338, ISSN 0022-3913
- Ribeiro, D.G.; Pavarina, A.C.; Machado, A.L.; Giampaolo, E.T. & Vergani, C.E. (2008). Flexural strength and hardness of reline and denture base acrylic resins after different exposure times of microwave disinfection. *Quintessence International*, Vol.39, No.10, (November 2008), pp. 833-840, ISSN 0033-6572

- Ribeiro, D.G.; Pavarina, A.C.; Dovigo, L.N.; Palomari Spolidorio, D.M.; Giampaolo, E.T. & Vergani, C.E. (2009). Denture disinfection by microwave irradiation: a randomized clinical study. *Journal of Dentistry*, Vol.37, No.9, (September 2009), pp. 666-672, ISSN 0300-5712
- Ribeiro, R.C.; Giampaolo, E.T.; Machado, A.L.; Vergani, C.E. & Pavarina, A.C. (2008). Effect of microwave disinfection on the bond strength of denture teeth to acrylic resins. *The International Journal of Adhesion and Adhesives*, Vol.28, No.6, (September 2008), pp. 296-301, ISSN 0143-7496
- Ribeiro, R.C.; Izumida, F.E.; Moffa, E.B.; Basso, M.F.M.; Giampaolo, E.T.; & Vergani, C.E. (2009). Color stability of reline resin after microwave disinfection: clinical evaluation. *Journal of Dental Research*, Vol.88, Special Issue Letter A, Abstract number 1755, ISSN 0022-0345.
- Rizzo, R. (1993). The effects of sterilization with microwaves on diamond burs. *Minerva Stomatologica*, Vol.42, No.3, (March 1993), pp. 93-96, ISSN 0026-4970
- Rohrer, M.D.; Bulard, R.A. (1985). Microwave sterilization. *Journal of the American Dental Association*, Vol.110, No.2, (February 1985), p.194-198, ISSN 0002-8177
- Rohrer, M.D.; Terry, M.A.; Bulard, R.A.; Graves, D.C. & Taylor, E.M. (1986). Microwave sterilization of hydrophilic contact lenses. *American Journal of Ophthalmology*, Vol.101, No.5, (January 1986), pp. 49-57, ISSN 0002-9394
- Rosaspina, S.; Salvatorelli, G.; Anzanel, D. & Bovolenta, R. (1994). Effect of microwave radiation on *Candida albicans*. *Microbios*, Vol.78, No.314, pp. 55-59, ISSN 0026-2633
- Rossi, T.; Peltonen, R.; Laine, J.; Eerola, E.; Vuopio-Varkila, J. & Kotilainen, P. (1996). Eradication of the long-term carriage of methicillin-resistant *Staphylococcus aureus* in patients wearing dentures: a follow-up of 10 patients. *The Journal of Hospital Infection*, Vol.34, No.4, (December 1996), pp. 311-320, ISSN 0195-6701
- Sande, M.A.; Gadot, F. & Wenzel, R.P. (1975). Point source epidemic of *Mycoplasma pneumoniae* infection in a prosthodontics laboratory. *The American Review of Respiratory Disease*, Vol.112, No.2, (August 1975), pp. 213-217, ISSN 0003-0805
- Sanitá, P.V.; Vergani, C.E.; Giampaolo, E.T.; Pavarina, A.C. & Machado, A.L. (2009). Growth of *Candida* species on complete dentures: effect of microwave disinfection. *Mycoses*, Vol.52, No.2, (March 2009), pp. 154-160, ISSN 0933-7407
- Sanitá, P.V.; Vergani, C.E.; Machado, A.L.; Giampaolo, E.T. & Pavarina, A.C. (2010). Denture microwave disinfection for reducing *Candida* infection in diabetic patients. *Journal of Dental Research*, Vol.89, Special Issue Letter B, Abstract number 1798, ISSN 0022-0345
- Sanitá, P.V.; Machado, A.L.; Dovigo, L.N.; Giampaolo, E.T.; Pavarina, A.C. & Vergani, C.E. (2011). Denture microwave disinfection in treating diabetics with denture stomatitis. *Journal of Dental Research*, Vol.90, Special Issue Letter A, Abstract number 2416, ISSN 0022-0345
- Sartori, E.A.; Schmidt, C.B.; Walber, L.F. & Shinkai, R.S. (2006). Effect of microwave disinfection on denture base adaptation and resin surface roughness. *Brazilian Dental Journal*, Vol.17, No.3, (2006), pp. 195-200, ISSN 0103-6440

- Sartori, E.A.; Schmidt, C.B.; Mota, E.G.; Hirakata, L.M. & Shinkai, R.S. (2008). Cumulative effect of disinfection procedures on microhardness and tridimensional stability of a poly(methyl methacrylate) denture base resin. *Journal of Biomedical Materials Research. Part B, Applied Biomaterials*, Vol. 86B, No.2, (August 2008), pp. 360-364, ISSN 1552-4973
- Seó, R.S.; Vergani, C.E.; Giampaolo, E.T.; Pavarina, A.C. & Machado, A.L. (2007a). Effect of a post-polymerization treatment on the flexural strength and Vickers hardness of relined and acrylic denture base resins. *Journal of Applied Oral Science*, Vol.15, No.6, (December 2007), pp. 506-511, ISSN 1678-7757
- Seó, R.S.; Vergani, C.E.; Pavarina, A.C.; Compagnoni, M.A. & Machado, A.L. (2007b). Influence of microwave disinfection on the dimensional stability of intact and relined acrylic resin denture bases. *The Journal of Prosthetic Dentistry*, Vol.98, No.3, (September 2007), pp. 216-223, ISSN 0022-3913
- Seó, R.S.; Vergani, C.E.; Giampaolo, E.T.; Pavarina, A.C.; dos Santos Nunes Reis, J.M. & Machado, A.L. (2008). Effect of disinfection by microwave irradiation on the strength of intact and relined denture bases and the water sorption and solubility of denture base and relined materials. *Journal of Applied Polymer Science*, Vol.107, No.1, (January 2008), pp. 300-308, ISSN 1097-4628
- Sideridou, I.; Achilias, D.S. & Kyrikou, E. (2004). Thermal expansion characteristics of light-cured dental resins and resin composites. *Biomaterials*, Vol.25, No.15, (July 2004), pp. 3087-3097, ISSN 0142-9612
- Silva, M.M.; Vergani, C.E.; Giampaolo, E.T.; Neppelenbroek, K.H.; Spolidorio, D.M.P. & Machado, A.L. (2006). Effectiveness of microwave irradiation on the disinfection of complete dentures. *The International Journal of Prosthodontics*, Vol.19, No.3, (May-June 2006), pp. 151-156, ISSN 0893-2174
- Silva, M.M.; Vergani, C.E.; Mima, E.G.O.; Pavarina, A.C.; Machado, A.L. & Giampaolo, E.T. (2008). Effect of microwave disinfection in the treatment of denture stomatitis. *Journal of Dental Research*, Vol.87, Special Issue Letter B, Abstract number 1276, ISSN 0022-0345
- Sofou, A.; Larsen, T.; Fiehn, N.E. & Owall, B. (2002). Contamination level of alginate impressions arriving at a dental laboratory. *Clinical Oral Investigations*, Vol.6, No.3, (September 2002), pp. 161-165, ISSN 1432-6981
- Spolidorio, D.; Tardivo, T.; dos Reis Derceli, J.; Neppelenbroek, K.; Duque, C.; Spolidorio, L. & Pires, J. (2011). Evaluation of two alternative methods for disinfection of toothbrushes and tongue scrapers. *International Journal of Dental Hygiene*, (February 2011), doi: 10.1111/j.1601-5037.2011.00503.x. [Epub ahead of print], ISSN 1601-5029
- Takahashi, Y.; Chai, J. & Kawaguchi, M. (1998). Effect of water sorption on the resistance to plastic deformation of a denture base material relined with four different denture relined materials. *The International Journal of Prosthodontics*, Vol.11, No.1, (January-February 1998), pp. 49-54, ISSN 0893-2174
- Tarantino, L.; Tomassini, E.; Petti, S. & Simonetti D'Arca, A. (1997). Use of a microwave device for dental instrument sterilization: possibilities and limitations. *Minerva Stomatologica*, Vol.46, No.10, (October 1997), pp. 561-566, ISSN 0026-4970

- Tate, W.H.; Goldschmidt, M.C.; Ward, M.T. & Grant, R.L. (1995). Disinfection and sterilization of composite polishing instruments. *American Journal of Dentistry*, Vol.8, No.5, (October 1995), pp. 270-272, ISSN 0894-8275
- Tate, W.H.; Goldschmidt, M.C. & Powers, J.M. (1996). Performance of composite finishing and polishing instruments after sterilization. *American Journal of Dentistry*, Vol.9, No.2, (April 1996), pp. 61-64, ISSN 0894-8275
- Thomas, C.J. & Webb, B.C. (1995). Microwaving of acrylic resin dentures. *The European Journal of Prosthodontics and Restorative Dentistry*, Vol.3, No.4, (June 1995), pp. 179-182, ISSN 0965-7452
- Tuncer, N.; Tufekçioğlu, H.B. & Calikkocaoglu, S. (1993). Investigation on the compressive strength of several gypsum products dried by microwave oven with different programs. *The Journal of Prosthetic Dentistry*, Vol.69, No.3, (March 1993), pp. 333-339, ISSN 0022-3913
- Vergani, C.E.; Giampaolo, E.T.; Pavarina, A.C.; Silva, M.M.; Mima, E.G.O. & Machado, A.L. (2008). Efficacy of denture microwave disinfection in treating denture stomatitis. *Journal of Dental Research*, Vol.87, Special Issue Letter B, Abstract number 1289, ISSN 0022-0345.
- Verran, J.; Kossar, S. & McCord, J.F. (1996). Microbiological study of selected risk areas in dental technology laboratories. *Journal of Dentistry*, Vol.24, No.1-2, (January-March 1996), pp. 77-80, ISSN 0300-5712
- Verran, J. & Maryan, C.J. (1997). Retention of *Candida albicans* on acrylic resin and silicone of different surface topography. *The Journal of Prosthetic Dentistry*, Vol.77, No.5, (May 1997), pp. 535-539, ISSN 0022-3913
- Wakefield, C.W. (1980). Laboratory contamination of dental prostheses. *The Journal of Prosthetic Dentistry*, Vol.44, No.2, (August 1980), pp. 143-146, ISSN 0022-3913
- Watanabe, K.; Kakita, Y.; Kashige, N.; Miake, F. & Tsukiji, T. (2000). Effect of ionic strength on the inactivation of micro-organisms by microwave irradiation. *Letters in Applied Microbiology*, Vol.31, No.1, (July 2000), pp. 52-56, ISSN 0266-8254
- Webb, B.C.; Thomas, C.J.; Harty, D.W. & Willcox, M.D. (1998). Effectiveness of two methods of denture sterilization. *Journal of Oral Rehabilitation*, Vol.25, No.6, (June 1998), pp. 416-423, ISSN 0305-182X
- Webb, B.C.; Thomas, C.J. & Whittle, T. (2005). A 2-year study of *Candida*-associated denture stomatitis treatment in aged care subjects. *Gerodontology*, Vol.22, No.3, (September 2005), pp. 168-176, ISSN 0734-0664
- Williams, H.N.; Falkler, W.A.Jr.; Hasler, J.F. & Libonati, J.P. (1985). The recovery and significance of nonoral opportunistic pathogenic bacteria in dental laboratory pumice. *The Journal of Prosthetic Dentistry*, Vol.54, No.5, (November 1985), pp. 725-730, ISSN 0022-3913
- Woo, I.; Rhee, I. & Park, H. (2000). Differential damage in bacterial cells by microwave radiation on the basis of cell wall structure. *Applied and Environmental Microbiology*, Vol.66, No.5, (May 2000), pp. 2243-2237, ISSN 0099-2240
- Wright, P.S.; Clark, P. & Hardie, J.M. (1985). The prevalence and significance of yeasts in persons wearing complete dentures with soft-lining materials. *Journal of Dental Research*, Vol.64, No.2, (February 1985), pp. 122-125, ISSN 0022-0345

- Yannikakis, S.; Zissis, A.; Polyzois, G. & Andreopoulos, A. (2002). Evaluation of porosity in microwave-processed acrylic resin using a photographic method. *The Journal of Prosthetic Dentistry*, Vol.87, No.6, (June 2002), pp. 613-619, ISSN 0022-3913
- Yeo, C.B.; Watson, I.A.; Stewart-Tull, D.E. & Koh, V.H. (1999). Heat transfer analysis of *Staphylococcus aureus* on stainless steel with microwave radiation. *Journal of Applied Microbiology*, Vol.87, No.3, (September 1999), pp. 396-401, ISSN 1364-5072



Microwave Heating

Edited by Dr. Usha Chandra

ISBN 978-953-307-573-0

Hard cover, 370 pages

Publisher InTech

Published online 27, July, 2011

Published in print edition July, 2011

The Microwave heating has not only revolutionized the food industry but also has extended its wings widely towards its multidimensional applications. Thus it has opened new vistas of potential research in science and technology. The book is compiled into Seventeen Chapters highlighting different aspects varying from epistemological discussion to applicability of conceptual constructs. The inclusion of discussion on the avenues in the field of Chemistry, Health & Environment, Medical Sciences and Technology makes it an exquisite work for the aspirant Researchers. As the text book for the beginners, it is designed fundamentally to be a reference monograph to the experts providing a passage for future research. The plethora of literatures are available on Microwave Applications but they seldom direct their readers to concentrate on the key aspects behind the success in microwave applications in different fields. Here is the attempt to fill up the gap with this book.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Carlos Vergani, Daniela Garcia Ribeiro, Livia Nordi Dovigo, Paula Volpato Sanita and Ana Claudia Pavarina (2011). Microwave assisted disinfection method in dentistry, Microwave Heating, Dr. Usha Chandra (Ed.), ISBN: 978-953-307-573-0, InTech, Available from: <http://www.intechopen.com/books/microwave-heating/microwave-assisted-disinfection-method-in-dentistry>

INTECH

open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the [Creative Commons Attribution-NonCommercial-ShareAlike-3.0 License](#), which permits use, distribution and reproduction for non-commercial purposes, provided the original is properly cited and derivative works building on this content are distributed under the same license.