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# Novel Insights Into the Role of Inflammation in Promoting Breast Cancer Development

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## 1. Introduction

In the past decades the major focus of cancer research has been the transformed tumor cells itself, while the role of cellular microenvironment in tumorigenesis has not been widely explored. Several studies have demonstrated the ability of stroma to regulate the growth and differentiation state of breast cancer cells, and the invasive behaviour, and polarity of normal mammary epithelial and breast carcinomas are influenced by tumor microenvironment, immune and stromal cells (Bissell, et al., 2002, Radisky & Radisky, 2007, Tlsty, 2001, Tlsty & Hein, 2001). In addition, genetic abnormalities, such as loss of heterozygosity, occur not only in cancer cells, but in stromal cells as well (Kurose, et al., 2002, Kurose, et al., 2001, Moinfar, et al., 2000).

It is believed that a better understanding of the tumor microenvironment could help render more accurate diagnostics or assist in predicting tumor aggressiveness (i.e., bad prognosis) thus facilitating the design of personalized treatments.

By the end of the nineteenth century, the English surgeon S. Paget suggested the idea that, in order for breast cancer to develop, a specific “seeding” process must occur and, for this primary onset to metastasize to a specific distant organ, particular stromal features would be required postulating his “seed and soil” hypothesis (Paget, 1889). His work greatly contributed to somewhat earlier observations by T. Langhans who first used the word stroma to describe the connective tissue, vessels and other components between tumors (Langhans, 1879) and to the theory postulated by R. Virchow suggesting a possible origin of cancer at sites of chronic inflammation (Balkwill & Mantovani, 2001). A century later, researchers such as B. Mintz and K. Illmensee in general, as well as M. Bissell, in breast cancer in particular, pointed to the tumor milieu as an essential component of neoplasias, not only for cancer evolution but also for cancer instigation (Mintz & Illmensee, 1975; Lochter & Bissell, 1995). Together these and additional findings had painted a broad picture of the complexity of tumor microenvironment, where diverse stromal cells interact with

each other and with the cancer cells playing important roles in tumorigenesis (Soto & Sonnenschein, 2004; Egeblad et al., 2010).

It is clear now that metastatic tumors represent the greatest threat to cancer patient mortality. Indeed, when breast cancer is diagnosed early and metastases are not present, 5-year survival is >88%; however, if metastases are also present, long-term survival is significantly diminished (~10%) (Jemal, et al., 2011). Thereby, the major cause of mortality of breast cancer and different types of cancer is due to metastasis to distant organs, such as lung, bone, liver and brain (Lu & Kang, 2007). A notable feature of this process is the variation in metastatic organ tropism displayed by different types of cancer (Chambers, et al., 2002, Fidler, 2002). A classic view has proposed that purely mechanical factors regulate the fate of blood-borne metastasis tumor cells (MacDonald, et al., 2002); however, this does not fully explain the non-random distribution and distinct pattern of metastasis in each tumor type (Lu & Kang, 2007). However, tumor microenvironment has also shown an important role in the regulation of this process (Valdivia-Silva, et al., 2009). A number of different molecules present in the microenvironment have been associated to the metastasis of breast cancer, among them, chemokines, which have been associated with regulation of cell migration and invasion of tumor cells into specific organs (Muller, et al., 2001, Zlotnik, 2006). Chemokines are a superfamily of chemotactic cytokines characterized by their ability to induce directed migration of leukocytes, during haematopoiesis, lymphoid organ development, and in disease (Sallusto, et al., 2000); their expression may be inducible, primarily by pro-inflammatory cytokines such as TNF- $\alpha$  and IL-1- $\beta$  (Ben-Baruch, 2003). Chemokine receptor expression in many cancer cells have shown to be a non-random process (Shields, et al., 2007, Zlotnik, 2006) and to have a role in organ-specific metastasis: for example, CXCR4 expression and metastasis to lung, bone and lymph nodes (Muller, et al., 2001), CCR7 to lymph nodes (Shields, et al., 2007), CX3CR1 to brain (Mourad, et al., 2005), CCR9 to liver and small bowel (Amersi, et al., 2008, Letsch, et al., 2004), and CCR5 and CXCR2 to lung, liver, vessel endothelial cells and bone (Gross & Meier, 2009, Keeley, et al., 2010, Miller, et al., 1998).

Here, we will discuss the ability of the chemokines to affect tumor cell-microenvironment interactions, increasing the invasive behaviour and metastasis, confirming the importance of the host inflammatory response that may differ between tumor types, disease stages, and/or many other host factors; and the role of stromal contribution of the inflammatory microenvironment to cancer progression and metastasis.

## **2. Inflammatory mediators as regulator of breast cancer development and metastasis**

The link between inflammation and cancer has been observed over 150 years ago when Rudolf Virchow noted that cancers tend to occur at sites of chronic inflammation. Indeed, epidemiological studies indicate that inflammatory and infectious diseases are often associated with an increased risk of cancer (Coussens & Werb, 2002). The microenvironment of tumors mimics that of tissues during the height of an inflammatory response to injury (Joyce & Pollard, 2009). However, unlike the organized morphology of normal tissue, and the ultimate resolution of the inflammation that occurs during healing, tumors exist in a state of chronic inflammation characterized by the presence of cancer cells, immune cells, aberrant vascular cells, and the persistence of inflammatory mediators, such as cytokines and chemokines.

The presence and significance of leukocyte infiltrates in developing neoplasms is now undisputed (Allen, et al., 2007, Moser & Loetscher, 2001, Moser & Willimann, 2004). It has been demonstrated that leukocyte infiltration in developing tumors is one of the host's main immune mechanisms to eradicate malignant cells. However, while some leukocytes certainly have this potential, i.e., cytotoxic T lymphocytes (CTLs) and natural killer (NK) cells (Luster, 1998), other leukocyte cell types, most notably innate immune cells, i.e., mast cells (MCs), immature myeloid cells, granulocytes, and macrophages, instead potentiate tumor progression (Baggiolini, et al., 1997, Chen, et al., 2006, Joyce & Pollard, 2009), and enhance neoplastic cell survival. Upon entry into the neoplastic microenvironment, infiltrating leukocytes become alternatively activated and manifest a pro-tumor phenotype as defined by activation of cellular programs involved in immune tolerance and tissue remodelling (Mishra, et al., 2011, Strieter, et al., 2006). During premalignant progression, a consequence of alternative activation of leukocytes is promotion and elaboration of a microenvironment rich in extracellular matrix (ECM) remodelling proteases, and increased presence of pro-survival, pro-growth and pro-angiogenic factors that further enhance proliferative and invasive capacities of neoplastic cells (Li, et al., 2007, Orimo, et al., 2005). Such pro-tumor inflammatory microenvironments promote not only malignant conversion and development of solid tumors, but also dissemination of neoplastic cells into blood vasculature by driving invasive capacity of malignant cells, expansion of angiogenic vasculature, and neoplastic cell entry into blood vessels (and lymphatics) (Keeley, et al., 2010).

Breast carcinomas are highly infiltrated by different types of host leukocytes, including primarily T cells, and monocytes that differentiate into tumor-associated macrophages (TAM) at the tumor site (Ben-Baruch, 2003, Crowther, et al., 2001). The presence of the cellular infiltrate in breast tumors was initially regarded as evidence for the potential activity of immune mechanisms against the growing neoplasm. As explained above, several studies suggest that T-cell antitumor responses are impaired in advanced stages of breast carcinoma, and there is no definite conclusion regarding the efficacy of T-cell-dependent immune mechanisms, or regarding the correlation between the type of T-cell infiltration and tumor progression in most subtypes of breast carcinoma (Hsiao, et al., 2010). The only exception is the relatively infrequent type of medullary carcinoma, in which favourable prognosis was correlated with intensive lymphoid infiltration (Hadden, 1999). In contrast to T lymphocytes, large evidence suggests that high levels of TAM are correlated with poor prognosis in breast carcinoma. Many studies have shown a positive relationship between high levels of TAM and lymph node metastases, and suggested that the density of TAM is associated with clinical aggressiveness (Crowther, et al., 2001, O'Sullivan & Lewis, 1994). Again, the potential contribution of TAM to tumor elimination, in view of several potential antimalignant activities that may be exerted by these cells, such as antigen presentation, cytotoxicity, or/and phagocytosis, was contradictory with the promalignant activities of TAM in breast carcinoma. These promalignant activities of TAM are the result of their ability to express numerous tumor-promoting mediators, such as growth factors for breast tumor cells, angiogenic molecules, ECM degrading enzymes, inflammatory cytokines, and chemokines (Balkwill & Mantovani, 2001, Colotta, et al., 2009). In addition, TAM might contribute to tumor progression by the release of reactive oxygen intermediates, which may induce mutagenic changes that could result in increased DNA damage and generation of new subtypes of cancer cells within the tumor (Colotta, et al., 2009). A major TAM-derived

inflammatory cytokine shown to be highly expressed in breast carcinomas is tumor necrosis factor alpha (TNF- $\alpha$ ) (Leek, et al., 1998), which is a multifactorial cytokine. Tumor necrosis factor alpha was first isolated as an anti-cancer cytokines more than two decades ago (Aggarwal, 2003). However, these effects may depend on multiple factors, such as estrogen therapy and the expression of members of the epidermal growth factor receptor family. The fact that TNF- $\alpha$  activities vary under different physiological conditions and in a cell-type-dependent manner contributes to a sense of ambiguity regarding its antitumor effects (Kanoh, et al., 2001, Offersen, et al., 2002). A number of reports indicate that TNF- $\alpha$  induces cellular transformation, proliferation, and tumor promotion (Balkwill & Mantovani, 2001, Li, et al., 2007). An interesting study reported that human TNF- $\alpha$  is more effective than the chemical tumor promoters okadaic acid and 12-O-tetradecanoylphorbol-13-acetate in inducing cancer (Komori, et al., 1993).

The number of cells expressing TNF- $\alpha$  in inflammatory breast carcinoma has been correlated with increasing tumor grade and node involvement (Ben-Baruch, 2003, Leek, et al., 1998). Furthermore, patients with more progressed tumor phenotypes were shown to have significantly higher TNF- $\alpha$  and IL-2 serum concentration (Tesarová, et al., 2000). The tumor-promoting functions of TNF- $\alpha$  may be mediated by its ability to induce pro-angiogenic functions, to promote the expression of matrix metalloproteinases (MMP) and endothelial adhesion molecules, and to cause DNA damage via reactive oxygen, the overall effect of which is promotion of tumor-related processes (Garg & Aggarwal, 2002).

In addition, several inflammatory interleukins have been linked with carcinogenesis and tumor progression. Among these, IL-6 and IL-1 have been widely studied in breast carcinoma. In different types of cancer, IL-1 promotes growth and confers chemoresistance (Arlt, et al., 2002, Woodworth, et al., 1995). Furthermore, IL-1 secretion into the tumor milieu also induces several angiogenic factors from tumor and stromal cells that promotes tumor growth through hyperneovascularization (Zhou, et al., 2011). IL-6 may act as a paracrine growth factor for multiple myeloma, non-Hodgkin's lymphoma, bladder cancer, colorectal cancer, and renal carcinoma (Angelo, et al., 2002, Landi, et al., 2003, Okamoto, et al., 1995, Voorzanger, et al., 1996). However, contradictory studies suggested that elevated levels of IL-6 might contribute to breast cancer progression (Karczewska, et al., 2000, Kurebayashi, 2000). Initial analyses regarding IL-1b indicated that its levels were significantly higher in invasive carcinoma than in ductal carcinoma *in situ* or in benign lesions, implying that elevated levels of IL-1b are directly correlated with a more advanced disease (Jin, et al., 1997). Of interest is the fact that the two cytokines (IL-6 and IL-1b) and TNF- $\alpha$  are interrelated and may act in an additive manner, suggesting that these three cytokines form a network of related factors that may affect tumor cell progression in a cooperative manner.

Cyclooxygenase (COX)-2, an inducible enzyme with expression regulated by NF- $\kappa$ B, mediates tumorigenesis. COX-2, the inducible isoform of prostaglandin H synthase has been implicated in the growth and progression of a variety of human cancers, and its expression can be induced by various growth factors, cytokines, oncogenes, and other tumor factors. IL-1 has been reported to upregulate COX-2 expression in human colorectal cancer cells via multiple signalling pathways (Liu, et al., 2003). COX-2 is expressed at an intermediate or high level in epithelial cells of invasive breast cancers (Chang, et al., 2005, Half, et al., 2002). Expression of COX-2 in breast cancer correlates with poor prognosis, and COX-2 enzyme inhibitors reduce breast cancer incidence in humans. COX-2 overexpression has also been found in the mammary gland of transgenic mice induced mammary cancer (Kundu & Fulton, 2002).

Hypoxia is also an important cellular stressor that triggers a survival program by which cells attempt to adapt to the new environment. This primarily involves adaptation of metabolism and/or stimulation of oxygen delivery. These cell-rescuing mechanisms can be conducted rapidly by a transcription factor that reacts to hypoxic conditions, the hypoxia-inducible factor-1 (HIF-1a) (Semenza & Wang, 1992). HIF-1a stimulates processes such as angiogenesis, glycolysis, and erythropoiesis (Jiang, et al., 1996) by activating genes that are responsible for these processes. Cancer cells are able to survive and proliferate in extreme microenvironmental conditions and show changes in oncogenes and tumor suppressor genes. Hypoxia and HIF-1a have been implicated in carcinogenesis and in clinical behaviour of tumors. Upregulation of HIF-1a was noted during breast carcinogenesis (Bos, et al., 2001) especially in the poorly differentiated pathway. Hypoxia is related to poor response to therapy in various cancer types. In invasive breast cancer, high HIF-1a concentrations were associated with poor survival in lymph node-negative patients (Bos, et al., 2003). As prognosis in breast cancer is closely related to proliferation rate (van Diest & Baak, 1991) and poorly differentiated tumors usually exhibit high proliferation and HIF-1a overexpression, the prognostic value of HIF-1a might well be explained by a close association between HIF-1a and proliferation. Additionally, HIF-1a has shown to be a master regulator for surviving hypoxia interacting with cell cycle-related proteins. High concentrations of HIF-1a are associated with overexpression of p53 and markers of proliferation during the late S/G2 phase of the cell cycle (Bos, et al., 2004).

### **3. Role of chemokines and their receptors in breast cancer progression and metastasis**

While most evidence presented above suggests that proinflammatory cytokines and enzymes play an important role in mediating tumorigenesis, and tumor progression, the molecular mechanisms of metastasis and its relationship with the organotropism of cancer cell remain unclear. However, recent studies focused on the chemokines and their receptors, and the different interactions with inflammatory cytokines in the tumor microenvironment have provided additional information that might better explain the non-random patterns of organotropism during metastasis, including atypical metastasis to rare organs (Franco-Barraza, et al., 2010, Valdivia-Silva, et al., 2009).

Chemokine activities in different malignancy including breast cancer are mediated primarily by their ability to induce chemotaxis of leukocytes, endothelial cells, and/or the tumor cells. Chemokines induce migration of leukocyte subpopulations to tumor sites that may promote antitumor activities (such as Th1 cells or natural killer cells), while other chemokines are responsible for large quantities of deleterious tumor-associated macrophages (TAM) at tumor sites (Allavena, et al., 2008, Ben-Baruch, 2008, Soria & Ben-Baruch, 2008) as discussed above. Moreover, specific chemokines upregulate endothelial cell migration and proliferation, and promoting angiogenesis, whereas other chemokines have powerful angiostatic properties (Strieter, et al., 2006, Struyf, et al., 2011). Another very important activity of chemokines is induction of tumor cell invasion and migration, thereby playing key roles in dictating site-directed metastasis formation (Ben-Baruch, 2008, Zlotnik, 2006). Chemokines and their receptors can execute such multifaceted roles in malignancy because cells of the tumor microenvironment, and in many cases also by the tumor cells themselves express them. As such, they can affect through autocrine pathways the ability of

the cancer cells to express tumor-promoting functions, and can also act in paracrine manners on host cells, thereby influencing their roles in malignancy.

Breast cancer metastasis is the result of several sequential steps and represents a highly organized, non-random and organ selective process dependent on intricate stroma-stroma interactions at the target organ (Ben-Baruch, 2006, Lu & Kang, 2007), causing high mortality by invasion of vital organs, such as bone, lung, brain and liver. Important evidence suggests that chemokines have an important role in regulating trafficking and metastasis (Bagley, et al., 2010). Indeed, breast cancer cells express chemokine receptors in a non-random manner, and these observations pointed to several chemokine/ receptor pairs that control cell-cell migration (Zlotnik, 2008). Association of chemokine receptors with various cancers including breast carcinoma has been widely documented (Ali & Lazennec, 2007, Karnoub & Weinberg, 2006, Koizumi, et al., 2007, Ruffini, et al., 2007). Accumulative evidence, in particular from clinical retrospective studies, presents a compelling picture indicating that the experimental evidence derived from *in vitro* experiments and animal models pointing to a pivotal role of chemokine receptors in cancer metastasis. CXCR4 and CCR7 are the most widely expressed in many different cancers, and the expression of CXCL12 and CCL21, their specific ligands, respectively, are highest in lung, liver bone marrow for the first one and lymph nodes for both (Nevo, et al., 2004, Schimanski, et al., 2008). Additionally, the expression of CCR7 in patients with several types of cancer has an excellent correlation with the ability of the tumor to spread to the lymph nodes (Takanami, 2003, Wang, et al., 2005). Other chemokine receptors may participate in the regulation of metastasis of specific cancers and in tumor progression. CX3CR1 is involved in homing metastasis to brain for glioblastoma and breast cancer (Andre, et al., 2010, Lavergne, et al., 2003) and to bone and bone marrow endothelial cell for prostate cancer (Shulby, et al., 2004). CCR9/CCL25 axis was found in melanoma (Letsch, et al., 2004), ovarian cancer (Johnson-Holiday, et al., 2007), prostate cancer (Singh, et al., 2004), nasopharyngeal carcinoma (Ou, et al., 2006), acute lymphoblastic leukaemia (Annels, et al., 2004) and probably breast cancer (Johnson-Holiday, et al., 2011); most of the cases are related to metastatic lesions in the gastrointestinal tract included the liver. Additionally, elevated expression levels of CXCR2 and CCR5 and their ligands, CXCL8 and CCL5, respectively, in breast carcinoma and other neoplasias were significant associated with increased malignancy, advance disease, early relapse and poor prognosis (Ben-Baruch, 2006, Yaal-Hahoshen, et al., 2006). Moreover, it has been demonstrated that tumor cells can generate autocrine gradients of ligands of chemokine receptors (i.e., CCR7) that guide their migration in direction of a physiological level of interstitial flow towards functional lymphatics, even if lymphatic endothelial cells are absent; although the effect is greatly amplified when both flow and cells are present (Shields, et al., 2007). This data suggests that the chemokine-chemokine receptor interaction is of particular importance in the metastatic destination of many cancers.

However, a couple of questions are very important to make in this point: Is the chemokine receptor expression in cancer cells constant? Or might the tumor microenvironment or inflammation regulate the chemokine receptor expression in cancer cells? Interestingly, these questions, which are product of logic thinking on the tumor microenvironment, were not made until recently by our group (Valdivia-Silva, et al., 2009). Indeed, the chemokine receptor expression has not been thoroughly studied under inflammatory conditions.

Although there are reports demonstrating that tumor and leukocytes increase expression of chemokines and cytokines during disease progression, it is not clear what are the chemokine

receptors involved in regulation of metastasis. Most of the previously reported studies had focused in analysing chemokine receptors expressed in different neoplasias without evaluating their phenotypic changes and functionality during the progress of the disease (Ben-Baruch, 2008). However, it has not been clearly demonstrated any type of regulation of the microenvironment in these changes. Finally, the chemokine receptors expressed under non-stimulated conditions by cancer cells were considered biomarkers to specific homing to organs, but it does not explained atypical metastasis of cancer to rare organs (Charalabopoulos, et al., 2004, Johnson, 2010, Kilgore, et al., 2007, Saisho, et al., 2005).

Within the tumor microenvironment, chemokines and their receptors play different roles in modulating several functions as described above, and through these processes, help to define the progression of the cancer. Stromal, and immune cells, including leukocytes differentiating into tumor-associated macrophages (TAM) at the tumor site, express numerous promoting factors, such as growth factors, angiogenic mediators, extracellular matrix-degrading enzymes, inflammatory cytokines, and more chemokines (Polyak & Kalluri, 2010). Interestingly, pro-inflammatory cytokines like IL-1, IL-6, IFN- $\gamma$  and TNF- $\alpha$ , which are important modulators of chemokine receptors expression in different tissues, have demonstrated to regulate their expression in cancer cells in a non-random manner (Valdivia-Silva, et al., 2009). Similar to cytokines regulate for CXCR4 and CCR5 in astrocytes (Croitoru-Lamoury, et al., 2003), CXCR2 in human mesangial cells (Schwarz, et al., 2002), and CX3CR1 in smooth muscle cells (Chandrasekar, et al., 2003), synovium (Nanki, et al., 2002), and different epithelial cells (Fujimoto, et al., 2001, Matsumiya, et al., 2001); different doses and times of exposition allowed the expression of specific type of chemokine receptor in several breast cancer cell lines and the change of their phenotypes into more invasiveness ones (Franco-Barraza, et al., 2010).

We have analysed the human breast carcinoma MCF-7 cell line as a model of pre-invasive stage to demonstrate the regulation by an inflammatory microenvironment on chemokine receptor expression and functionality (Valdivia-Silva, et al., 2009). The comparison of the expression of CXCR4, CX3CR1, CXCR2, CCR9 and CCR5 at the transcriptional, protein, and functional levels under two different *in vitro* conditions (basal versus cytokine-stimulation) showed clearly the regulation of the specific cytokine over specific chemokine receptor, independently of the genetic background of MCF-7, which presents very low levels of these receptors under basal conditions. This was also observed in the highly metastatic MDA-MB-231, MDA-MB-361 and in the poorly metastatic T47D breast cancer cell lines; although the levels of expression observed after cytokine stimulation were different than those obtained in the MCF-7 cell line. A direct suggestion of these results, affirms that basal expression of a given chemokine receptor is not by itself a good marker of homing or aggressiveness and is subject to change by the microenvironment. Another important outcome in that work was the absence of correlation between the functionality of the receptor and their expression (gen or protein). For example, an increase in CXCR2 expression in MCF-7 cell line does not correlates with an increase in the migration index. In contrast, CX3CR, induced by TNF- $\alpha$ , had a small but significant increase at the protein level, which had an impact on their chemotactic activity. A considerable increase of chemokine receptors was found in non-migratory cancer cells indicating that that chemokine receptor expression does not necessarily result in migration response to a chemoattractant ligand. It also suggests that only a fraction of the cells have the potential to form metastases and capable to invade different organs. In fact, genetic analysis of the MDA-MB-231 breast cancer cell line subpopulations, obtained from *in vivo* experiments, identified a gene set whose expression



pattern is associated to metastasis to bone but not adrenal medulla (Kirschmann, et al., 1999, Xu, et al., 2010). Interestingly this signature is retained through repeated passage of the metastatic cell population both in vitro and in vivo. Therefore, breast cancer cells with a defined tissue-specific metastatic ability pre-exist in the parental tumor cell population and may have a distinctive metastasis gene expression signature. Thus, these data suggested that inflammatory stimulation in the tumor microenvironment might affect cancer cells migration by different mechanisms. Importantly, not all cancer cell population, including cell lines, had the same behaviour under the same cytokine stimulation. Finally, other important finding in this study suggested that cancer cells require constant inflammatory stimuli by the microenvironment to trigger their invasive and metastatic activity, because of after a short time without stimuli (hours to days), the cells diminished their specific-stimuli chemokine receptor expression.

Altogether, these data allowed us to propose that exist sub-populations expressing different levels of chemokine receptor expression, which under a particular stimuli in the host microenvironment, change their expression levels and thus their aggressiveness. Then, atypical metastasis of breast cancer to others organs, which are relatively rare, could fall under this scheme. The biological inflammatory global response in the tumor microenvironment might be triggering the expression of different chemokine receptors and determining a new homing for these cancer cells. More broadly, these observations strongly support the overall model where chemokines determine the metastatic destinations of cancer cells (Fig 1.)

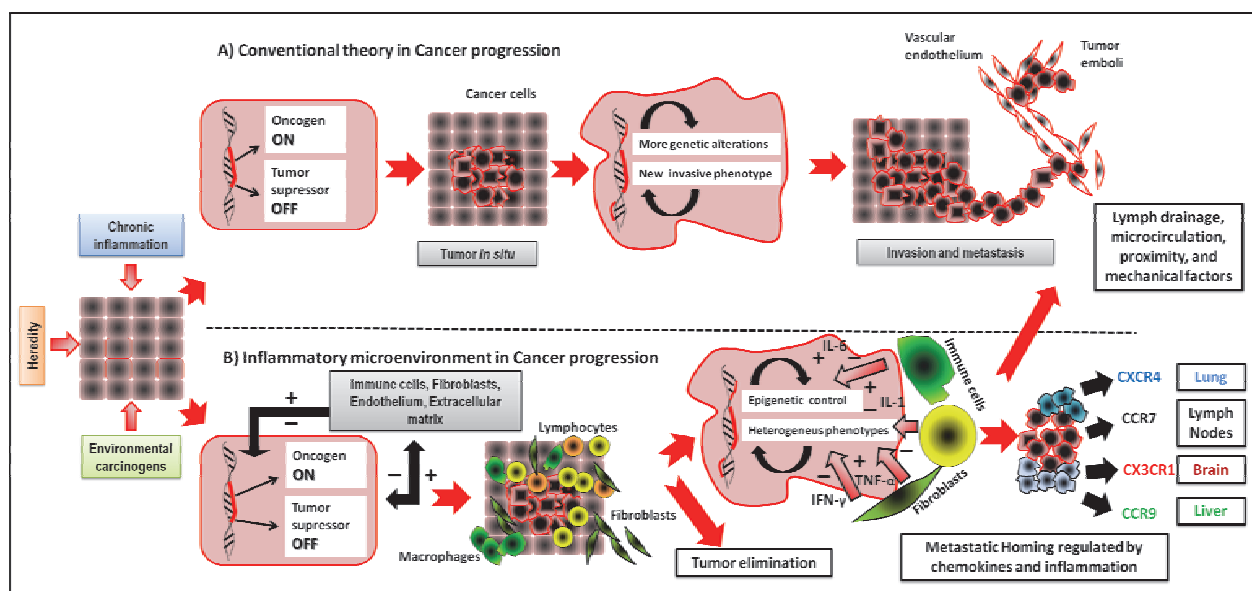


Fig. 1. Microenvironment and cancer progression.

Two theories have been proposed to explain this process, a conventional theory based on genetic alterations and a second view that involves participation of an inflammatory microenvironment. A) Initially, susceptible cells to different carcinogenic factors (e.g., genetic susceptibility obtained by inheritance) suffer specific DNA mutations that trigger tumorigenesis. The conventional theory is focused on the view that cancer progression is initially dependent on a sequence of genetic alterations and, finally, purely mechanical factors regulate the fate of blood-borne metastasis tumour cells (e.g. proximity,

microcirculation, direction of lymph or circulation drainage, etc.). B) A second view, based on the participation of an inflammatory microenvironment, takes into account constant interactions between tumor cells and surrounding cells during the different stages of cancer development. Therefore, the final response is the result of positive and negative effects and not only dependent on internal genetic changes in cancer cells but on interactions and epigenetic control of multiple inflammatory molecules released into the tumor microenvironment. Therefore, the final metastatic homing, which is mediated by expression of chemokines and chemokine receptors, will be dependent on the deregulation of the host immune response

#### **4. Targeting chemokines for breast cancer metastasis**

As a consequence of studies focusing almost exclusively on cancer cells, nearly all of the currently used cancer therapeutic agents target the cancer cells that, due to their inherent genomic instability, frequently acquire therapeutic resistance (Rajagopalan, et al., 2003). In part due to frequent therapeutic failures during the course of treatment of advanced stage tumors, increasing emphasis has been placed on targeting various stromal cells, particularly endothelial cells, via therapeutic interventions. Since these cells are thought to be normal and genetically stable, they are less likely to develop acquired resistance to cancer therapy. Thus, isolating, and characterizing each cell type (epithelial, myoepithelial, and various stromal cells) comprising non-malignant and cancerous breast tissue would not only help us to understand the role these cells play in breast tumorigenesis, but would likely give us new molecular targets for cancer intervention and treatment.

There is now an abundant literature documenting the associations of chemokine receptors with various types of cancer (Zlotnik, 2006) and their importance to mediate the establishment or development of metastatic foci. In fact, some anticancer drugs currently in use -like Herceptin- may involve the downregulation of chemokine receptors as part of their mechanism of action (Li, et al., 2004). This would provide the ultimate validation of the hypothesis, and would also point to future opportunities for therapeutic intervention as we discussed below. Current therapies such as surgery, radiotherapy and chemotherapy are primarily concerned with destruction of cancer. Targeting chemokines and chemokine receptors will allow limiting angiogenesis or metastasis and may enable such therapies to act as chemotherapeutic agents alone or in synergism with conventional agents. The up-regulation of certain chemokine molecules in tumor as compared with normal cells offers a potential avenue – where cancer cells and their metastases can be specifically targeted. This selective destruction of cells is also pre-requisite of non-toxic treatment regimens.

Manipulation of the tumor microenvironment by treatment with chemokines can be used to recruit either immature dendritic cells for the initiation of anti-tumor responses or effector cells for cytotoxic responses. Intratumoral delivery of CCL21 using pox virus vaccine into established tumors derived from murine colon cancer line, CT26 results in enhanced infiltration of CD4 T cells which correlated with inhibition of tumor growth (Flanagan, et al., 2004). Non-immunogenic murine breast carcinoma is rejected after transducing cells with CCL19. The rejection of tumor was mediated by activated NK and CD4+ cells (Braun, et al., 2000). Adenoviral delivery of the CCL16 is able to inhibit growth of mammary tumors and prevent metastatic growth (Okada, et al., 2004). Importantly, in treatment involving delivery of chemokines to the tumor environment, there is a major problem of heterogeneity of the tumor cells. Chemokines may have dual effects, can be beneficial to one patient might be

harmful to another. However, this problem can be circumvented by chemokine typing every tumor prior to deciding on an appropriate therapy regime. They may be used as an adjunct to increase the efficacy of currently available therapies. Targeting specific chemokines can also modulate tumor infiltrating leukocytes or angiogenesis. High CXCL8 expression levels render tumor cells highly tumorigenic, angiogenic and invasive (Chavey, et al., 2007, Freund, et al., 2003, Freund, et al., 2004). In a murine model of breast cancer treatment with Met-CCL5, an antagonist of CCR1 and CCR5 led to a reduction in the total number of infiltrating inflammatory cells, in particular a decrease in macrophage infiltration and reduced growth of tumors (Liang, et al., 2004, Robinson, et al., 2003). The 7-transmembrane structure of chemokine receptors makes them attractive targets for small molecule inhibitors (Seaton, et al., 2009).

In summary, the exploration and manipulation of the chemokine network has just started and is likely to improve efficiency of current tumor therapies. However, since these chemotactic cytokines are also utilized in a plethora of normal interactions, caution is needed especially when extrapolating *in vitro* data into the clinical situation. Differences amongst tumor entities are obvious and the same chemokine/chemokine-receptor system seems to have divergent functions in different tumor entities. A more in-depth analysis of the real players in tumor immunosuppression, for example characterization of the subtypes of infiltrating immune cells and thorough analysis of the cytokine and chemokine milieu of primary tumors, will be necessary to pave the way for more efficient therapeutic interventions.

## **5. Tumor stroma: A permissive substrate for breast cancer development and progression**

The stroma of carcinomas is an intricate ecosystem where heterogeneous cell populations coexist. This structural and functional connective tissue niche is inhabited by immune and inflammatory cells such as macrophages and monocytes, mesenchymal bone marrow-derived stem cells, endothelial and pericyte cells, lipocytes, additional smooth muscle cells and activated fibroblastic cells known as myofibroblasts, which are believed to be responsible for producing and maintaining the altered extracellular matrix (ECM) (Beacham & Cukierman, 2005; Li et al., 2007; Xouri & Christian, 2010). It is well accepted that the altered and excessive deposition of ECM, which is part of a process named desmoplasia, is directly associated with rapid progression and bad prognosis in carcinomas such as breast, pancreas, colon and prostate to name a few (Beacham & Cukierman, 2005; Arendt et al., 2010; Franco et al., 2010). In fact, we and others have suggested that stroma progression could be staged (analogously to classic tumor staging) into discrete stromagenic stages (Bissell et al., 2002; Mueller & Fusenig, 2002; Beacham & Cukierman, 2005; Quiros et al., 2008; Castello-Cros et al., 2009). Briefly, under normal (i.e., homeostatic) conditions, the breast stroma maintains the tissue architecture where a specialized ECM rich in collagen IV and laminin-1 known as basement membrane (BM) demarks a barrier between epithelium and the mesenchyme (Gudjonsson et al., 2002). A particular feature of the glandular epithelium in breast tissue is that both alveolar and ductal epithelial cells are not in direct contact with the BM. Instead, they are supported by a monolayer of myoepithelial cells that resides in between. Myoepithelial cells play an important role in supporting epithelial cell differentiation and controlling proliferation and cell polarity. These cells secrete the BM proteins and together with adjacent stromal fibroblasts maintain the integrity of this

specialized gland (Gudjonsson et al., 2002; Polyak & Kalluri, 2010). Under physiological conditions, a normal stroma preserves and drives regular breast tissue morphogenesis (Kuperwasser et al., 2004) and, at the same time, suppresses the transformation of epithelial cells thus preventing the development of breast carcinoma *in situ* (CIS) and inhibiting progression towards invasive cancer (Hu et al., 2008). Although not much information is available to describe the mechanistic events responsible for normal stroma prevention of carcinoma progression, recent data suggests that the tumor microenvironment lacks the regulatory mechanisms that are necessary to maintain a normal epithelial phenotype (Postovit et al., 2008). As shown by interesting work conducted by Mintz and Illmensee in 1975 where they observed that a normal embryo microenvironment is repressive of teratoma tumorigenesis (Mintz & Illmensee, 1975), more recent work by Postovit *et al* looking at specific human embryonic stem cells-secreted factors also concluded that embryonic microenvironments can control and sustain a normal behaviour of invasive tumor cells (Postovit et al., 2008). In summary, one could state that the normal stroma is a natural barrier or a non-permissive environment for tumor progression.

In an effort to understand premature events that occur during stroma progression (i.e., stromagenesis (Cukierman, 2009)), researchers have used animal models where they have shown stromal cells alterations at early stages of tumorigenesis. For example, prostate smooth muscle cells, known to support homeostasis and epithelium differentiation and considered to be analogous to normal myoepithelial cells in breast, have been shown to undergo alterations during early tumorigenesis (Wong & Tam, 2002). Similar to myoepithelial cells, smooth muscle cells are also lost in advanced stages of tumor progression, but prior to this they lower the expression levels of differentiation markers such as myosin, desmin, and laminin (Wong & Tam, 2002). This fact strongly suggests the advent of a discrete intermediate state between normal and activated stroma. To this end, the up-regulated expression of proteins, such as fibroblast activation protein, has been suggested as potential markers of this intermediate or primed stromal stage (Mathew et al., 1995; Huber et al., 2003; Santos et al., 2009). Another such molecule is tenascin-C, an ECM protein expressed in breast cancer at early stages of the tumorigenesis, which has been shown to have a diagnostic value (Adams et al., 2002; Guttery et al., 2010).

Once the stroma becomes activated, many histological features are evident. This stage is commonly described by pathologists as desmoplasia and is characterized by increased interstitial ECM-deposition. The desmoplastic ECM is believed to be produced by a highly proliferating fibroblastic and alpha-smooth muscle actin ( $\alpha$ -SMA) expressing myofibroblastic cell population. It is common in many cancers including breast, and it can constitute up to 50% of the tumor mass (Kunz-Schughart & Knuechel, 2002a, b; Desmouliere et al., 2004). The altered architecture of the desmoplastic stroma reaction is characterized by the over expression of ECM proteins such as collagen I and differential spliced fibronectin isoforms such as EDA and EDB (Matsumoto et al., 1999; Desmouliere et al., 2004). The desmoplastic ECM is highly organized in a parallel fiber pattern, which is clearly oriented *in vivo* perpendicular to the tumor border (Provenzano et al., 2006). In fact, this particular feature of the tumor associated-ECM (TA-ECM) has been suggested to facilitate migration of breast cancer cells *in vitro*, in a  $\beta$ 1-integrin dependent manner (Castello-Cros et al., 2009). Moreover, there is evidence to suggest that TA-ECM can induce a phenotypic switch upon naïve fibroblasts thus inducing a myofibroblastic (or activated) conformation (Amatangelo et al., 2005). To this end, in a xenograft model of human breast cancer, it was shown that

activated fibroblasts influence the local microenvironment to promote invasion (Orimo et al., 2005; Hu et al., 2008).

## **6. Tumor- or carcinoma-associated fibroblasts: A bad myofibroblastic influence**

Fibroblasts are the main cellular component of the stroma and responsible for producing the mesenchymal (i.e., interstitial) ECM. These cells have been described as non-epithelial, non-inflammatory and non-vascular semi-differentiated connective tissue cells (Tarin & Croft, 1969). They are best known for their role in maintaining the tissue's integrity while they become quickly activated (e.g., myofibroblastic) and can modify the plasticity of the resident's tissue under conditions that alter the homeostatic equilibrium such as during wound healing, organogenesis, cancer and other pathological and inflammatory conditions (Kalluri & Zeisberg, 2006). In fact, fibroblasts are known as tissue remodelers capable of renovating ECMs while, at the same time, facilitating access to ECM stored growth factors, such as transforming growth factor-beta (TGF- $\beta$ ), through a tightly regulated release and activation of matrix digestive enzymes such as matrix metalloproteinases (MMPs) (Jodele et al., 2006).

The fibroblastic cell population, known as carcinoma-associated fibroblasts (CAFs) or tumor-associated fibroblasts (Barsky et al., 1984), presents a myofibroblastic phenotype that is very similar to the one observed in activated fibroblasts during wound healing (Barsky et al., 1984). CAFs are the main stromal cell component of solid epithelial carcinomas (Shao et al., 2000). In addition to a characteristic, high proliferation rate and increased ECM deposition, the development of contractile cell features affects the physico-chemical characteristics of TA-ECM (Tomasek et al., 2002; Butcher et al., 2009; Cukierman & Bassi, 2010). Interestingly, CAFs are capable of establishing interactions with inflammatory, endothelial, and tumor cells by means of cytokines/chemokines secretions such as interleukin (IL)- $1\beta$ , IL-6, CXCL-8, stromal derived factor-1 (SDF-1), also known as CXCL-12, and the monocyte chemotactic protein (MCPs/CCLs) among others (Silzle et al., 2004; Mishra et al., 2011). In an effort to find a discrete set of CAF specific markers, proteins such as  $\gamma$ - and  $\alpha$ -SMA (Brouty-Boye et al., 1991; Kunz-Schughart & Knuechel, 2002b; Desmouliere et al., 2004; Xouri & Christian, 2010) specific isoforms of the actin binding protein palladin, (Ronty et al., 2006; Goicoechea et al., 2010; Gupta et al., 2011) as well as the intermediate filament proteins vimentin and desmin (Schmid et al., 1982) have been suggested. Furthermore, the specific breast cancer microenvironmental niche has been shown to contain increased levels of expression of ECM stabilizing (e.g., cross-linking) enzymes such as prolyl-4 hydroxylase (Orimo et al., 2005) and lysyl oxidase (Chang et al., 2005; Levental et al., 2009; Barry-Hamilton et al., 2010). Additional proteins have been shown to be specifically overexpressed at the tumor-associated stroma such as fibroblast activation protein (LeBeau et al., 2009; Lee, 2011), endosialin (Becker et al., 2008; Christian et al., 2008) S100A4 (Ambartsumian et al., 1996; Ryan et al., 2003; Katoh et al., 2010), and a plethora of MMPs, among others (Rasanen & Vaheri, 2010). In fact, some of these have already been proposed to serve as stromal monitoring or prognostic markers (Erkan et al., 2008; Gupta et al., 2011).

Nevertheless, this hardly consistent signature of myofibroblastic markers strongly suggests that the tumor stroma is a heterogeneous milieu (Sugimoto et al., 2006). The variety of

myofibroblastic phenotypes is also suggestive of the eliciting of different roles played by these cell populations at the tumor stroma. Interestingly, this heterogeneity could have been originated (i.e., differentiated) by the multiple cell lineages known to produce myofibroblastic CAFs. These are: local fibroblasts (Kalluri & Zeisberg, 2006), bone marrow recruited mesenchymal cells (Ishii et al., 2003; Goldstein et al., 2010), as well as endothelial and tumor (i.e., epithelial) cells (Petersen et al., 2003; Kalluri & Zeisberg, 2006; Zeisberg et al., 2007), among others. In all these cases, TGF- $\beta$  has been closely associated with tumor-induced myofibroblastic activation or differentiation (Zeisberg et al., 2007; Hinz, 2010; Taylor et al., 2010). The myofibroblastic differentiation is a complex and not yet fully understood process that is believed to play a central role during breast tumorigenesis (Cukierman, 2004; McAllister & Weinberg, 2010). Even though a plethora of molecules has been implicated in regulating fibroblastic activation, the specific desmoplastic response in breast cancer is believed to be driven by four main groups of inducers; i) growth factors, ii) TA-ECM, iii) acute inflammation and iv) microenvironmental stress denoted by nutrient and oxygen deprivation as well as low pH.

- i. Specific growth factor presence at the tumor microenvironment may constitute the most studied aspect believed to trigger a myofibroblastic switch of the otherwise quiescent homeostatic fibroblasts. Determined mainly *in vitro* by an increment in proliferation rate, induction of  $\alpha$ -SMA expression, and an up-regulation of ECM components, the growth factors most commonly implicated in this process are TGF- $\alpha$ , TGF- $\beta$ , insulin-like growth factors I and II (TGF-I and TGF-II), the platelet-derived growth factor (PDGF), and the basic fibroblast growth factor (bFGF) (Beacham & Cukierman, 2005; Kalluri & Zeisberg, 2006; Rasanen & Vaheri, 2010; Xouri & Christian, 2010). Although many questions remain regarding specific triggers for breast cancer desmoplasia, work from Walker and Dearing implicated TGF- $\beta_1$ , TGF- $\beta_2$  and TGF- $\beta$  receptor as vital contributors of breast tumorigenesis associated with a stromal increment of fibronectin and tenascin in the tumor stroma (Walker & Dearing, 1992; Walker et al., 1994). Moreover, TGF- $\beta$  known to induce myofibroblastic differentiation and to increase collagen I deposition during the wound healing process (Desmouliere et al., 2005), has also been implicated as a main factor in inducing breast cancer associated bone marrow-derived myofibroblasts differentiation (Goldstein et al., 2010). Similarly, PDGF has been shown to increase the breast myofibroblastic population by 30% while greatly increasing the amount of interstitial collagen I *in vivo* (Shao et al., 2000). In the context of epithelial to mesenchymal transition (EMT)-derived myofibroblasts, hepatocyte growth factor (HGF) and epidermal growth factor (EGF), in addition to the above-mentioned PDGF and TGF- $\beta$ , have also been implicated (Mimeault & Batra, 2007; Kalluri & Weinberg, 2009).
- ii. Breast TA-ECMs' features are known to become altered in both their molecular composition (Chen, S.T. et al., 2008; Levental et al., 2009; Ronnov-Jessen & Bissell, 2009) and their architectural characteristics (Provenzano et al., 2006). Together these two altered features can modulate tumorigenic behaviours of cancer cells and promote or delay the evolution of carcinomas in a permissive or restrictive manner (Ronnov-Jessen & Bissell, 2009; Cukierman & Bassi, 2010). In addition, it has been suggested that the physico-chemical characteristics of the ECM also affect the behaviour of mesenchymal cells (Discher et al., 2005). Fibroblasts are influenced by stromal stiffness, which exerts mechanical forces that modulate their cell behaviour. Thus, it has been demonstrated

that as the substrate stiffness increases, fibroblastic cells change exhibiting three discrete phenotypic switch stages: normal or naive fibroblasts, intermediate or proto-myofibroblastic and activated myofibroblastic (Hinz, 2010). The phenotype transition induced by the increased tension in the substratum is also accompanied by the maturation or elongation of focal adhesions, together with cytoskeletal changes known to build-up contractile stress fibers (Hinz, 2010). Interestingly, studies of normal breast revealed a relatively limp tissue composition (0.15 kPa, expressed in  $E$  values of a Young modulus) compared to the stiffer and highly desmoplastic ~4 kPa tissue that has been affected by breast cancer (Butcher et al., 2009). The altered (i.e., myofibroblastic) phenotype of fibroblasts is linked to the stiffer ECM during tumor progression as these cells are responsible for the production of the TA-ECM (Cukierman & Bassi, 2010). Indeed increments of mammographic density, suggesting excessive collagen deposition, have been associated with higher risk in breast cancer (Boyd et al., 1998). Moreover, increases in cross-linked collagen due to over expression of LOX together with patterned linearization of the TA-ECM and specific ECM receptor, integrin, clustering and enhanced phosphoinositide 3-kinase (PI3K) activity, have all been correlated with breast cancer progression (Levental et al., 2009). Additionally, it has been shown that the interstitial ECM can function as a reservoir for diffusible molecules, such as the above-mentioned TGF- $\beta$  which is secreted by both stromal and tumor cells in its inactivated form (Wipff & Hinz, 2008), but can be both activated and released due to the intrinsic myofibroblastic forces that increase the tension of TA-ECM's fibrils (Wipff et al., 2007; Tenney & Discher, 2009).

- iii. Recently, an inflammatory microenvironment has been suggested as the seventh hallmark of cancer (Colotta et al., 2009). This cancer hallmark is also believed to play an important role in desmoplasia as a fibroblast phenotypic-switch activator. To this end, it has been demonstrated that stromal inflammatory responses that result from wounding can trigger tumorigenesis (Arwert et al., 2010). The importance of an inflammatory component has also been suggested for the breast cancer stroma (Hu & Polyak, 2008), and its repercussion in inducing or promoting cancer aggressiveness and metastasis has been highlighted in numerous occasions (Pantschenko et al., 2003; Elaraj et al., 2006; Valdivia-Silva et al., 2009; Franco-Barraza et al., 2010; Goldberg & Schwertfeger, 2010). However, our current knowledge regarding fibroblastic responses to inflammatory cytokines in breast cancer remains relatively modest. Work conducted at the Polyak laboratory suggested that cytokines could participate in triggering a fibroblast phenotypic switch at the breast cancer microenvironment (Hu et al., 2009). This work and the work of others has opened up the possibility of targeting inflammatory cytokines for the treatment of neoplasias as in the case of COX-2 and arachidonic acid inhibitors (Chen, X. et al., 2006; Hu et al., 2009). In fact, in the kidney, it has been shown that collagen I regulates COX-2 expression in a pro-proliferative type of response (Alique et al., 2011). Interestingly, CAFs are known to promote inflammation in an NF $\kappa$ -b dependent manner, suggesting a vicious cycle between inflammation and stromal activation during tumorigenesis (Erez et al., 2010). Moreover, it has been shown that CAFs effectively suppress anti-tumor inflammation while, at the same time, maintaining acute inflammatory (pro-tumor) conditions (Kraman et al., 2010). As established before, the cytokine/growth factor TGF- $\beta$  imparts a pleiotropic and decisive role in the promotion of the desmoplastic tumor microenvironment thus

supporting tumor progression (Yang et al., 2010). In addition, this same factor plays an additional important stromal role in inducing the expression of NADPH oxidase family protein, Nox4 (Bondi et al., 2010). Nox4 is a potent regulator of reactive oxygen species (ROS) (Barnes & Gorin, 2011) and has been shown to induce the accumulation of ROS in damaged tissues while transactivation of fibroblasts into myofibroblasts (Cucoranu et al., 2005; Rocic & Lucchesi, 2005). In breast cancer, the oxidative stress present at the tumor stroma is also considered to be an inductor for myofibroblastic differentiation, as recently shown in a JunD deficient mouse model, where the absence of this transcription factor allowed the accumulation of Ras-mediated production of ROS with the subsequent conversion of fibroblasts into myofibroblasts and shortening of the tumor free survival rate (Toullec et al., 2010).

- iv. It is well known that as tumors progress increased regions of nutrient deprivation, low pH and low oxygen tension (hypoxia) are evident. Under these hypoxic stress conditions, breast cancer tissues are known to up-regulate the expression of hypoxia-inducible family (HIF) genes such as HIF-1 $\alpha$  (Chen, C.L. et al., 2010). HIF proteins are known to participate in many cellular events such as angiogenesis, through the induction of vascular endothelium growth factor (VEGF), angiopoietin-2, PDGF and FGF (Allen & Louise Jones, 2011) which in turn can also activate stromal myofibroblastic differentiation in breast cancers (Shao et al., 2000). Finally, other molecules known to be induced by HIF-1 $\alpha$  in carcinomas (and other fibrotic conditions) are the above mentioned ECM-cross-linkers (i.e., LOX) which have been associated with aggressive breast tumorigenesis (Chang et al., 2005; Levental et al., 2009; Barry-Hamilton et al., 2010).

## 7. Fibroblasts as moderators of signals at the tumor microenvironment

At the tumor microenvironment, intercellular communications resemble a social network emitting signals (either static or diffusible molecules) that in turn are collected, processed and emitted to additional cells. Using this analogy, it seems that CAFs play a decisive role during cancer progression acting as microenvironment signals moderators that sense extracellular signals and, after intracellular processing, emit new ones that in turn modulate both stromal and neoplastic neighbouring cells' behaviours (Bhowmick et al., 2004). In fact during cancer progression, CAFs constitute a very important source of the exogenous stimulants such as the above-mentioned TGF- $\beta$  (Kalluri & Zeisberg, 2006). To this end, using an elegant humanized stromal reconstruction model of human breast cancer in mouse, Kuperwasser *et al* demonstrated that CAFs facilitate tumor development in a fibroblastic TGF- $\beta$ - and HGF-dependent manner (Kuperwasser et al., 2004). Additionally, recent findings have demonstrated that epigenetic changes induced by mesenchymal cells on breast cancer cells that are regulated by the TGF- $\beta$ /TGF- $\beta$ R/Smad2 signalling axis provoke the silencing of critical epithelial genes resulting in the pro-tumorigenic EMT process (Papageorgis et al., 2010). To this end, in support of the above proposed vicious cycle effect, it is interesting to note that following quiescent fibroblasts transdifferentiation into CAFs, these cells support an invasive phenotype of mammary carcinomas where they secrete inflammatory cytokines (Powell et al., 1999; Buckley et al., 2001; Silzle et al., 2004) thus activating NF- $\kappa$ b and promoting EMT as well as promoting aggressiveness of breast cancer cells (Sullivan et al., 2009; Wu et al., 2009). An ever more complicated interplay between CAFs, cytokines and neoplastic cells has recently been proposed in breast cancers where,



due to the presence of an altered TA-ECM, an integrin-dependent activation of Src family kinases results in the increase of NF- $\kappa$ B activity which blocks the production of certain microRNAs such as Let-7. Under these conditions, IL-6 production is promoted resulting in the increased secretion of this pro-tumorigenic cytokine, which in turn induces or promotes a positive feedback in tumor cells (Iliopoulos et al., 2009). Moreover, activated myofibroblastic and cancer cells are known to remodel the stromal ECM by means of increased secretion of MMPs and urokinase-type plasminogen activator (uPA). These enzymes cleave the ECM molecules to release fragments that contain chemotactic properties called matrikines that activate leukocytes to also release inflammatory cytokines (Maquart et al., 2004; Silzle et al., 2004). For example, a special feature of MMP-2, -3 and -9 is that these proteases can increase the availability of IL-1b at the tumor microenvironment by cleavage of the pIL-1b (immature IL-1b) (Schonbeck et al., 1998). Also, analyses of co-cultures containing both breast cancer cells and CAFs have shown increases in stromal MMP-2 and MMP-9 expression (Singer et al., 2002). These observations concur with observations stemming from an immunohistochemical study where tissue arrays of breast cancer patients showed that intratumor stromal fibroblasts express MMP-2, -7, and -14, while fibroblast at the invasive front highly express MMP-9. What is more, this specific profile of stromal MMPs staining was found to be a predictor of future distant metastases occurrences (Del Casar et al., 2009). Another uncovered effect of released MMPs into the tumor stroma is the capacity of these molecules to promote a permissive environment that supports epithelial tumorigenic progression including the promotion of genomic alterations (Radisky, E.S. & Radisky, 2007). In the mammary glands of transgenic mice, the overexpression of MMP-3 has been shown to be sufficient to stimulate myofibroblastic presence, increased fibrosis, epithelial hyperplasia, and development of mammary carcinoma (Thomasset et al., 1998). What is more, mammary epithelial cells exposed to stromal MMP-3 showed activation of a genotoxic metabolic pathway, where the over expression of the spliced variant Rac1b produced DNA-damaging superoxide radicals and induced EMT (Radisky, D.C. et al., 2005). Interestingly, the epithelial genomic alterations induced by stromal MMPs *in vitro*, suggest a possible mechanism to understand the presence of areas with genomic imbalance patterns detected in histologically normal tissues adjacent to the tumor stroma (Ellsworth et al., 2004; Holliday et al., 2009).

## 8. Targeting fibroblasts as an anti-cancer therapy

Various aspects of the tumor microenvironment have been explored as putative therapeutic targets in the fight against cancer (Andre et al., 2010; Cukierman & Khan, 2010; Allen & Louise Jones, 2011). Since a desmoplastic reaction is an ECM component-rich substratum and some of the TA-ECM components are believed to be specific for discrete types of carcinomas, they constitute a promising basis for therapeutics (i.e., inhibitory functional antibodies). For example, in glioblastoma patients an iodine-131 radiolabeled anti-tenascin-C monoclonal antibody has produced encouraging results in phase II trials (Reardon et al., 2006). Similarly, the development of radioactive or bioactive molecules coupled to antibodies against TA-ECM specific EDB, the L-19 antibody, showed encouraging results when tested in various carcinomas (Kaspar et al., 2006). The TA-ECM has been considered as both a target as well as a means to attract anti-tumoral drugs. For example, as albumin binds efficiently to the TA-ECM protein osteonectin (also known as SPARC), known to be upregulated in a plethora of cancer stromas and often associated with bad prognosis (Tai &

Tang, 2008), paclitaxel delivered through nanoparticles conjugated to albumin (nab-paclitaxel) are being tested (Vishnu & Roy, 2010; Robert et al., 2011; Volk et al., 2011). Moreover inhibition of the serine protease activity of the CAF specific fibroblast activation protein has been suggested as a therapeutic target in a plethora of cancers including breast (Mersmann et al., 2001). In fact, antibodies against fibroblast activation protein induced a marked decrease in desmoplastic collagen I expression resulting in an increased (up to 70%) increment in chemotherapeutic drugs uptake (Loeffler et al., 2006). Therefore, it is not surprising that fibroblast activation protein has been suggested as a tumor targeting molecule for the delivery of peptide protoxins (amongst others) thus diminishing non-tumoral side effect toxicities (LeBeau et al., 2009).

Pro-inflammatory molecules have also been used as effective targets. For example TNF- $\alpha$  antagonists have been shown to have good results preventing disease acceleration in a considerable number of breast cancer patients (Madhusudan et al., 2004; Brown et al., 2008). The SDF-1 $\alpha$ /CXCR4 chemokine axis has been proposed as a general target for anticancer strategies (Guleng et al., 2005), and recently a compound derived from marine organisms that blocks CXCR4 has been shown effective as well (He et al., 2008). Antibodies blocking the TGF- $\beta$  signalling pathway have been developed and showed promising synergistic effects when added to known chemotherapeutics and, thus, have been regarded as anti angiogenesis-depending tumor stromal agents in breast cancer (Takahashi et al., 2001). Finally, it was recently shown that eliminating pro-tumorigenic macrophages in pancreas causes desmoplastic shrinkage and subsequent tumor stalling (Beatty et al., 2011). We believe that these types of treatments, together with similar novel ones, could provide increased hope in the common fight against breast cancers.

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## 10. References

- Adams, M.; Jones, J.L.; Walker, R.A.; Pringle, J.H. & Bell, S.C. (2002). Changes in tenascin-C isoform expression in invasive and preinvasive breast disease. *Cancer research*, Vol.62, No.11, pp. 3289-3297, ISSN 0008-5472
- Ali, S. & Lazennec, G. (2007). Chemokines: novel targets for breast cancer metastasis. *Cancer metastasis reviews*, Vol. 26, No. pp. 401-420,
- Alique, M.; Calleros, L.; Luengo, A.; Grieria, M.; Iniguez, M.A.; Punzon, C.; Fresno, M.; Rodriguez-Puyol, M. & Rodriguez-Puyol, D. (2011). Changes in extracellular matrix composition regulate cyclooxygenase-2 expression in human mesangial cells. *Am J Physiol Cell Physiol*, Vol.300, No.4, pp. C907-918, ISSN 1522-1563
- Allavena, P.; Sica, A.; Solinas, G.; Porta, C. & Mantovani, A. (2008). The inflammatory micro-environment in tumor progression: The role of tumor-associated macrophages. *Critical reviews in oncology/hematology*, Vol. 66, No. 1, pp. 1-9, 1040-8428
- Allen, M. & Louise Jones, J. (2011). Jekyll and Hyde: the role of the microenvironment on the progression of cancer. *The Journal of pathology*, Vol.223, No.2, pp. 162-176, ISSN 1096-9896

- Allen, S.J.; Crown, S.E. & Handel, T.M. (2007). Chemokine: receptor structure, interactions, and antagonism. *Annual review of immunology*, Vol. 25, No. pp. 787-820,
- Amatangelo, M.D.; Bassi, D.E.; Klein-Szanto, A.J. & Cukierman, E. (2005). Stroma-derived three-dimensional matrices are necessary and sufficient to promote desmoplastic differentiation of normal fibroblasts. *The American journal of pathology*, Vol.167, No.2, pp. 475-488, ISSN 0002-9440
- Ambartsumian, N.S.; Grigorian, M.S.; Larsen, I.F.; Karlstrom, O.; Sidenius, N.; Rygaard, J.; Georgiev, G. & Lukanidin, E. (1996). Metastasis of mammary carcinomas in GRS/A hybrid mice transgenic for the mts1 gene. *Oncogene*, Vol.13, No.8, pp. 1621-1630, ISSN 0950-9232
- Amersi, F.F.; Terando, A.M.; Goto, Y.; Scolyer, R.A.; Thompson, J.F.; Tran, A.N.; Faries, M.B.; Morton, D.L. & Hoon, D.S.B. (2008). Activation of CCR9/CCL25 in Cutaneous Melanoma Mediates Preferential Metastasis to the Small Intestine. *Clinical cancer research*, Vol. 14, No. 3, pp. 638-645, ISSN 1557-3265
- Andre, F.; Berrada, N. & Desmedt, C. (2010). Implication of tumor microenvironment in the resistance to chemotherapy in breast cancer patients. *Curr Opin Oncol*, Vol. 22, No. 6, pp. 547-551, 1531-703X (Electronic)
- Angelo, L.S.; Talpaz, M. & Kurzrock, R. (2002). Autocrine Interleukin-6 Production in Renal Cell Carcinoma. *Cancer research*, Vol. 62, No. 3, pp. 932-940,
- Annels, N.E.; Williemze, A.J.; van der Velden, V.H.; Faaij, C.M.; E. van Wering, E.; Sie-Go, D.M.; R.M. Egeler, R.M.; van Tol, M.J. & Revesz, T. (2004). Possible link between unique chemokine and homing receptor expression at diagnosis and relapse location in a patient with childhood T-ALL. *Blood*, Vol. 103, No. pp. 2806-2808,
- Arendt, L.M.; Rudnick, J.A.; Keller, P.J. & Kuperwasser, C. (2010). Stroma in breast development and disease. *Seminars in cell & developmental biology*, Vol.21, No.1, pp. 11-18, ISSN 1096-3634
- Arlt, A.; Vorndamm, J.; Mfi?erkfi?ster, S.; Yu, H.; Schmidt, W.E.; Ffi?lsch, U.R. & Schfi?fer, H. (2002). Autocrine Production of Interleukin 1â? Confers Constitutive Nuclear Factor â?B Activity and Chemoresistance in Pancreatic Carcinoma Cell Lines. *Cancer research*, Vol. 62, No. 3, pp. 910-916,
- Arwert, E.N.; Lal, R.; Quist, S.; Rosewell, I.; van Rooijen, N. & Watt, F.M. (2010). Tumor formation initiated by nondividing epidermal cells via an inflammatory infiltrate. *Proceedings of the National Academy of Sciences of the United States of America*, Vol.107, No.46, pp. 19903-19908, ISSN 1091-6490
- Baggiolini, M.; Dewald, B. & Moser, B. (1997). Human chemokines: an update. *Annual review of immunology*, Vol. 15, No. pp. 675-705,
- Bagley, R.G.; Nannuru, K.C.; Singh, S. & Singh, R.K. (2010). Chemokines and Metastasis. In: *Chemokines and Metastasis* Book. pp. 601-631. Springer New York, 978-1-4419-6615-5
- Balkwill, F. & Mantovani, A. (2001). Inflammation and cancer: back to Virchow? *Lancet*, Vol. 357, No. 9255, pp. 539-545, ISSN 0140-6736
- Barnes, J.L. & Gorin, Y. (2011). Myofibroblast differentiation during fibrosis: role of NAD(P)H oxidases. *Kidney international*, ISSN 1523-1755

- Barry-Hamilton, V.;Spangler, R.;Marshall, D.;McCauley, S.;Rodriguez, H.M.;Oyasu, M.;Mikels, A.;Vaysberg, M.;Ghermazien, H.;Wai, C.;Garcia, C.A.;Velayo, A.C.;Jorgensen, B.;Biermann, D.;Tsai, D.;Green, J.;Zaffryar-Eilot, S.;Holzer, A.;Ogg, S.;Thai, D.;Neufeld, G.;Van Vlasselaer, P. & Smith, V. (2010). Allosteric inhibition of lysyl oxidase-like-2 impedes the development of a pathologic microenvironment. *Nature medicine*, Vol.16, No.9, pp. 1009-1017, ISSN 1546-170X
- Barsky, S.H.;Green, W.R.;Grotendorst, G.R. & Liotta, L.A. (1984). Desmoplastic breast carcinoma as a source of human myofibroblasts. *The American journal of pathology*, Vol.115, No.3, pp. 329-333, ISSN 0002-9440
- Beacham, D.A. & Cukierman, E. (2005). Stromagenesis: the changing face of fibroblastic microenvironments during tumor progression. *Seminars in cancer biology*, Vol.15, No.5, pp. 329-341, ISSN 1044-579X
- Beatty, G.L.;Chiorean, E.G.;Fishman, M.P.;Saboury, B.;Teitelbaum, U.R.;Sun, W.;Huhn, R.D.;Song, W.;Li, D.;Sharp, L.L.;Torigian, D.A.;O'Dwyer, P.J. & Vonderheide, R.H. (2011). CD40 agonists alter tumor stroma and show efficacy against pancreatic carcinoma in mice and humans. *Science*, Vol.331, No.6024, pp. 1612-1616, ISSN 1095-9203
- Becker, R.;Lenter, M.C.;Vollkommer, T.;Boos, A.M.;Pfaff, D.;Augustin, H.G. & Christian, S. (2008). Tumor stroma marker endosialin (Tem1) is a binding partner of metastasis-related protein Mac-2 BP/90K. *FASEB J*, Vol.22, No.8, pp. 3059-3067, ISSN 1530-6860
- Ben-Baruch, A. (2003). Host microenvironment in breast cancer development: inflammatory cells, cytokines and chemokines in breast cancer progression: reciprocal tumor-microenvironment interactions. *Breast cancer research.*, Vol. 5, No. pp. 31-36,
- Ben-Baruch, A. (2006). Inflammation-associated immune suppression in cancer: The roles played by cytokines, chemokines and additional mediators. *Seminars in cancer biology*, Vol. 16, No. 1, pp. 38-52, 1044-579X
- Ben-Baruch, A. (2008). Organ selectivity in metastasis: regulation by chemokines and their receptors. *Clinical and experimental metastasis*, Vol. 25, No. 4, pp. 345-356,
- Bhowmick, N.A.;Neilson, E.G. & Moses, H.L. (2004). Stromal fibroblasts in cancer initiation and progression. *Nature*, Vol.432, No.7015, pp. 332-337, ISSN 1476-4687
- Bissell, M.J.;Radisky, D.C.;Rizki, A.;Weaver, V.M. & Petersen, O.W. (2002). The organizing principle: microenvironmental influences in the normal and malignant breast. *Differentiation*, Vol.70, No.9-10, pp. 537-546
- Bondi, C.D.;Manickam, N.;Lee, D.Y.;Block, K.;Gorin, Y.;Abboud, H.E. & Barnes, J.L. (2010). NAD(P)H oxidase mediates TGF-beta1-induced activation of kidney myofibroblasts. *Journal of the American Society of Nephrology : JASN*, Vol.21, No.1, pp. 93-102, ISSN 1533-3450
- Bos, R.; van der Groep, P.; Greijer, A.E.; Shvarts, A.; Meijer, S.; Pinedo, H.M.; Semenza, G.L.; van Diest, P.J. & van der Wall, E. (2003). Levels of hypoxia-inducible factor-1? independently predict prognosis in patients with lymph node negative breast carcinoma. *Cancer*, Vol. 97, No. 6, pp. 1573-1581, 1097-0142
- Bos, R.; van Diest, P.; van der Groep, P.; Shvarts, A.; Greijer, A. & van der Wall, E. (2004). Expression of hypoxia-inducible factor-1alpha and cell cycle proteins in invasive

- breast cancer are estrogen receptor related. *Breast Cancer Res*, Vol. 6, No. 4, pp. R450 - R459, ISSN 1465-5411
- Bos, R.; Zhong, H.; Hanrahan, C.F.; Mommers, E.C.; Semenza, G.L.; Pinedo, H.M.; Abeloff, M.D.; Simons, J.W.; van Diest, P.J. & van der Wall, E. (2001). Levels of hypoxia-inducible factor-1? during breast carcinogenesis. *Journal of the national cancer institute*, Vol. 93, No. pp. 309-314,
- Boyd, N.F.;Lockwood, G.A.;Martin, L.J.;Knight, J.A.;Byng, J.W.;Yaffe, M.J. & Tritchler, D.L. (1998). Mammographic densities and breast cancer risk. *Breast disease*, Vol.10, No.3-4, pp. 113-126, ISSN 0888-6008
- Braun, S.E.; Chen, K.; Foster, R.G.; Kim, C.H.; Hromas, R.; Kaplan, M.H.; Broxmeyer, H.E. & Cornetta, K. (2000). The CC Chemokine CKfi1-11/MIP-3fi1/ELC/Exodus 3 Mediates Tumor Rejection of Murine Breast Cancer Cells Through NK Cells. *The Journal of Immunology*, Vol. 164, No. 8, pp. 4025-4031,
- Brouty-Boye, D.;Raux, H.;Azzarone, B.;Tamboise, A.;Tamboise, E.;Beranger, S.;Magnien, V.;Pihan, I.;Zardi, L. & Israel, L. (1991). Fetal myofibroblast-like cells isolated from post-radiation fibrosis in human breast cancer. *Int J Cancer*, Vol.47, No.5, pp. 697-702, ISSN 0020-7136
- Brown, E.R.;Charles, K.A.;Hoare, S.A.;Rye, R.L.;Jodrell, D.I.;Aird, R.E.;Vora, R.;Prabhakar, U.;Nakada, M.;Corringham, R.E.;DeWitte, M.;Sturgeon, C.;Propper, D.;Balkwill, F.R. & Smyth, J.F. (2008). A clinical study assessing the tolerability and biological effects of infliximab, a TNF-alpha inhibitor, in patients with advanced cancer. *Annals of oncology : official journal of the European Society for Medical Oncology / ESMO*, Vol.19, No.7, pp. 1340-1346, ISSN 1569-8041
- Buckley, C.D.;Pilling, D.;Lord, J.M.;Akbar, A.N.;Scheel-Toellner, D. & Salmon, M. (2001). Fibroblasts regulate the switch from acute resolving to chronic persistent inflammation. *Trends in immunology*, Vol.22, No.4, pp. 199-204, ISSN 1471-4906
- Butcher, D.T.;Alliston, T. & Weaver, V.M. (2009). A tense situation: forcing tumor progression. *Nature reviews. Cancer*, Vol.9, No.2, pp. 108-122, ISSN 1474-1768
- Castello-Cros, R.;Khan, D.R.;Simons, J.;Valianou, M. & Cukierman, E. (2009). Staged stromal extracellular 3D matrices differentially regulate breast cancer cell responses through PI3K and beta1-integrins. *BMC Cancer*, Vol.9, pp. 94, ISSN 1471-2407
- Chambers, A.F.; Groom, A.C. & MacDonald, I.C. (2002). Dissemination and growth of cancer cells in metastatic sites. *Nature review in cancer*, Vol. 2, No. 8, pp. 563-572, ISSN 1474-175X
- Chandrasekar, B.; Mummidi, S.; Perla, R.P.; Bysani, S.; Dulin, N.O.; Liu, F. & Melby, P.C. (2003). Fractalkine (CX3CL1) stimulated by nuclear factor kappaB (NF-kappaB)-dependent inflammatory signals induces aortic smooth muscle cell proliferation through an autocrine pathway. *The biochemical journal*, Vol. 373, No. 2, pp. 547-558,
- Chang, H.Y.;Nuyten, D.S.;Sneddon, J.B.;Hastie, T.;Tibshirani, R.;Sorlie, T.;Dai, H.;He, Y.D.;van't Veer, L.J.;Bartelink, H.;van de Rijn, M.;Brown, P.O. & van de Vijver, M.J. (2005). Robustness, scalability, and integration of a wound-response gene expression signature in predicting breast cancer survival. *Proceedings of the*

- National Academy of Sciences of the United States of America, Vol.102, No.10, pp. 3738-3743, ISSN 0027-8424
- Chang, S.-H.; Ai, Y.; Breyer, R.M.; Lane, T.F. & Hla, T. (2005). The Prostaglandin E2 Receptor EP2 Is Required for Cyclooxygenase 2<sub>1</sub>-Mediated Mammary Hyperplasia. *Cancer research*, Vol. 65, No. 11, pp. 4496-4499,
- Charalabopoulos, K.; Dalavaga, Y.; Stefanou, D.; Charalabopoulos, A.; Bablekos, G. & Constantopoulos, S. (2004). Direct endobronchial metastasis is a rare metastatic pattern in breast cancer. *International Journal of Clinical Practice*, Vol. 58, No. 6, pp. 641-644, ISSN 1742-1241
- Chavey, C.; Bibeau, F.; Gourgou-Bourgade, S.; Burlincho, S.; Boissiere, F.; Laune, D.; Roques, S. & Lazenec, G. (2007). Oestrogen receptor negative breast cancers exhibit high cytokine content. *Breast Cancer Research*, Vol. 9, No. 1, pp. R15, 1465-5411
- Chen, C.L.; Chu, J.S.; Su, W.C.; Huang, S.C. & Lee, W.Y. (2010). Hypoxia and metabolic phenotypes during breast carcinogenesis: expression of HIF-1alpha, GLUT1, and CAIX. *Virchows Archiv : an international journal of pathology*, Vol.457, No.1, pp. 53-61, ISSN 1432-2307
- Chen, G.S.; Yu, H.S.; Lan, C.C.E.; Chow, K.C.; Lin, T.Y.; Kok, L.F.; Lu, M.P.; Liu, C.H. & Wu, M.T. (2006). CXC chemokine receptor CXCR4 expression enhances tumorigenesis and angiogenesis of basal cell carcinoma. *British Journal of Dermatology*, Vol. 154, No. 5, pp. 910-918, ISSN 1365-2133
- Chen, S.T.; Pan, T.L.; Juan, H.F.; Chen, T.Y.; Lin, Y.S. & Huang, C.M. (2008). Breast tumor microenvironment: proteomics highlights the treatments targeting secretome. *J Proteome Res*, Vol.7, No.4, pp. 1379-1387, ISSN 1535-3893
- Chen, X.; Sood, S.; Yang, C.S.; Li, N. & Sun, Z. (2006). Five-lipoxygenase pathway of arachidonic acid metabolism in carcinogenesis and cancer chemoprevention. *Curr Cancer Drug Targets*, Vol.6, No.7, pp. 613-622, ISSN 1873-5576
- Christian, S.; Winkler, R.; Helfrich, I.; Boos, A.M.; Besemfelder, E.; Schadendorf, D. & Augustin, H.G. (2008). Endosialin (Tem1) is a marker of tumor-associated myofibroblasts and tumor vessel-associated mural cells. *The American journal of pathology*, Vol.172, No.2, pp. 486-494, ISSN 0002-9440
- Colotta, F.; Allavena, P.; Sica, A.; Garlanda, C. & Mantovani, A. (2009). Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis*, Vol. 30, No. 7, pp. 1073-1081, 1460-2180 (Electronic)
- Colotta, F.; Allavena, P.; Sica, A.; Garlanda, C. & Mantovani, A. (2009). Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis*, Vol.30, No.7, pp. 1073-1081, ISSN 1460-2180
- Coussens, L.M. & Werb, Z. (2002). Inflammation and cancer. *Nature*, Vol. 420, No. pp. 860-867,
- Croitoru-Lamoury, J.; Guillemin, G.J.; Boussin, F.D.; Mognetti, B.; Gigout, L.I.; Cheret, A.; Vaslin, B.; Le Grand, R.; Brew, B.J. & Dormont, D. (2003). Expression of chemokines and their receptors in human and simian astrocytes: evidence for a central role of TNF alpha and IFN gamma in CXCR4 and CCR5 modulation. *Glia*, Vol. 41, No. pp. 354-370,

- Crowther, M.; Brown, N.J.; Bishop, E.T. & Lewis, C.E. (2001). Microenvironmental influence on macrophage regulation of angiogenesis in wounds and malignant tumors. *Journal of Leukocyte Biology*, Vol. 70, No. 4, pp. 478-490,
- Cucoranu, I.; Clempus, R.; Dikalova, A.; Phelan, P.J.; Ariyan, S.; Dikalov, S. & Sorescu, D. (2005). NAD(P)H oxidase 4 mediates transforming growth factor-beta1-induced differentiation of cardiac fibroblasts into myofibroblasts. *Circulation research*, Vol.97, No.9, pp. 900-907, ISSN 1524-4571
- Cukierman, E. (2004). A visual-quantitative analysis of fibroblastic stromagenesis in breast cancer progression. *Journal of mammary gland biology and neoplasia*, Vol.9, No.4, pp. 311-324, ISSN 1083-3021
- Cukierman, E. (2009). Stromagenesis. *Encyclopedia of Cancer*. M. Schwab. Heidelberg/Germany, Springer. 4: 2843-2845.
- Cukierman, E. & Bassi, D.E. (2010). Physico-mechanical aspects of extracellular matrix influences on tumorigenic behaviors. *Seminars in cancer biology*, Vol.20, No.3, pp. 139-145, ISSN 1096-3650 (
- Cukierman, E. & Khan, D.R. (2010). The benefits and challenges associated with the use of drug delivery systems in cancer therapy. *Biochem Pharmacol*, Vol.80, No.5, pp. 762-770, ISSN 1873-2968
- Del Casar, J.M.; Gonzalez, L.O.; Alvarez, E.; Junquera, S.; Marin, L.; Gonzalez, L.; Bongera, M.; Vazquez, J. & Vizoso, F.J. (2009). Comparative analysis and clinical value of the expression of metalloproteases and their inhibitors by intratumor stromal fibroblasts and those at the invasive front of breast carcinomas. *Breast cancer research and treatment*, Vol.116, No.1, pp. 39-52, ISSN 1573-7217
- Desmouliere, A.; Chaponnier, C. & Gabbiani, G. (2005). Tissue repair, contraction, and the myofibroblast. *Wound Repair Regen*, Vol.13, No.1, pp. 7-12, ISSN 1067-1927
- Desmouliere, A.; Guyot, C. & Gabbiani, G. (2004). The stroma reaction myofibroblast: a key player in the control of tumor cell behavior. *Int J Dev Biol*, Vol.48, No.5-6, pp. 509-517, ISSN 0214-6282
- Discher, D.E.; Janmey, P. & Wang, Y.L. (2005). Tissue cells feel and respond to the stiffness of their substrate. *Science*, Vol.310, No.5751, pp. 1139-1143, ISSN 1095-9203
- Egeblad, M.; Nakasone, E.S. & Werb, Z. (2010). Tumors as organs: complex tissues that interface with the entire organism. *Dev Cell*, Vol.18, No.6, pp. 884-901, ISSN 1878-1551
- Elaraj, D.M.; Weinreich, D.M.; Varghese, S.; Puhlmann, M.; Hewitt, S.M.; Carroll, N.M.; Feldman, E.D.; Turner, E.M. & Alexander, H.R. (2006). The role of interleukin 1 in growth and metastasis of human cancer xenografts. *Clinical cancer research : an official journal of the American Association for Cancer Research*, Vol.12, No.4, pp. 1088-1096, ISSN 1078-0432
- Ellsworth, D.L.; Ellsworth, R.E.; Love, B.; Deyarmin, B.; Lubert, S.M.; Mittal, V. & Shriver, C.D. (2004). Genomic patterns of allelic imbalance in disease free tissue adjacent to primary breast carcinomas. *Breast cancer research and treatment*, Vol.88, No.2, pp. 131-139, ISSN 0167-6806
- Erez, N.; Truitt, M.; Olson, P.; Arron, S.T. & Hanahan, D. (2010). Cancer-Associated Fibroblasts Are Activated in Incipient Neoplasia to Orchestrate Tumor-Promoting

- Inflammation in an NF-kappaB-Dependent Manner. *Cancer cell*, Vol.17, No.2, pp. 135-147, ISSN 1878-3686
- Erkan, M.;Michalski, C.W.;Rieder, S.;Reiser-Erkan, C.;Abiatari, I.;Kolb, A.;Giese, N.A.;Esposito, I.;Friess, H. & Kleeff, J. (2008). The Activated Stroma Index Is a Novel and Independent Prognostic Marker in Pancreatic Ductal Adenocarcinoma. *Clin Gastroenterol Hepatol*, Vol.6, No.10, pp. 1155-1161, ISSN 1542-7714 (Electronic)
- Fidler, I.J. (2002). Critical determinants of metastasis. *Seminars in cancer biology*, Vol. 12, No. 2, pp. 89-96, ISSN 1044-579X
- Flanagan, K.; Glover, R.T.; H\_rig, H.; Yang, W. & Kaufman, H.L. (2004). Local delivery of recombinant vaccinia virus expressing secondary lymphoid chemokine (SLC) results in a CD4 T-cell dependent antitumor response. *Vaccine*, Vol. 22, No. 21-22, pp. 2894-2903, 0264-410X
- Franco-Barraza, J.; Valdivia-Silva, J.E.; Zamudio-Meza, H.; Castillo, A.; Garcia-Zepeda, E.A.; Benitez-Bribiesca, L. & Meza, I. (2010). Actin cytoskeleton participation in the onset of IL-1beta induction of an invasive mesenchymal-like phenotype in epithelial MCF-7 cells. *Archives of medical research*, Vol. 41, No. 3, pp. 170-181, 1873-5487 (Electronic)
- Franco, O.E.;Shaw, A.K.;Strand, D.W. & Hayward, S.W. (2010). Cancer associated fibroblasts in cancer pathogenesis. *Seminars in cell & developmental biology*, Vol.21, No.1, pp. 33-39, ISSN 1096-3634
- Freund, A.; Chauveau, C.; Brouillet, J.; Lucas, A.; Lacroix, M.; Licznar, A.; Vignon, F. & Lazennec, G. (2003). IL-8 expression and its possible relationship with estrogen-receptor-negative status of breast cancer cells. *Oncogene*, Vol. 22, No. pp. 256 - 265,
- Freund, A.; Jolivel, V.; Durand, S.; Kersual, N.; Chalbos, D.; Chavey, C.; Vignon, F. & Lazennec, G. (2004). Mechanisms underlying differential expression of interleukin-8 in breast cancer cells. *Oncogene*, Vol. 23, No. pp. 6105 - 6114,
- Fujimoto, K.; Imaizumi, T.; Yoshida, H.; Takanashi, S.; Okumura, K. & Satoh, K. (2001). Interferon-gamma Stimulates Fractalkine Expression in Human Bronchial Epithelial Cells and Regulates Mononuclear Cell Adherence. *Am. J. Respir. Cell Mol. Biol.*, Vol. 25, No. 2, pp. 233-238,
- Garg, A.K. & Aggarwal, B.B. (2002). Reactive oxygen intermediates in TNF signalling. *Molecular Immunology*, Vol. 39, No. 9, pp. 509-517, 0161-5890
- Goicoechea, S.M.;Bednarski, B.;Stack, C.;Cowan, D.W.;Volmar, K.;Thorne, L.;Cukierman, E.;Rustgi, A.K.;Brentnall, T.;Hwang, R.F.;McCulloch, C.A.;Yeh, J.J.;Bentrem, D.J.;Hochwald, S.N.;Hingorani, S.R.;Kim, H.J. & Otey, C.A. (2010). Isoform-specific upregulation of palladin in human and murine pancreas tumors. *PloS one*, Vol.5, No.4, pp. e10347, ISSN 1932-6203
- Goldberg, J.E. & Schwertfeger, K.L. (2010). Proinflammatory cytokines in breast cancer: mechanisms of action and potential targets for therapeutics. *Current drug targets*, Vol.11, No.9, pp. 1133-1146, ISSN 1873-5592
- Goldstein, R.H.;Reagan, M.R.;Anderson, K.;Kaplan, D.L. & Rosenblatt, M. (2010). Human bone marrow-derived MSCs can home to orthotopic breast cancer tumors and



- promote bone metastasis. *Cancer research*, Vol.70, No.24, pp. 10044-10050, ISSN 1538-7445
- Gross, N. & Meier, R. (2009). Chemokines in neuroectodermal cancers: The crucial growth signal from the soil. *Seminars in cancer biology*, Vol. 19, No. 2, pp. 103-110, ISSN 1044-579X
- Gudjonsson, T.; Ronnov-Jessen, L.; Villadsen, R.; Rank, F.; Bissell, M.J. & Petersen, O.W. (2002). Normal and tumor-derived myoepithelial cells differ in their ability to interact with luminal breast epithelial cells for polarity and basement membrane deposition. *Journal of cell science*, Vol.115, No.Pt 1, pp. 39-50, ISSN 0021-9533
- Guleng, B.; Tateishi, K.; Ohta, M.; Kanai, F.; Jazag, A.; Ijichi, H.; Tanaka, Y.; Washida, M.; Morikane, K.; Fukushima, Y.; Yamori, T.; Tsuruo, T.; Kawabe, T.; Miyagishi, M.; Taira, K.; Sata, M. & Omata, M. (2005). Blockade of the stromal cell-derived factor-1/CXCR4 axis attenuates in vivo tumor growth by inhibiting angiogenesis in a vascular endothelial growth factor-independent manner. *Cancer research*, Vol.65, No.13, pp. 5864-5871, ISSN 0008-5472
- Gupta, V.; Bassi, D.E.; Simons, J.D.; Al-Saleem, T.I.; Devarajan, K.; Uzzo, R.G. & Cukierman, E. (2011). Elevated expression of stromal palladin predicts poor clinical outcome in renal cell carcinoma. *PloS one*, Vol. 6, No.6, pp. e21494, ISSN
- Guttery, D.S.; Hancox, R.A.; Mulligan, K.T.; Hughes, S.; Lambe, S.M.; Pringle, J.H.; Walker, R.A.; Jones, J.L. & Shaw, J.A. (2010). Association of invasion-promoting tenascin-C additional domains with breast cancers in young women. *Breast Cancer Res*, Vol.12, No.4, pp. R57, ISSN 1465-542X
- Hadden, J.W. (1999). The immunology and immunotherapy of breast cancer: an update. *International journal of immunopharmacology*, Vol. 21, No. pp. 79-101,
- Half, E.; Tang, X.M.; Gwyn, K.; Sahin, A.; Wathen, K. & Sinicrope, F.A. (2002). Cyclooxygenase-2 Expression in Human Breast Cancers and Adjacent Ductal Carcinoma in Situ. *Cancer research*, Vol. 62, No. 6, pp. 1676-1681,
- He, X.; Fang, L.; Wang, J.; Yi, Y.; Zhang, S. & Xie, X. (2008). Bryostatins blocks stromal cell-derived factor-1 induced chemotaxis via desensitization and down-regulation of cell surface CXCR4 receptors. *Cancer research*, Vol.68, No.21, pp. 8678-8686, ISSN 1538-7445
- Hinz, B. (2010). The myofibroblast: paradigm for a mechanically active cell. *Journal of biomechanics*, Vol.43, No.1, pp. 146-155, ISSN 1873-2380
- Holliday, C.; Rummel, S.; Hooke, J.A.; Shriver, C.D.; Ellsworth, D.L. & Ellsworth, R.E. (2009). Genomic instability in the breast microenvironment? A critical evaluation of the evidence. *Expert review of molecular diagnostics*, Vol.9, No.7, pp. 667-678, ISSN 1744-8352
- Hsiao, Y.H.; Chou, M.C.; Fowler, C.; Mason, J.T. & Man, Y. (2010). Breast cancer heterogeneity: mechanisms, proofs, and implications. *Journal of cancer*, Vol. 1, No. pp. 6-13,
- Hu, M. & Polyak, K. (2008). Molecular characterisation of the tumor microenvironment in breast cancer. *European journal of cancer*, Vol.44, No.18, pp. 2760-2765, ISSN 1879-0852

- Hu, M.;Peluffo, G.;Chen, H.;Gelman, R.;Schnitt, S. & Polyak, K. (2009). Role of COX-2 in epithelial-stromal cell interactions and progression of ductal carcinoma in situ of the breast. *Proceedings of the National Academy of Sciences of the United States of America*, Vol.106, No.9, pp. 3372-3377, ISSN 1091-6490
- Hu, M.;Yao, J.;Carroll, D.K.;Weremowicz, S.;Chen, H.;Carrasco, D.;Richardson, A.;Violette, S.;Nikolskaya, T.;Nikolsky, Y.;Bauerlein, E.L.;Hahn, W.C.;Gelman, R.S.;Allred, C.;Bissell, M.J.;Schnitt, S. & Polyak, K. (2008). Regulation of in situ to invasive breast carcinoma transition. *Cancer cell*, Vol.13, No.5, pp. 394-406, ISSN 1878-3686
- Huber, M.A.;Kraut, N.;Park, J.E.;Schubert, R.D.;Rettig, W.J.;Peter, R.U. & Garin-Chesa, P. (2003). Fibroblast activation protein: differential expression and serine protease activity in reactive stromal fibroblasts of melanocytic skin tumors. *The Journal of investigative dermatology*, Vol.120, No.2, pp. 182-188, ISSN 0022-202X
- Iliopoulos, D.;Hirsch, H.A. & Struhl, K. (2009). An epigenetic switch involving NF-kappaB, Lin28, Let-7 MicroRNA, and IL6 links inflammation to cell transformation. *Cell*, Vol.139, No.4, pp. 693-706, ISSN 1097-4172
- Ishii, G.;Sangai, T.;Oda, T.;Aoyagi, Y.;Hasebe, T.;Kanomata, N.;Endoh, Y.;Okumura, C.;Okuhara, Y.;Magae, J.;Emura, M.;Ochiya, T. & Ochiai, A. (2003). Bone-marrow-derived myofibroblasts contribute to the cancer-induced stromal reaction. *Biochem Biophys Res Commun*, Vol.309, No.1, pp. 232-240, ISSN 0006-291X
- Jemal, A.; Bray, F.; Center, M.M.; Ferlay, J.; Ward, E. & Forman, D. (2011). Global cancer statistics. *CA: A Cancer Journal for Clinicians*, Vol. 61, No. 2, pp. 69-90, ISSN 1542-4863
- Jiang, B.H.; Rue, E.; Wang, G.; Roe, R. & Semenza, G.L. (1996). Dimerization, DNA binding, and transactivation properties of hypoxia-inducible factor 1. *J Biol Chem*, Vol. 271:, No. pp. 17771-17778,
- Jin, L.; Yuan, R.Q.; Fuchs, A.; Yao, Y.; Joseph, A.; Schwall, R.; Schnitt, S.J.; Guida, A.; Hastings, H.M.; Andres, J.; Turkel, G.; Polverini, P.J.; Goldberg, I.D. & Rosen, E.M. (1997). Expression of interleukin-1? in human breast carcinoma. *Cancer*, Vol. 80, No. 3, pp. 421-434, 1097-0142
- Jodele, S.;Blavier, L.;Yoon, J.M. & DeClerck, Y.A. (2006). Modifying the soil to affect the seed: role of stromal-derived matrix metalloproteinases in cancer progression. *Cancer metastasis reviews*, Vol.25, No.1, pp. 35-43, ISSN 0167-7659
- Johnson-Holiday, C.; Singh, S.; Johnson, E.; Singh, U. & Lillard, J.W. (2007). CCR9- CCL25 interaction mediates breast cancer cell survival via Akt activation. *Journal of immunology*, Vol. 178, No. pp. 49.22,
- Johnson, H.L. (2010). A rare presentation of metastatic breast cancer in a woman with apparent cholangiocarcinoma. *Journal of the american academy of physician assistants*, Vol. 23, No. 3, pp. 32-36,
- Joyce, J.A. & Pollard, J.W. (2009). Microenvironmental regulation of metastasis. *Nature reviews. Cancer*, Vol. 9, No. 4, pp. 239-252, ISSN 1474-175X
- Kalluri, R. & Weinberg, R.A. (2009). The basics of epithelial-mesenchymal transition. *The Journal of clinical investigation*, Vol.119, No.6, pp. 1420-1428, ISSN 1558-8238
- Kalluri, R. & Zeisberg, M. (2006). Fibroblasts in cancer. *Nature reviews. Cancer*, Vol.6, No.5, pp. 392-401, ISSN 1474-175X

- Kanoh, K.; Shimura, T.; Tsutsumi, S.; Suzuki, H.; Kashiwabara, K.; Nakajima, T. & Kuwano, H. (2001). Significance of contracted cholecystitis lesions as high risk for gallbladder carcinogenesis. *Cancer letters*, Vol. 169, No. 1, pp. 7-14, 0304-3835
- Karczewska, A.; Nawrocki, S.; Br?borowicz, D.; Filas, V. & Mackiewicz, A. (2000). Expression of interleukin-6, interleukin-6 receptor, and glycoprotein 130 correlates with good prognoses for patients with breast carcinoma. *Cancer*, Vol. 88, No. 9, pp. 2061-2071, 1097-0142
- Karnoub, A.E. & Weinberg, R.A. (2006). Chemokine networks and breast cancer metastasis. *Breast disease*, Vol. 26, No. pp. 75-85,
- Kaspar, M.;Zardi, L. & Neri, D. (2006). Fibronectin as target for tumor therapy. *International journal of cancer. Journal international du cancer*, Vol.118, No.6, pp. 1331-1339, ISSN 0020-7136
- Katoh, H.;Hosono, K.;Ito, Y.;Suzuki, T.;Ogawa, Y.;Kubo, H.;Kamata, H.;Mishima, T.;Tamaki, H.;Sakagami, H.;Sugimoto, Y.;Narumiya, S.;Watanabe, M. & Majima, M. (2010). COX-2 and prostaglandin EP3/EP4 signaling regulate the tumor stromal proangiogenic microenvironment via CXCL12-CXCR4 chemokine systems. *The American journal of pathology*, Vol.176, No.3, pp. 1469-1483, ISSN 1525-2191
- Keeley, E.C.; Mehrad, B.; Strieter, R.M.; George, F.V.W. & George, K. (2010). CXC Chemokines in Cancer Angiogenesis and Metastases. In: *CXC Chemokines in Cancer Angiogenesis and MetastasesBook*. pp. 91-111. Academic Press, ISBN 0065-230X
- Kilgore, T.; Grewal, A.; Bechtold, M.; Miick, R.; Diaz-Arias, A.; Ibdah, J. & Bragg, J. (2007). Breast Cancer Metastasis To The Colon: A Case Report And Review Of The Literature. *The Internet Journal of Gastroenterology*, Vol. 6, No. 1, pp. on-line, ISSN 1528-8323
- Kirschmann, D.A.; Seftor, E.A.; Nieva, D.R.C.; Mariano, E.A. & Hendrix, M.J.C. (1999). Differentially expressed genes associated with the metastatic phenotype in breast cancer. *Breast cancer research and treatment*, Vol. 55, No. 2, pp. 125-134, 0167-6806
- Koizumi, K.; Hojo, S.; Akashi, T.; Yasumoto, K. & Saiki, I. (2007). Chemokine receptors in cancer metastasis and cancer cell-derived chemokines in host immune response. *Cancer science*, Vol. 98, No. pp. 1652-1658,
- Komori, A.; Yatsunami, J.; Sukanuma, M.; Okabe, S.; Abe, S.; Sakai, A.; Sasaki, K. & Fujiki, H. (1993). Tumor Necrosis Factor Acts as a Tumor Promoter in BALB/3T3 Cell Transformation. *Cancer research*, Vol. 53, No. 9, pp. 1982-1985,
- Kraman, M.;Bambrough, P.J.;Arnold, J.N.;Roberts, E.W.;Magiera, L.;Jones, J.O.;Gopinathan, A.;Tuveson, D.A. & Fearon, D.T. (2010). Suppression of antitumor immunity by stromal cells expressing fibroblast activation protein-alpha. *Science*, Vol.330, No.6005, pp. 827-830, ISSN 1095-9203
- Kundu, N. & Fulton, A.M. (2002). Selective Cyclooxygenase (COX)-1 or COX-2 Inhibitors Control Metastatic Disease in a Murine Model of Breast Cancer. *Cancer research*, Vol. 62, No. 8, pp. 2343-2346,
- Kunz-Schughart, L.A. & Knuechel, R. (2002a). Tumor-associated fibroblasts (part II): Functional impact on tumor tissue. *Histol Histopathol*, Vol.17, No.2, pp. 623-637, ISSN 0213-3911

- Kunz-Schughart, L.A. & Knuechel, R. (2002b). Tumor-associated fibroblasts (part I): Active stromal participants in tumor development and progression? *Histol Histopathol*, Vol.17, No.2, pp. 599-621, ISSN 0213-3911
- Kuperwasser, C.;Chavarria, T.;Wu, M.;Magrane, G.;Gray, J.W.;Carey, L.;Richardson, A. & Weinberg, R.A. (2004). Reconstruction of functionally normal and malignant human breast tissues in mice. *Proceedings of the National Academy of Sciences of the United States of America*, Vol.101, No.14, pp. 4966-4971, ISSN 0027-8424
- Kurebayashi, J. (2000). Regulation of interleukin-6 secretion from breast cancer cells and its clinical implications. *Breast cancer*, Vol. 7, No. 2, pp. 124-129,
- Kurose, K.; Gilley, K.; Matsumoto, S.; Watson, P.H.; Zhou, X.-P. & Eng, C. (2002). Frequent somatic mutations in PTEN and TP53 are mutually exclusive in the stroma of breast carcinomas. *Nat Genet*, Vol. 32, No. 3, pp. 355-357, ISSN 1061-4036
- Kurose, K.; Hoshaw-Woodard, S.; Adeyinka, A.; Lemeshow, S.; Watson, P.H. & Eng, C. (2001). Genetic model of multi-step breast carcinogenesis involving the epithelium and stroma: clues to tumor – microenvironment interactions. *Human molecular genetics*, Vol. 10, No. 18, pp. 1907-1913, ISSN 1460-2083
- Landi, S.; Moreno, V.; Gioia-Patricola, L.; Guino, E.; Navarro, M.; de Oca, J.; Capella, G.; Canzian, F. & Group, f.t.B.C.C.S. (2003). Association of Common Polymorphisms in Inflammatory Genes Interleukin (IL)6, IL8, Tumor Necrosis Factor  $\alpha$ , NFKB1, and Peroxisome Proliferator-activated Receptor  $\alpha$ ? with Colorectal Cancer. *Cancer research*, Vol. 63, No. 13, pp. 3560-3566,
- Langhans, T. (1879). Pulsirende cavernose Geschwulst der Miltz mit metastatischen Knoten in der Leber. *Virchows Archiv*, Vol.75, pp. 273-291, ISSN (N/A)
- Lavergne, E.; Combadiere, B.; Bonduelle, O.; Iga, M.; Gao, J.L.; Maho, M.; Boissonnas, A.; Murphy, P.M.; Debre, P. & Combadiere, C. (2003). Fractalkine mediates natural killer-dependent antitumor responses in vivo. *Cancer research*, Vol. 63, No. pp. 7468-7474,
- LeBeau, A.M.;Brennen, W.N.;Aggarwal, S. & Denmeade, S.R. (2009). Targeting the cancer stroma with a fibroblast activation protein-activated promelittin protoxin. *Molecular cancer therapeutics*, Vol.8, No.5, pp. 1378-1386, ISSN 1538-8514
- Lee, H.O.M., S.R.; Franco-Barraza, J.; Valinaou, M.; Cukierman, E. & Cheng, J.D. (2011). Fap-overexpressing fibroblasts produce an extracellular matrix that enhances invasive velocity and directionality of pancreatic cancer cells. *BMC Cancer*, ISSN Vol. 13, No.11, pp. 245, ISSN
- Leek, R.D.; Landers, R.; Fox, S.B.; Ng, F.; Harris, A.L. & Lewis, C.E. (1998). Association of tumor necrosis factor alpha and its receptors with thymidine phosphorylase expression in invasive breast carcinoma. *British journal of cancer*, Vol. 77, No. 12, pp. 2246-2251,
- Letsch, A.; Keilholz, U.; Schadendorf, D.; Assfalg, G.; Asemissen, A.M.; Thiel, E. & Scheibenbogen, C. (2004). Functional CCR9 Expression Is Associated with Small Intestinal Metastasis. *J Investig Dermatol*, Vol. 122, No. 3, pp. 685-690, ISSN 0022-202X
- Levental, K.R.;Yu, H.;Kass, L.;Lakins, J.N.;Egeblad, M.;Erler, J.T.;Fong, S.F.;Csiszar, K.;Giaccia, A.;Weninger, W.;Yamauchi, M.;Gasser, D.L. & Weaver, V.M. (2009).

- Matrix crosslinking forces tumor progression by enhancing integrin signaling. *Cell*, Vol.139, No.5, pp. 891-906, ISSN 1097-4172
- Li, H.; Fan, X. & Houghton, J. (2007). Tumor microenvironment: the role of the tumor stroma in cancer. *J Cell Biochem*, Vol.101, No.4, pp. 805-815, ISSN 0730-2312
- Li, Y.M.; Pan, Y.; Wei, Y.; Cheng, X.; Zhou, B.P.; Tan, M.; Zhou, X.; Xia, W.; Hortobagyi, G.N.; Yu, D. & Hung, M.-C. (2004). Upregulation of CXCR4 is essential for HER2-mediated tumor metastasis. *Cancer cell*, Vol. 6, No. 5, pp. 459-469, 1535-6108
- Liang, Z.; Wu, T.; Lou, H.; Yu, X.; Taichman, R.S.; Lau, S.K.; Nie, S.; Umbreit, J. & Shim, H. (2004). Inhibition of Breast Cancer Metastasis by Selective Synthetic Polypeptide against CXCR4. *Cancer research*, Vol. 64, No. 12, pp. 4302-4308,
- Liu, W.; Reinmuth, N.; Stoeltzing, O.; Parikh, A.A.; Tellez, C.; Williams, S.; Jung, Y.D.; Fan, F.; Takeda, A.; Akagi, M.; Bar-Eli, M.; Gallick, G.E. & Ellis, L.M. (2003). Cyclooxygenase-2 Is Up-Regulated by Interleukin-1 $\alpha$  in Human Colorectal Cancer Cells via Multiple Signaling Pathways. *Cancer research*, Vol. 63, No. 13, pp. 3632-3636,
- Lochter, A. & Bissell, M.J. (1995). Involvement of extracellular matrix constituents in breast cancer. *Seminars in cancer biology*, Vol.6, No.3, pp. 165-173, ISSN 1044-579X
- Loeffler, M.; Kruger, J.A.; Niethammer, A.G. & Reisfeld, R.A. (2006). Targeting tumor-associated fibroblasts improves cancer chemotherapy by increasing intratumoral drug uptake. *The Journal of clinical investigation*, Vol.116, No.7, pp. 1955-1962, ISSN 0021-9738
- Lu, X. & Kang, Y. (2007). Organotropism of breast cancer metastasis. *Journal of mammary gland biology and neoplasia*, Vol. 12, No. 2-3, pp. 153-162, ISSN 1083-3021
- Luster, A.D. (1998). Chemokines - Chemotactic Cytokines That Mediate Inflammation. *New England Journal of Medicine*, Vol. 338, No. 7, pp. 436-445,
- MacDonald, I.C.; Groom, A.C. & Chambers, A.F. (2002). Cancer spread and micrometastasis development: quantitative approaches for in vivo models. *Bioessays*, Vol. 24, No. pp. 885-893,
- Madhusudan, S.; Foster, M.; Muthuramalingam, S.R.; Braybrooke, J.P.; Wilner, S.; Kaur, K.; Han, C.; Hoare, S.; Balkwill, F.; Talbot, D.C.; Ganesan, T.S. & Harris, A.L. (2004). A phase II study of etanercept (Enbrel), a tumor necrosis factor alpha inhibitor in patients with metastatic breast cancer. *Clinical cancer research : an official journal of the American Association for Cancer Research*, Vol.10, No.19, pp. 6528-6534, ISSN 1078-0432
- Maquart, F.X.; Pasco, S.; Ramont, L.; Hornebeck, W. & Monboisse, J.C. (2004). An introduction to matrikines: extracellular matrix-derived peptides which regulate cell activity. Implication in tumor invasion. *Critical reviews in oncology/hematology*, Vol.49, No.3, pp. 199-202, ISSN 1040-8428
- Mathew, S.; Scanlan, M.J.; Mohan Raj, B.K.; Murty, V.V.; Garin-Chesa, P.; Old, L.J.; Rettig, W.J. & Chaganti, R.S. (1995). The gene for fibroblast activation protein alpha (FAP), a putative cell surface-bound serine protease expressed in cancer stroma and wound healing, maps to chromosome band 2q23. *Genomics*, Vol.25, No.1, pp. 335-337, ISSN 0888-7543

- Matsumiya, T.; Imaizumi, T.; Fujimoto, K.; Cui, X.; Shibata, T.; Tamo, W.; Kumagai, M.; Tanji, K.; Yoshida, H.; Kimura, H. & Satoh, K. (2001). Soluble Interleukin-6 Receptor [alpha] Inhibits the Cytokine-Induced Fractalkine/CX3CL1 Expression in Human Vascular Endothelial Cells in Culture. *Experimental Cell Research*, Vol. 269, No. 1, pp. 35-41, 0014-4827
- Matsumoto, E.; Yoshida, T.; Kawarada, Y. & Sakakura, T. (1999). Expression of fibronectin isoforms in human breast tissue: production of extra domain A+/extra domain B+ by cancer cells and extra domain A+ by stromal cells. *Jpn J Cancer Res*, Vol.90, No.3, pp. 320-325, ISSN 0910-5050
- McAllister, S.S. & Weinberg, R.A. (2010). Tumor-host interactions: a far-reaching relationship. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, Vol.28, No.26, pp. 4022-4028, ISSN 1527-7755
- Mersmann, M.; Schmidt, A.; Rippmann, J.F.; Wuest, T.; Brocks, B.; Rettig, W.J.; Garin-Chesa, P.; Pfizenmaier, K. & Moosmayer, D. (2001). Human antibody derivatives against the fibroblast activation protein for tumor stroma targeting of carcinomas. *International journal of cancer. Journal international du cancer*, Vol.92, No.2, pp. 240-248, ISSN 0020-7136
- Miller, L.J.; Kurtzman, S.H.; Wang, Y.; Anderson, K.H.; Lindquist, R.R. & Kreutzer, D.L. (1998). Expression of interleukin-8 receptors on tumor cells and vascular endothelial cells in human breast cancer tissue. *Anticancer research*, Vol. 18, No. 1A, pp. 77-88, ISSN 0250-7005
- Mimeault, M. & Batra, S.K. (2007). Interplay of distinct growth factors during epithelial mesenchymal transition of cancer progenitor cells and molecular targeting as novel cancer therapies. *Annals of oncology : official journal of the European Society for Medical Oncology / ESMO*, Vol.18, No.10, pp. 1605-1619, ISSN 1569-8041
- Mintz, B. & Illmensee, K. (1975). Normal genetically mosaic mice produced from malignant teratocarcinoma cells. *Proceedings of the National Academy of Sciences of the United States of America*, Vol.72, No.9, pp. 3585-3589, ISSN 0027-8424
- Mishra, P.; Banerjee, D. & Ben-Baruch, A. (2011). Chemokines at the crossroads of tumor-fibroblast interactions that promote malignancy. *J Leukoc Biol*, Vol. 89, No. 1, pp. 31-39, 1938-3673 (Electronic)
- Moinfar, F.; Man, Y.G.; Arnould, L.; Bratthauer, G.L.; Ratschek, M. & Tavassoli, F.A. (2000). Concurrent and Independent Genetic Alterations in the Stromal and Epithelial Cells of Mammary Carcinoma: Implications for Tumorigenesis. *Cancer research*, Vol. 60, No. 9, pp. 2562-2566, ISSN: 1538-7445
- Moser, B. & Loetscher, P. (2001). Lymphocyte traffic control by chemokines. *Nat Immunol*, Vol. 2, No. 2, pp. 123-128, ISSN 1529-2908
- Moser, B. & Willmann, K. (2004). Chemokines: role in inflammation and immune surveillance. *Annals of the Rheumatic Diseases*, Vol. 63, No. suppl 2, pp. ii84-ii89, ISSN 1468-2060
- Mourad, P.D.; Farrell, L.; Stamps, L.D.; Chicoine, M.R. & Silbergeld, D.L. (2005). Why are systemic glioblastoma metastases rare? Systemic and cerebral growth of mouse glioblastoma. *Surgical neurology*, Vol. 63, No. 6, pp. 511-519, ISSN 0090-3019

- Mueller, M.M. & Fusenig, N.E. (2002). Tumor-stroma interactions directing phenotype and progression of epithelial skin tumor cells. *Differentiation*, Vol.70, No.9-10, pp. 486-497, ISSN 1538-7445
- Muller, A.; Homey, B.; Soto, H.; Ge, N.; Catron, D.; Buchanan, M.E.; McClanahan, T.; Murphy, E.; Yuan, W.; Wagner, S.N.; Barrera, J.L.; Mohar, A.; Verastegui, E. & Zlotnik, A. (2001). Involvement of chemokine receptors in breast cancer metastasis. *Nature medicine*, Vol. 410, No. pp. 50-56,
- Nanki, T.; Imai, T.; Nagasaka, K.; Urasaki, Y.; Nonomura, Y.; Taniguchi, K.; Hayashida, K.; Hasegawa, J.; Yoshie, O. & Miyasaka, N. (2002). Migration of CX3CR1-positive T cells producing type 1 cytokines and cytotoxic molecules into the synovium of patients with rheumatoid arthritis. *Arthritis & Rheumatism*, Vol. 46, No. 11, pp. 2878-2883, 1529-0131
- Nevo, I.; Sagi-Assif, O.; Meshel, T.; Geminder, H.; Goldberg-Bittman, L.; Ben-Menachem, S.; Shalmon, B.; Goldberg, I.; Ben-Baruch, A. & Witz, I.P. (2004). The tumor microenvironment: CXCR4 is associated with distinct protein expression patterns in neuroblastoma cells. *Immunology Letters*, Vol. 92, No. pp. 163-169,
- O'Sullivan, C. & Lewis, C.E. (1994). Tumor-associated leucocytes: Friends or foes in breast carcinoma. *The Journal of pathology*, Vol. 172, No. 3, pp. 229-235, 1096-9896
- Offersen, B.V.; Knap, M.M.; Marcussen, N.; Horsman, M.R.; Hamilton-Dutoit, S. & Overgaard, J. (2002). Intense inflammation in bladder carcinoma is associated with angiogenesis and indicates good prognosis. *British journal of cancer*, Vol. 87, No. 12, pp. 1422-1430,
- Okada, N.; Gao, J.-Q.; Sasaki, A.; Niwa, M.; Okada, Y.; Nakayama, T.; Yoshie, O.; Mizuguchi, H.; Hayakawa, T.; Fujita, T.; Yamamoto, A.; Tsutsumi, Y.; Mayumi, T. & Nakagawa, S. (2004). Anti-tumor activity of chemokine is affected by both kinds of tumors and the activation state of the host's immune system: implications for chemokine-based cancer immunotherapy. *Biochemical and Biophysical Research Communications*, Vol. 317, No. 1, pp. 68-76, 0006-291X
- Okamoto, M.; Kawamata, H.; Kawai, K. & Oyasu, R. (1995). Enhancement of Transformation in Vitro of a Nontumorigenic Rat Urothelial Cell Line by Interleukin 6. *Cancer research*, Vol. 55, No. 20, pp. 4581-4585,
- Orimo, A.; Gupta, P.B.; Sgroi, D.C.; Arenzana-Seisdedos, F.; Delaunay, T.; Naeem, R.; Carey, V.J.; Richardson, A.L. & Weinberg, R.A. (2005). Stromal Fibroblasts Present in Invasive Human Breast Carcinomas Promote Tumor Growth and Angiogenesis through Elevated SDF-1/CXCL12 Secretion. *Cell*, Vol. 121, No. 3, pp. 335-348, ISSN 0092-8674
- Orimo, A.; Gupta, P.B.; Sgroi, D.C.; Arenzana-Seisdedos, F.; Delaunay, T.; Naeem, R.; Carey, V.J.; Richardson, A.L. & Weinberg, R.A. (2005). Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion. *Cell*, Vol.121, No.3, pp. 335-348, ISSN 0092-8674
- Ou, D.L.; Chen, C.L.; Lin, S.B.; Hsu, C.H. & L.I. Lin, L.I. (2006). Chemokine receptor expression profiles in nasopharyngeal carcinoma and their association with metastasis and radiotherapy. *Journal of pathology*, Vol. 210, No. pp. 363-373,

- Paget, S. (1889). The distribution of secondary growths in cancer of the breast *Lancet*, Vol.133, pp. 571-573, ISSN
- Pantschenko, A.G.;Pushkar, I.;Anderson, K.H.;Wang, Y.;Miller, L.J.;Kurtzman, S.H.;Barrows, G.&Kreutzer, D.L. (2003). The interleukin-1 family of cytokines and receptors in human breast cancer: implications for tumor progression. *International journal of oncology*, Vol.23, No.2, pp. 269-284, ISSN 1019-6439
- Papageorgis, P.;Lambert, A.W.;Ozturk, S.;Gao, F.;Pan, H.;Manne, U.;Alekseyev, Y.O.;Thiagalingam, A.;Abdolmaleky, H.M.;Lenburg, M. & Thiagalingam, S. (2010). Smad signaling is required to maintain epigenetic silencing during breast cancer progression. *Cancer research*, Vol.70, No.3, pp. 968-978, ISSN 1538-7445
- Petersen, O.W.;Nielsen, H.L.;Gudjonsson, T.;Villadsen, R.;Rank, F.;Niebuhr, E.;Bissell, M.J. & Ronnov-Jessen, L. (2003). Epithelial to mesenchymal transition in human breast cancer can provide a nonmalignant stroma. *The American journal of pathology*, Vol.162, No.2, pp. 391-402, ISSN 0002-9440
- Polyak, K. & Kalluri, R. (2010). The role of the microenvironment in mammary gland development and cancer. *Cold Spring Harb Perspect Biol*, Vol.2, No.11, pp. a003244, ISSN 1943-0264
- Postovit, L.M.;Margaryan, N.V.;Seftor, E.A.;Kirschmann, D.A.;Lipavsky, A.;Wheaton, W.W.;Abbott, D.E.;Seftor, R.E. & Hendrix, M.J. (2008). Human embryonic stem cell microenvironment suppresses the tumorigenic phenotype of aggressive cancer cells. *Proceedings of the National Academy of Sciences of the United States of America*, Vol.105, No.11, pp. 4329-4334, ISSN 1091-6490
- Powell, D.W.;Mifflin, R.C.;Valentich, J.D.;Crowe, S.E.;Saada, J.I. & West, A.B. (1999). Myofibroblasts. I. Paracrine cells important in health and disease. *The American journal of physiology*, Vol.277, No.1 Pt 1, pp. C1-9, ISSN 0002-9513
- Provenzano, P.P.;Eliceiri, K.W.;Campbell, J.M.;Inman, D.R.;White, J.G. & Keely, P.J. (2006). Collagen reorganization at the tumor-stromal interface facilitates local invasion. *BMC Med*, Vol.4, No.1, pp. 38, ISSN 1741-7015
- Quiros, R.M.;Valianou, M.;Kwon, Y.;Brown, K.M.;Godwin, A.K. & Cukierman, E. (2008). Ovarian normal and tumor-associated fibroblasts retain in vivo stromal characteristics in a 3-D matrix-dependent manner. *Gynecologic Oncology*, Vol.110, No.1, pp. 99-109, ISSN 0090-8258
- Radisky, D.C.;Levy, D.D.;Littlepage, L.E.;Liu, H.;Nelson, C.M.;Fata, J.E.;Leake, D.;Godden, E.L.;Albertson, D.G.;Nieto, M.A.;Werb, Z. & Bissell, M.J. (2005). Rac1b and reactive oxygen species mediate MMP-3-induced EMT and genomic instability. *Nature*, Vol.436, No.7047, pp. 123-127, ISSN 1476-4687
- Radisky, E.S. & Radisky, D.C. (2007). Stromal induction of breast cancer: inflammation and invasion. *Reviews in endocrine & metabolic disorders*, Vol.8, No.3, pp. 279-287, ISSN 1389-9155
- Rajagopalan, H.; Nowak, M.A.; Vogelstein, B. & Lengauer, C. (2003). The significance of unstable chromosomes in colorectal cancer. *Nature reviews. Cancer*, Vol. 3, No. 9, pp. 695-701, ISSN 1474-175X
- Rasanen, K. & Vaheri, A. (2010). Activation of fibroblasts in cancer stroma. *Exp Cell Res*, Vol.316, No.17, pp. 2713-2722, ISSN 1090-2422



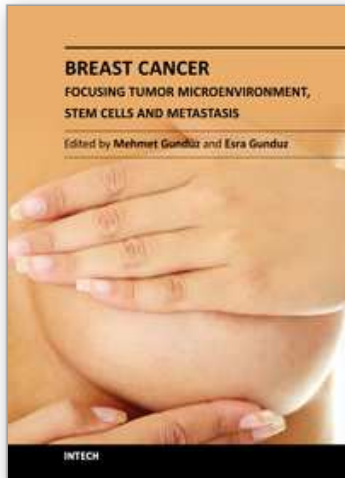
- Reardon, D.A.;Akabani, G.;Coleman, R.E.;Friedman, A.H.;Friedman, H.S.;Herndon, J.E., 2nd;McLendon, R.E.;Pegram, C.N.;Provenzale, J.M.;Quinn, J.A.;Rich, J.N.;Vredenburgh, J.J.;Desjardins, A.;Gururangan, S.;Badruddoja, M.;Dowell, J.M.;Wong, T.Z.;Zhao, X.G.;Zalutsky, M.R. & Bigner, D.D. (2006). Salvage radioimmunotherapy with murine iodine-131-labeled antitenascin monoclonal antibody 81C6 for patients with recurrent primary and metastatic malignant brain tumors: phase II study results. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, Vol.24, No.1, pp. 115-122, ISSN 1527-7755
- Robert, N.;Krekow, L.;Stokoe, C.;Clawson, A.;Iglesias, J. & O'Shaughnessy, J. (2011). Adjuvant dose-dense doxorubicin plus cyclophosphamide followed by dose-dense nab-paclitaxel is safe in women with early-stage breast cancer: a pilot study. *Breast cancer research and treatment*, Vol.125, No.1, pp. 115-120, ISSN 1573-7217
- Robinson, S.C.; Scott, K.A.; Wilson, J.L.; Thompson, R.G.; Proudfoot, A.E.I. & Balkwill, F.R. (2003). A Chemokine Receptor Antagonist Inhibits Experimental Breast Tumor Growth. *Cancer research*, Vol. 63, No. 23, pp. 8360-8365,
- Rocic, P. & Lucchesi, P.A. (2005). NAD(P)H oxidases and TGF-beta-induced cardiac fibroblast differentiation: Nox-4 gets Smad. *Circulation research*, Vol.97, No.9, pp. 850-852, ISSN 1524-4571
- Ronnov-Jessen, L. & Bissell, M.J. (2009). Breast cancer by proxy: can the microenvironment be both the cause and consequence? *Trends Mol Med*, Vol.15, No.1, pp. 5-13, ISSN 1471-4914
- Ronty, M.J.;Leivonen, S.K.;Hinz, B.;Rachlin, A.;Otey, C.A.;Kahari, V.M. & Carpen, O.M. (2006). Isoform-specific regulation of the actin-organizing protein palladin during TGF-beta1-induced myofibroblast differentiation. *The Journal of investigative dermatology*, Vol.126, No.11, pp. 2387-2396, ISSN 1523-1747
- Ruffini, P.A.; Morandi, P.; Cabioglu, N.; Altundag, K. & Cristofanilli, M. (2007). Manipulating the chemokine-chemokine receptor network to treat cancer. *Cancer*, Vol. 109, No. pp. 2392-2404,
- Ryan, D.G.;Taliana, L.;Sun, L.;Wei, Z.G.;Masur, S.K. & Lavker, R.M. (2003). Involvement of S100A4 in stromal fibroblasts of the regenerating cornea. *Invest Ophthalmol Vis Sci*, Vol.44, No.10, pp. 4255-4262, ISSN 0146-0404
- Saisho, S.; Takashima, S.; Ohsumi, S.; Saeki, S.; Aogi, K.; Saeki, T.; Mandai, K.; Iwata, S. & Takeda, T. (2005). Two Cases with Long-Term Disease-Free Survival after Resection and Radiotherapy for Solitary Brain Metastasis from Breast Cancer with Extensive Nodal Metastases. *Breast cancer*, Vol. 12, No. pp. 221-225, ISSN 1880-4233
- Sallusto, F.; Mackay, C.R. & Lanzavecchia, A. (2000). The role of chemokine receptors in primary, effector, and memory immune responses. *Annual review of immunology*, Vol. 18, No. pp. 593-620,
- Santos, A.M.;Jung, J.;Aziz, N.;Kissil, J.L. & Pure, E. (2009). Targeting fibroblast activation protein inhibits tumor stromagenesis and growth in mice. *The Journal of clinical investigation*, Vol.119, No.12, pp. 3613-3625, ISSN 1558-8238
- Schimanski, C.C.; Galle, P.R. & Moehler, M. (2008). Chemokine receptor CXCR4- prognostic factor for gastrointestinal tumors. *World journal of gastroenterology*, Vol. 14, No. pp. 4721-4724,

- Schmid, E.; Osborn, M.; Rungger-Brandle, E.; Gabbiani, G.; Weber, K. & Franke, W.W. (1982). Distribution of vimentin and desmin filaments in smooth muscle tissue of mammalian and avian aorta. *Exp Cell Res*, Vol.137, No.2, pp. 329-340, ISSN 0014-4827
- Schonbeck, U.; Mach, F. & Libby, P. (1998). Generation of biologically active IL-1 beta by matrix metalloproteinases: a novel caspase-1-independent pathway of IL-1 beta processing. *Journal of immunology*, Vol.161, No.7, pp. 3340-3346, ISSN 0022-1767
- Schwarz, M.; Wahl, M.; Resch, K. & Radeke, H.H. (2002). IFN $\gamma$  induces functional chemokine receptor expression in human mesangial cells. *Clinical & Experimental Immunology*, Vol. 128, No. 2, pp. 285-294, 1365-2249
- Seaton, A.; Maxwell, P.J.; Hill, A.; Gallagher, R.; Pettigrew, J.; Wilson, R.H. & Waugh, D.J.J. (2009). Inhibition of constitutive and cxc-chemokine-induced NF- $\kappa$ B activity potentiates ansamycin-based HSP90-inhibitor cytotoxicity in castrate-resistant prostate cancer cells. *British journal of cancer*, Vol. 101, No. 9, pp. 1620-1629, 0007-0920
- Semenza, G.L. & Wang, G.L. (1992). A nuclear factor induced by hypoxia via de novo protein synthesis binds to the human erythropoietin gene enhancer at a site required for transcriptional activation. *Mol. Cell. Biol.*, Vol. 12, No. 12, pp. 5447-5454,
- Shao, Z.M.; Nguyen, M. & Barsky, S.H. (2000). Human breast carcinoma desmoplasia is PDGF initiated. *Oncogene*, Vol.19, No.38, pp. 4337-4345, ISSN 0950-9232
- Shields, J.D.; Fleury, M.E.; Yong, C.; Tomei, A.A.; Randolph, G.J. & Swartz, M.A. (2007). Autologous chemotaxis as a mechanism of tumor cell homing to lymphatics via interstitial flow and autocrine CCR7 signalling. *Cancer cell*, Vol. 11, No. pp. 526-538,
- Shulby, S.A.; Dolloff, N.G.; Stearns, M.E.; Meucci, O. & Fatatis, A. (2004). CX3CR1-fractalkine expression regulates cellular mechanisms involved in adhesion, migration, and survival of human prostate cancer cells. *Cancer research*, Vol. 64, No. pp. 4693-4698,
- Silzle, T.; Randolph, G.J.; Kreutz, M. & Kunz-Schughart, L.A. (2004). The fibroblast: sentinel cell and local immune modulator in tumor tissue. *International journal of cancer*. *Journal international du cancer*, Vol.108, No.2, pp. 173-180, ISSN 0020-7136
- Singer, C.F.; Kronsteiner, N.; Marton, E.; Kubista, M.; Cullen, K.J.; Hirtenlehner, K.; Seifert, M. & Kubista, E. (2002). MMP-2 and MMP-9 expression in breast cancer-derived human fibroblasts is differentially regulated by stromal-epithelial interactions. *Breast cancer research and treatment*, Vol.72, No.1, pp. 69-77, ISSN 0167-6806
- Singh, S.; Singh, U.P.; Grizzle, W.E. & J.W. Lillard, J.W.J. (2004). CXCL12-CXCR4 interactions modulate prostate cancer cell migration, metalloproteinase expression and invasion. *Laboratory investigation*, Vol. 84, No. pp. 1666-1676,
- Soria, G. & Ben-Baruch, A. (2008). The inflammatory chemokines CCL2 and CCL5 in breast cancer. *Cancer letters*, Vol. 267, No. 2, pp. 271-285, 0304-3835
- Soto, A.M. & Sonnenschein, C. (2004). The somatic mutation theory of cancer: growing problems with the paradigm? *Bioessays*, Vol.26, No.10, pp. 1097-1107, ISSN 0265-9247

- Strieter, R.M.; Burdick, M.D.; Mestas, J.; Gomperts, B.; Keane, M.P. & Belperio, J.A. (2006). Cancer CXC chemokine networks and tumor angiogenesis. *European journal of cancer (Oxford, England : 1990)*, Vol. 42, No. 6, pp. 768-778, ISSN 0959-8049
- Struyf, S.; Salogni, L.; Burdick, M.D.; Vandercappellen, J.; Gouwy, M.; Noppen, S.; Proost, P.; Opdenakker, G.; Parmentier, M.; Gerard, C.; Sozzani, S.; Strieter, R.M. & Van Damme, J. (2011). Angiostatic and chemotactic activities of the CXC chemokine CXCL4L1 (platelet factor-4 variant) are mediated by CXCR3. *Blood*, Vol. 117, No. 2, pp. 480-488, ISSN 1528-0020
- Sugimoto, H.;Mundel, T.M.;Kieran, M.W. & Kalluri, R. (2006). Identification of fibroblast heterogeneity in the tumor microenvironment. *Cancer biology & therapy*, Vol.5, No.12, pp. 1640-1646, ISSN 1538-4047
- Sullivan, N.J.;Sasser, A.K.;Axel, A.E.;Vesuna, F.;Raman, V.;Ramirez, N.;Oberyszyn, T.M. & Hall, B.M. (2009). Interleukin-6 induces an epithelial-mesenchymal transition phenotype in human breast cancer cells. *Oncogene*, Vol.28, No.33, pp. 2940-2947, ISSN 1476-5594
- Tai, I.T. & Tang, M.J. (2008). SPARC in cancer biology: its role in cancer progression and potential for therapy. *Drug Resist Updat*, Vol.11, No.6, pp. 231-246, ISSN 1532-2084
- Takahashi, N.;Haba, A.;Matsuno, F. & Seon, B.K. (2001). Antiangiogenic therapy of established tumors in human skin/severe combined immunodeficiency mouse chimeras by anti-endoglin (CD105) monoclonal antibodies, and synergy between anti-endoglin antibody and cyclophosphamide. *Cancer research*, Vol.61, No.21, pp. 7846-7854, ISSN 0008-5472
- Takanami, I. (2003). Overexpression of CCR7 mRNA in nonsmall cell lung cancer: correlation with lymph node metastasis. *International journal of cancer*, Vol. 105, No. pp. 186-189,
- Tarin, D. & Croft, C.B. (1969). Ultrastructural features of wound healing in mouse skin. *J Anat*, Vol.105, No.Pt 1, pp. 189-190, ISSN 0021-8782
- Taylor, M.A.;Parvani, J.G. & Schiemann, W.P. (2010). The pathophysiology of epithelial-mesenchymal transition induced by transforming growth factor-beta in normal and malignant mammary epithelial cells. *Journal of mammary gland biology and neoplasia*, Vol.15, No.2, pp. 169-190, ISSN 1573-7039
- Tenney, R.M. & Discher, D.E. (2009). Stem cells, microenvironment mechanics, and growth factor activation. *Curr Opin Cell Biol*, Vol.21, No.5, pp. 630-635, ISSN 1879-0410
- Tesarov, P.; Kvasnika, J.; Umlaufov, A.; Homolkov, H.; Jirsa, M. & Tesar, V. (2000). Soluble TNF and IL-2 receptors in patients with breast cancer. *Medical science monitor*, Vol. 6, No. 4, pp. 661-664,
- Thomasset, N.;Lochter, A.;Sympson, C.J.;Lund, L.R.;Williams, D.R.;Behrendtsen, O.;Werb, Z. & Bissell, M.J. (1998). Expression of autoactivated stromelysin-1 in mammary glands of transgenic mice leads to a reactive stroma during early development. *The American journal of pathology*, Vol.153, No.2, pp. 457-467, ISSN 0002-9440
- Tlsty, T.D. (2001). Stromal cells can contribute oncogenic signals. *Seminars in cancer biology*, Vol. 11, No. 2, pp. 97-104, 1044-579X

- Ilstiy, T.D. & Hein, P.W. (2001). Know thy neighbor: stromal cells can contribute oncogenic signals. *Current Opinion in Genetics & Development*, Vol. 11, No. 1, pp. 54-59, 0959-437X
- Tomasek, J.J.; Gabbiani, G.; Hinz, B.; Chaponnier, C. & Brown, R.A. (2002). Myofibroblasts and mechano-regulation of connective tissue remodelling. *Nature reviews. Molecular cell biology*, Vol.3, No.5, pp. 349-363, ISSN 1471-0072
- Toullec, A.; Gerald, D.; Despouy, G.; Bourachot, B.; Cardon, M.; Lefort, S.; Richardson, M.; Rigai, G.; Parrini, M.C.; Lucchesi, C.; Bellanger, D.; Stern, M.H.; Dubois, T.; Sastre-Garau, X.; Delattre, O.; Vincent-Salomon, A. & Mechta-Grigoriou, F. (2010). Oxidative stress promotes myofibroblast differentiation and tumor spreading. *EMBO molecular medicine*, Vol.2, No.6, pp. 211-230, ISSN 1757-4684
- Valdivia-Silva, J.E.; Franco-Barraza, J.; Silva, A.L.; Pont, G.D.; Soldevila, G.; Meza, I. & Garcia-Zepeda, E.A. (2009). Effect of pro-inflammatory cytokine stimulation on human breast cancer: implications of chemokine receptor expression in cancer metastasis. *Cancer letters*, Vol. 283, No. 2, pp. 176-185, 1872-7980 (Electronic)
- van Diest, P.J. & Baak, J.P. (1991). The morphometric prognostic index is the strongest prognosticator in premenopausal lymph node-negative and lymph node-positive breast cancer patients. *Human pathology*, Vol. 22, No. pp. 326-330,
- Vishnu, P. & Roy, V. (2010). nab-paclitaxel: a novel formulation of taxane for treatment of breast cancer. *Womens Health (Lond Engl)*, Vol.6, No.4, pp. 495-506, ISSN 1745-5065
- Volk, L.D.; Flister, M.J.; Chihade, D.; Desai, N.; Trieu, V. & Ran, S. (2011). Synergy of Nab-paclitaxel and Bevacizumab in Eradicating Large Orthotopic Breast Tumors and Preexisting Metastases. *Neoplasia*, Vol.13, No.4, pp. 327-338, ISSN 1476-5586
- Voorzanger, N.; Tuitou, R.; Garcia, E.; Delecluse, H.-J.; Rousset, F.o.; Joab, I.n.; Favrot, M.C. & Blay, J.-Y. (1996). Interleukin (IL)-10 and IL-6 Are Produced in Vivo by Non-Hodgkin's Lymphoma Cells and Act as Cooperative Growth Factors. *Cancer research*, Vol. 56, No. 23, pp. 5499-5505,
- Walker, R.A. & Dearing, S.J. (1992). Transforming growth factor beta 1 in ductal carcinoma in situ and invasive carcinomas of the breast. *European journal of cancer*, Vol.28, No.2-3, pp. 641-644, ISSN 0959-8049
- Walker, R.A.; Dearing, S.J. & Gallacher, B. (1994). Relationship of transforming growth factor beta 1 to extracellular matrix and stromal infiltrates in invasive breast carcinoma. *British journal of cancer*, Vol.69, No.6, pp. 1160-1165, ISSN 0007-0920
- Wang, J.; Xi, L.; Gooding, W.; Godfrey, T.E. & Ferris, R.L. (2005). Chemokine receptors 6 and 7 identify a metastatic expression pattern in squamous cell carcinoma of the head and neck. *Advances in otorhinolaryngology*, Vol. 62, No. pp. 121-133,
- Wipff, P.J. & Hinz, B. (2008). Integrins and the activation of latent transforming growth factor beta1 - an intimate relationship. *European journal of cell biology*, Vol.87, No.8-9, pp. 601-615, ISSN 0171-9335
- Wipff, P.J.; Rifkin, D.B.; Meister, J.J. & Hinz, B. (2007). Myofibroblast contraction activates latent TGF-beta1 from the extracellular matrix. *The Journal of cell biology*, Vol.179, No.6, pp. 1311-1323, ISSN 1540-8140

- Wong, Y.C. & Tam, N.N. (2002). Dedifferentiation of stromal smooth muscle as a factor in prostate carcinogenesis. *Differentiation*, Vol.70, No.9-10, pp. 633-645, ISSN 0301-4681
- Woodworth, C.D.; McMullin, E.; Iglesias, M. & Plowman, G.D. (1995). Interleukin 1 alpha and tumor necrosis factor alpha stimulate autocrine amphiregulin expression and proliferation of human papillomavirus-immortalized and carcinoma-derived cervical epithelial cells. *Proceedings of the National Academy of Sciences*, Vol. 92, No. 7, pp. 2840-2844,
- Wu, Y.; Deng, J.; Rychahou, P.G.; Qiu, S.; Evers, B.M. & Zhou, B.P. (2009). Stabilization of snail by NF-kappaB is required for inflammation-induced cell migration and invasion. *Cancer cell*, Vol.15, No.5, pp. 416-428, ISSN 1878-3686
- Xouri, G. & Christian, S. (2010). Origin and function of tumor stroma fibroblasts. *Seminars in cell & developmental biology*, Vol.21, No.1, pp. 40-46, ISSN 1096-3634
- Xu, S.-G.; Yan, P.-J. & Shao, Z.-M. (2010). Differential proteomic analysis of a highly metastatic variant of human breast cancer cells using two-dimensional differential gel electrophoresis. *Journal of Cancer Research and Clinical Oncology*, Vol. 136, No. 10, pp. 1545-1556, 0171-5216
- Yaal-Hahoshen, N.; Shina, S.; Leider-Trejo, L.; Barnea, I.; Shabtai, E.L.; Azenshtein, E.; Greenberg, I.; Keydar, I. & Ben-Baruch, A. (2006). The chemokine CCL5 as a potential prognostic factor predicting disease progression in stage II breast cancer patients. *Clinical cancer research*, Vol. 12, No. pp. 4474- 4480,
- Yang, L.; Pang, Y. & Moses, H.L. (2010). TGF-beta and immune cells: an important regulatory axis in the tumor microenvironment and progression. *Trends in immunology*, Vol.31, No.6, pp. 220-227, ISSN 1471-4981
- Zeisberg, E.M.; Potenta, S.; Xie, L.; Zeisberg, M. & Kalluri, R. (2007). Discovery of endothelial to mesenchymal transition as a source for carcinoma-associated fibroblasts. *Cancer research*, Vol.67, No.21, pp. 10123-10128, ISSN 1538-7445
- Zhou, W.; Guo, S. & Gonzalez-Perez, R.R. (2011). Leptin pro-angiogenic signature in breast cancer is linked to IL-1 signalling. *British journal of cancer*, Vol. 104, No. 1, pp. 128-137, 0007-0920
- Zlotnik, A. (2006). Chemokines and cancer. *International journal of cancer*, Vol. 119, No. 9, pp. 2026-2029,
- Zlotnik, A. (2008). New insights on the role of CXCR4 in cancer metastasis. *The Journal of pathology*, Vol. 215, No. 3, pp. 211-213, ISSN 1096-9896



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Cancer is the leading cause of death in most countries and its consequences result in huge economic, social and psychological burden. Breast cancer is the most frequently diagnosed cancer type and the leading cause of cancer death among females. In this book, we discussed characteristics of breast cancer cell, role of microenvironment, stem cells and metastasis for this deadly cancer. We hope that this book will contribute to the development of novel diagnostic as well as therapeutic approaches.

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