1. Introduction

Rapidly progressive glomerulonephritis (RPGN) is one of the nephrology emergencies which needs special attention. RPGN is a clinical description which determines by symptoms and signs of glomerulonephritis (GN); edema, hypertension and gross hematuria, and evidence of acute renal failure (severe decrease in glomerular filtration rate presents as oliguria or anuria, and increased serum levels of BUN and creatinine). Definite diagnosis of the disorder is based on kidney biopsy's findings. Early diagnosis and appropriate treatment plays a critical role in renal saving and preventing permanent glomerular damage.

This chapter will focus on clinical manifestations, therapeutic protocols and prognostic factors in patients with different subtypes of RPGN.

Rapidly progressive glomerulonephritis (RPGN) is defined as a syndrome with abrupt or insidious onset of hematuria, proteinuria, anemia, and rapidly progressing acute renal failure (ARF), and special findings on light microscopy examination of kidney biopsy's specimen; crescentic lesions which usually involved most glomerular architectures (Hirayama, et al., 2008; Rutgers et al., 2004). It also characterized by rapid loss of renal function (GFR<50% within 3 months) with histological findings of crescent lesions which usually involves>50 % of glomeruli (Couser, 1988).

RPGN can be primary or secondary. Secondary forms occur in any form of severe glomerulonephritis including membranoproliferative GN, IgA nephropathy, post infectious GN, and systemic lupus erythematous (SLE). Primary RPGN is an autoimmune disease which is divided into three immunopathologic categories:(Rutgers et al., 2004; Haas & Eustace, 2004):

Type 1 RPGN: glomerulonephritis with antibodies directed against the glomerular basement membrane (GBM) (anti-GBM mediated GN)

Type II RPGN: immune-complex induced glomerulonephritis

Type III RPGN: Antineutrophil cytoplasmic antibody associated glomerulonephritis (ANCA-associated glomerulonephritis or pauci-immune GN)

RPGN type 1 and 2 are responsible for 10-20 % and 40 % of all cases respectively. RPGN type 2 can be found in different forms of systemic disease such as post infectious GN(PSGN), Ig-A nephropathy, Henoch–Schönlein purpura (HSP), SLE, membranous GN(MGN) or membrano- proliferative GN(MPGN). A few cases of idiopathic immune-complex-mediated RPGN have been reported (Jindal, 1999).
Interestingly different forms of RPGN share similar clinical features including hematuria, proteinuria, edema, hypertension and symptoms of ARF. Patients with Anti-GBM antibody or pauci-immune RPGN (ANCA-associated vasculitis) may have pulmonary hemorrhage and hemoptysis (Jindal, 1999). In pauci-immune RPGN the initial symptoms are non-specific; often fatigue, fever, night sweats and artheralgias are first clinical manifestations (Jindal, 1999).

2. Histopathology characteristics of RPGN and laboratory findings

Variable clinical manifestations and non-specific histologic changes complicate diagnosis and classification of vasculitis. To confirm the diagnosis light microscopy and immunofluorescence examinations of kidney biopsy should be accompanied by appropriate serologic tests, including ANCA (Vizjak et al., 2003).

2.1 Light microscopy findings

Histopathologically, RPGN is characterized by vasculitis which involves glomerular capillaries, and results in formation of cellular crescents within most glomeruli (Hricik et al., 1998). The hallmark histologic lesions are crescents; a morphologic expression of severe glomerular injury. In severe glomerular injury rupture of the glomerular capillaries allows inflammatory mediators to spill into Bowman’s space, resulting in epithelial cell proliferation and invasion of monocyte and macrophage to Bowman’s space (Couser, 1988; Jennette & Falk, 1998; Jennette & Thomas, 2001). Crescents are divided into cellular, fibro cellular or fibrous types. Halmarks of irreversible glomerular or tubulointerstitial injuries are glomerular sclerosis, fibrous or fibro cellular crescents, and interstitial fibrosis. The lesions usually are seen in various stages of activity or resolution. Necrotizing inflammation in small cortical arteries is reported in 10% of biopsy specimens. Inflammation of medullary vasa rectae with papillary necrosis is another finding that may be found (Lionakiet al., 2007).

In acute pauci-immune glomerulonephritis (RPGN type III) fibrinoid necrosis accompanies crescents. These lesions occur at the same frequency irrespective of the presence or absence of associated vasculitis (DAgati et al., 1986). Acute lesions range from focal segmental fibrinoid necrosis affecting less than 10% of glomeruli to severe diffuse necrotizing and crescentic glomerulonephritis that may injure all glomeruli. Periglomerular granulomatous inflammation may occur, but is not specific for pauci-immune glomerulonephritis (Lionakiet al., 2007).

3. Histopathology characteristics on immunofluorescent microscopy

Anti-GBM glomerulonephritis characterized by linear staining for IgG and usually C3 along the glomerular capillary. Immune complex-mediated glomerulonephritis, which is found in severe forms of various types of glomerulonephritis such as PSGN, IgA nephropathy, and lupus nephritis, characterized by granular glomerular staining for one or more immunoglobulins and/or complement components, and pauci-immune glomerulonephritis characterized by mild or absent glomerular tuft staining for immunoglobulins and/or complement (Rutgers et al., 2004).

Anti-neutrophil cytoplasmic antibodies (ANCA) associated glomerulonephritis are usually pauci immune; however, immunofluorescence microscopy often reveals a low level of
staining (less than +2, in the 0–4 scale (Harriset al., 1998). Figure 1 presents histologic findings in light and immunofluorescent microscopy.

3.1 Electron microscopy findings
On electron microscopy examination absence of electron-dense immune complex deposits (type I RPGN), multiple electron-dense deposits (type II RPGN), and few or no electron-dense deposits (type III RPGN) are main findings (figure 2) (Haas, M.&Eustace, 2004).

Fig. 1. Histopathologic findings in light and immunofluorescent microscopy (Lionakiet al., 2007) a/left; light microscopic demonstration of ANCA-associated necrotizing GN (with a crescent), arrows; (alveolar capillaritis with intra-alveolar hemorrhage); arrow; middle (and pulmonary necrotizing granulomatous inflammation with a multinucleated giant cell) arrow; right b/middle; Immunofluorescence microscopy can separate crescentic glomerulonephritis into anti-GBM with linear IgG staining, (left) immune complex with granular staining, (right); or pauci-immune categories with little or no immunoglobulin staining.

4. Anti-neutrophil cytoplasmic and anti-GBM antibodies
Anti-neutrophil cytoplasmic antibodies(ANCA) are characteristic markers of small vessel vasculitides; Wegener’s granulomatosis(WG), Microscopic polyangiitis(MPA), Churg-Strauss Syndrome(CSS), and idiopathic pauci-immune necrotizing glomerulonephritis for them the term ANCA associated vasculitides(AAV) has long being used(Jennette et al., 1989).
C: Electron microscopy showing multiple sub-epithelial electron-dense deposits, some appearing partially resorbed, with extension of the glomerular basement membrane (GBM) around the deposits (uranyl acetate and lead citrate stain, original magnification ×6300)

F: Electron microscopy showing a large subepithelial deposit in a “notch” area (arrow), as well as mesangial deposits (uranyl acetate and lead citrate stain, original magnification×3800).

The standard approach for detection of ANCA is indirect immunofluorescence (IIF) technique followed by antigen-specific quantitative assays. Myeloperoxidase (MPO) and proteinase3 (PR3) are major ANCA antigens (Falk & Jennette, 1988; Goldschmeding et al., 1989; Niles et al., 1989).

In patients with RPGN, there are two major sub classes of ANCA, namely perinuclear (p-ANCA) and cytoplasmic (c-ANCA) (Guillevin et al., 1999). The main epitope of p-ANCA is myeloperoxidase (MPO), and that of c-ANCA is proteinase-3 (PR3) (Asarodet al., 2000). MPO-ANCA is a useful serum marker for MPA and idiopathic pauci-immune crescentic GN, and PR3-ANCA is regarded as a serum marker for Wegener’s granulomatosis and MPA (Asarodet al., 2000).

Majority (approximately 80-85%) of cases of pauci-immune crescentic glomerulonephritis are ANCA positive (Hauere et al., 2002; Jennette, 2003). Greater than 95% of cytoplasmic ANCA are PR3-ANCA and >95% of perinuclear ANCA are MPO-ANCA (Lionakiet al., 2007). The c and p-ANCAs are mostly directed against the azurophilic granule proteins proteinase 3 (PR3) and myeloperoxidase (MPO), respectively. Detection of c and especially p-ANCAs are not equivalent to the presence of PR3 and MPO-ANCA. It is recommended to detect ANCA by an antigen-specific ELISA (Savige et al., 2000; Cohen Tervaert et al., 1991).

Anti-GBM antibodies which are directed to the non-collagenous part of the α3 chain of type IV collagen, can also be evaluated by both IIF and ELISA (Rutgers et al., 2004). The ANCA-GBM dot-blot is a qualitative assay that uses nitrocellulose strips on which purified antigens are blotted at preset spots. MPO and PR3 antigens that are used in these tests are produced from human leukocytes. The GBM-ANCA dot-blot assay has been revealed reactivities that had not been detected by ELISA (Table 1) (Rutgers et al., 2004).
Table 1. Characteristics of the GBM-ANCA Dot-Blot Assay (Rutgers et al., 2004)

<table>
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<tr>
<th>MPO-ANCA</th>
<th>PR3-ANCA</th>
<th>Anti-GBM antibodies</th>
</tr>
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<tbody>
<tr>
<td><strong>Sensitivity</strong></td>
<td>80–86%&lt;sup&gt;a&lt;/sup&gt;</td>
<td>92–95%&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td><strong>Specificity</strong></td>
<td>100%</td>
<td>100%</td>
</tr>
<tr>
<td><strong>Inter-observer effect</strong></td>
<td>5%</td>
<td>1%</td>
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5. **RPGN, Pulmonary–renal syndrome (PRS), and ANCA-associated vasculitis (AAVs)**

The ANCA-associated vasculitis (WG, MPA, and CSS) are a group of rare autoimmune conditions characterized by the development of necrotizing vasculitis. They share a number of clinical features and are therefore treated using similar treatment protocols. The AAVs are rare with an annual incidence of 20/million in Europe, with WG as the most common and CSS the least frequent (Ntatsaki et al., 2010). In far-east, MPA is more common than WG. It's thought that they arise from interaction between an environmental factor and a genetically predisposing agent (Ntatsaki et al., 2011).

Pulmonary–renal syndrome (PRS) is defined as combination of diffuse alveolar hemorrhage (DAH) and glomerulonephritis (Savage et al., 1997; Seo & Stone, 2004; Jennette & Falk, 2003). This syndrome is caused by different diseases, including various forms of primary systemic vasculitis especially WG and MPA, ANCA-associated systemic vasculitis (AVV), Good pasteure's syndrome, SLE, and infection-associated or drug induced glomerulonephritis (Westmann et al., 1997; Brusselle, 2007).

Immunologic injuries or non-immunologic mechanisms are involved in pathogenesis of PRS. Immunologic mechanisms such as production of anti-GBM antibodies, ANCA, immune complexes mediated injuries and non-immunologic mechanisms such as thrombotic microangiopathy (Rondeau et al., 1989) have been suggested. Pulmonary involvement in the majority of cases is the result of small-vessel vasculitis that involves arterioles, venules and alveolar capillaries (necrotic pulmonary capillaritis). These lesions are clinically expressed with DAH (Levy. & Wineals, 1994). In the majority of cases the underlying renal pathology is a form of focal proliferative glomerulonephritis (Jayne et al., 2000), with fibrinoid necrosis, as well as micro vascular thrombi, and extensive crescent formation accompanies glomerular tuft disease (Walters et al., 2010).

According to results of ANCA pulmonary renal syndrome can be categorized into two subgroups:
1. ANCA-positive Pulmonary–renal syndrome
2. ANCA-negative Pulmonary–renal syndrome

CirculatingANCAs are detected in the majority of pulmonary–renal syndromes (The Wegener's Granulomatosis Etanercept Trial [WGET] Research Group 2005; Dickersinet al., 1994.) Three major systemic vasculitis syndromes are associated with positive ANCAs consists of Wegener’s granulomatosis, microscopic polyangiitis and Churg-Strauss syndrome (The Wegener's Granulomatosis Etanercept Trial[ WGET] Research Group, 2005), and idiopathic pulmonary–renal syndrome (Bolton eturgill, 1989).
Majority of cases of pulmonary-renal syndromes are related to ANCA associated vasculitis (Brusselle, 2007). Pulmonary-renal syndrome in ANCA-negative systemic vasculitis is very rare and has been reported in Behçet’s disease, HSP, IgA nephropathy and in mixed cryoglobulinaemia and rarely in thrombotic thrombocytopenic purpura (TTP) (Naseri & Zabolinejad, 2008).

**ANCA-positive Pulmonary-renal syndrome**: More than 80% of patients with necrotizing pauci immune small-vessel vasculitis have circulating ANCA (Jennette & Falk, 1997). The main clinicopathological expressions of AAV are WG, MPA, CSS and renal-limited vasculitis (RLV). The highest incidence is for WG in northern Europe (Haugeberg et al., 1998; Watts et al., 2000, 2008; Lane et al., 2000; Reinhold-Keller et al., 2005), while the incidence of MPA/RLV is higher in Japan (Fujimoto et al., 2006).

**Wegener’s granulomatosis (WG)**: Friedrich Wegener in 1939 was the first investigator to recognize a group of diseases characterized by extra vascular necrotizing granulomatous inflammation of the respiratory tract with vasculitis and/or glomerulonephritis (Schultz & Tozman, 1995). In 1983, Fauci et al reported details of 85 patients with Wegener’s granulomatosis followed over a 21-year period. Their diagnosis was based on upper and lower respiratory tract complications, renal disease, and variable involvement of other organs with disseminated vasculitis. Tissue biopsies confirmed the characteristic clinical findings (Fauci et al., 1983). Pathologically Wegener’s granulomatosis was characterized by small-vessel necrotizing vasculitis and granulomatous inflammation involving mostly the upper and lower respiratory tracts and the kidneys (Sugimoto et al., 2007). Diffuse alveolar hemorrhage is the most serious complication in small-vessel vasculitis. Respiratory symptoms including cough and hemoptysis. CXR or chest CT-scan may reveal diffuse lung infiltrate. Clinical manifestations, pathologic and serologic findings play important role in the diagnosis of WG. 75-90% of patients with active disease have PR3-ANCAs (Langford, 2005; Ozaki, 2007). The role of PR3-ANCA in the pathogenesis of the disease is not clear, but in vitro evidence suggests that PR3-ANCA can directly or indirectly damage endothelial cells (Preston et al., 2002). With the recognition of association between ANCs and Wegener’s granulomatosis, the concept of Wegener’s granulomatosis has been modified and recently a less restrictive definition has been proposed, termed “Wegener’s vasculitis.” This term includes ANCA-positive patients with clinical presentations of WG such as sinusitis, pulmonary infiltrates, nephritis, and documented necrotizing vasculitis, but without biopsy-proven granulomatous inflammation. Both classic Wegener’s granulomatosis and Wegener’s vasculitis are different manifestations of the same disease process. The term “Wegener’s syndrome” is a more generic term suggested by the working classification of ANCA-associated vasculitides (Tervaert, & Stegeman, 2003).

Diagnostic criteria have been reported by the research group of intractable vasculitides, ministry of Health, Labor, and Welfare {MHLW} of Japan for definite diagnosis of AAV including WG (Table 2) (Ozaki, 2007). Necrotizing granulomatous lesions commonly affect the ear, nose and throat (E), lung (L), and kidney (K). Systemic symptoms in WG are classified as the following:

1. **E Symptoms**: nasal symptoms; purulent rhinorrhea, epistaxis, and a saddle nose, eye symptoms; ophthalmic pain, visual disturbance, and exophthalmia, ear symptoms; otalgia and otitis media, and throat symptoms; pharyngeal ulcer, hoarseness, and laryngeal obstruction.
2. lung (L) symptoms: bloody sputa, cough, and dyspnea
3. Kidney (K) symptoms: hematuria, proteinuria, rapidly progressive renal failure, edema, and hypertension

Table 2. Diagnostic criteria for Wegener's granulomatosis (Ozaki, 2007)

Biopsies of the nasal mucosa, lung, and kidney reveal necrotizing granulomatous vasculitis and necrotizing crescentic glomerulonephritis without immune deposits (Ozaki, 2007). WG with E, L, and K involvement is classified as the generalized form, while when there is no kidney involvement, E only, L only, or E + L, the term of limited form is used. The therapeutic strategies are different for each forms (Ozaki, 2007).

Microscopic polyangiitis (MPA)
Renal and pulmonary symptoms are characteristic in MPA, and interstitial pneumonitis and pulmonary hemorrhage are common clinical features. MPO-ANCA is positive in 50-75% of patients and biopsy of the lung and kidney reveals necrotizing vasculitis of arterioles, capillaries, and venules with few immune deposits necrotizing and crescentic GN (Ozaki; 2007). Granulomatous inflammation and asthma are not seen in MPA (Jennette & Falk, 1997).

Table 3 presents diagnostic criteria for microscopic polyangiitis (Ozaki, 2007).
Allergic granulomatous angiitis (AGA) or Churg-Strauss syndrome (CSS)

Churg and Strauss was firstly described allergic granulomatous angiitis (Churg, & Strauss, 1951). The disease is characterized by presence of asthma, eosinophilia, and necrotizing granulomatous inflammation. Clinical manifestations of small-vessel vasculitis; palpable purpura of the lower extremities, mononeuritis multiplex, abdominal pain, and gastrointestinal bleeding develop several years after the onset of asthma. Positive MPO-ANCA are seen and skin biopsy shows necrotizing vasculitis of small vessels with massive eosinophilic infiltration and extravascular granulomatosis (Ozaki, 2007). Table 4 presents diagnostic criteria for CSS (Ozaki, 2007).

<table>
<thead>
<tr>
<th>1. Symptoms</th>
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<tbody>
<tr>
<td>(1) Rapidly progressive glomerulonephritis</td>
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<tr>
<td>(2) Pulmonary hemorrhage</td>
</tr>
<tr>
<td>(3) Other organ symptoms:</td>
</tr>
<tr>
<td>Purpura, subcutaneous hemorrhage, gastrointestinal bleeding, and mononeuritis multiplex</td>
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<table>
<thead>
<tr>
<th>2. Histological findings</th>
</tr>
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<tbody>
<tr>
<td>Necrotizing vasculitis of arterioles, capillaries, and venules, and perivascular infiltration of inflammatory cells</td>
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<table>
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<tr>
<th>3. Laboratory findings</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1) Positive MPO-ANCA</td>
</tr>
<tr>
<td>(2) Positive CRP</td>
</tr>
<tr>
<td>(3) Proteinuria, hematuria, elevation of BUN and serum creatinine</td>
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<Diagnosis>

1. Definite MPA

(1) Positive for 2 or more of the symptoms, and positive histological findings

(2) Positive for 2 or more of the symptoms including the symptoms (1) and (2), and positive MPO-ANCA

2. Probable MPA

(1) Positive for 3 of the symptoms

(2) Positive for 1 of the symptoms, and positive MPO-ANCA

Table 3. Diagnostic criteria for microscopic polyangiitis (Ozaki, 2007)

SLE and AAV

Systemic lupus erythromatosis is an autoimmune disorder. Variety of autoantibodies are present in SLE patients including ANCA which have been reported in 3-69% of cases (Molnare et al., 2002; Edgar et al., 1995; Pradhan et al., 2004). Some studies (Chinet al., 2000; Nishiyama et al., 1997) have reported p-ANCA in 37.3-42% of patients with lupus nephritis, mainly those who have diffuse proliferative GN (DPGN), and minority of patients without renal involvement. In Pradhan et al’s study predominant ANCA pattern was p-ANCA while c-ANCA pattern was not found in any patient (Pradhan et al., 2004). Their study revealed that ANCA can be used as a serological marker to differentiate vasculitides in lupus nephritis cases from SLE without nephritis.
Table 4. Diagnostic criteria for Churg-Strauss syndrome (Ozaki, 2007)

<table>
<thead>
<tr>
<th>1. Symptoms</th>
<th>2. Characteristic clinical course</th>
<th>3. Histological findings</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1) Bronchial asthma and/or allergic rhinitis</td>
<td>(1) Symptoms (1) and (2) precede the development of (3)</td>
<td>(1) Granulomatous or necrotizing vasculitis of small vessels with marked infiltration of eosinophils</td>
</tr>
<tr>
<td>(2) Eosinophilia</td>
<td></td>
<td>(2) Extravascular granulomas</td>
</tr>
<tr>
<td>(3) Symptoms due to vasculitis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(a) General symptoms: fever (38°C or higher, 2 weeks or longer), weight loss (6 kg or more for 6 months)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(b) Local symptoms: mononeuritis multiplex, gastrointestinal bleeding, purpura, polyarthritis/polyarthralgia, and myalg (muscle weakness)</td>
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</table>

Table 5. Correlation of organ involvement with ANCA serology and other autoantibodies in SLE patients (Pradhan et al., 2004)

<table>
<thead>
<tr>
<th>Organ involvement</th>
<th>ANCA serology</th>
<th>Other autoantibodies</th>
</tr>
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<tbody>
<tr>
<td>Skin (50)</td>
<td>anti-MPO (12/59)</td>
<td>ANA (59/59)</td>
</tr>
<tr>
<td></td>
<td>anti-LF (10/59)</td>
<td>anti-dsDNA (45/59)</td>
</tr>
<tr>
<td></td>
<td>anti-CG (8/59)</td>
<td>anti-ssDNA (40/59)</td>
</tr>
<tr>
<td>Renal (41)</td>
<td>12</td>
<td>28</td>
</tr>
<tr>
<td>DPGN with crescents (21)</td>
<td>5</td>
<td>19</td>
</tr>
<tr>
<td>FPGN with crescents (14)</td>
<td>5</td>
<td>12</td>
</tr>
<tr>
<td>RPGN with crescents (4)</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>MPGN with crescents (2)</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>Joint (35) **</td>
<td>0</td>
<td>26</td>
</tr>
<tr>
<td>Serositis (16)</td>
<td>0</td>
<td>14</td>
</tr>
<tr>
<td>Haematological (8)</td>
<td>0</td>
<td>4</td>
</tr>
<tr>
<td>GI tract (8)</td>
<td>0</td>
<td>6</td>
</tr>
<tr>
<td>CNS (8)</td>
<td>0</td>
<td>3</td>
</tr>
</tbody>
</table>

Table 5. Correlation of organ involvement with ANCA serology and other autoantibodies in SLE patients (Pradhan et al., 2004)
Atypical or X-ANCA has been reported among SLE patients by Savige et al. This antibody showing specificities to cathepsin G and lactoferrin (Savige et al., 1996). Table 5 shows correlation of organ involvement with ANCA serology and other autoantibodies in SLE patients (Pradhan et al., 2004).

6. Treatment of RPGN

Untreated RPGN typically progresses to end-stage renal disease over a period of weeks to a few months. Early diagnosis and initiation of appropriate therapy is essential to minimize the degree of irreversible renal injuries. The therapy of most patients involves pulse methylprednisolone followed by daily oral prednisone, oral or intravenous (IV) cyclophosphamide, and in some cases plasmapheresis. Empiric therapy with IV methylprednisolone should be begun in patients with severe disease with adding of plasmapheresis especially if the patient has hemoptysis (Appel et al., 2010). If a renal biopsy performs soon after initiating empiric therapy the histological abnormalities will not alter.

6.1 Treatment of ANCA associated vasculitis (AAV)

Pauci-immune crescentic glomerulonephritis is a severe form of glomerular inflammation, which if left untreated, usually progresses to end-stage renal failure in weeks or months, AAV is responsible for 80% of cases (Sakai et al., 2002). The evidence-based studies for the management of the AAV is well established and the strategy of induction, consolidation and maintenance therapy is accepted. Guidelines have been designed by both the British Society for Rheumatology (Lapraik et al., 2007) and European League Against Rheumatism (Mukhtyar et al., 2009) on the management of the AAVs. The AAVs are conventionally treated with a strategy of remission induction followed by maintenance therapy using glucocorticoids combined with cyclophosphamide during induction and azathioprine (AZA) for maintenance (Ntatsaki et al., 2011). Current standard treatment is combination of cyclophosphamide and steroids, but the optimal doses, routes of administration, and duration of therapy remain poorly defined (Hotta, et al., 2005). As immunosuppressive therapies increases the risk of infection, therefore one of the most important aspect of successful treatment strategies should be sufficient attenuation of inflammation without serious immunosuppression which leading to life-threatening infection.

Corticosteroids

For the induction of remission corticosteroid regimen is recommended which includea daily intravenous pulse of methylprednisolone (15 mg/kg) for 3 days, followed by oral prednisone (1 mg/kg/day) for 3 weeks, which then tapered progressively (Pagnoux et al., 2008). In pauci-immune RPGN and MPA usefulness of combination of methylprednisolone pulse therapy for 3 days, oral corticosteroid of 1 mg/kg/day for 1 month, and cyclophosphamide of 2 mg/kg/day for 6 to 12 months has been confirmed in several studies (Salama, et al., 2002; Jindal, 1999; Hoffman, 1997; Guillemin et al., 1991).

Cyclophosphamide (CYC)

Different therapeutic agents have been used in treatment of AAV. Standard therapy for AAV is treatment with combination of cyclophosphamide and prednisolone (Pagnoux et al., 2008). After induction with daily oral or pulse intravenous cyclophosphamide therapy, relapse rates of 15% at 12 months (Haubitz et al., 1998), and 38% at 30 months (Guillemin et al., 1997)
have been reported. In long-time follow-up 50% of patients experience relapse within 5 years (Hoffman et al., 1992). Treatment with cyclophosphamide is effective, but also very toxic (Hoffman et al., 1992). Repeated treatment with cyclophosphamide increases adverse effects. To avoid cyclophosphamide-related toxicity, alternative induction treatments are needed (Stassen et al., 2007).

**Mycophenolate mofetil (MMF):** MMF is considered as a potent immuno suppressive drug with favorable side effects, so it has been considered as an alternative to cyclophosphamide treatment in patients with AAV (Stassen et al., 2007; Pesavento et al., 1999; Haidinger et al., 2000; Joy et al., 2005; Koukoulaki, & Jayne, 2006). MMF is a drug which usually is well tolerated. In patients with auto-immune diseases such as SLE, MMF is effective in inducing remission with short-term efficacy which is comparable with cyclophosphamide (Chan et al., 2000, 2005; Ong et al., 2005; Ginzler et al., 2005). Induction treatment with MMF and oral steroids consisted of oral MMF 1000 mg twice daily and oral prednisolone 1 mg/kg once daily (maximum 60 mg). If patients are still in remission after 1 year, MMF is tapered by 500 mg every 3 months and Prednisolone is tapered after 6 weeks by 10 mg every 2 weeks until a dose of 30 mg was reached, and by 5 mg every 2 weeks until 10 mg. Next, the dose is reduced 2.5 mg every month (Stassen et al., 2007). Stassen et al reported that combination of oral steroids and MMF induced complete and partial remission in 78% and 19% of patients respectively (Stassen et al., 2007). Therefore they suggested oral steroids with MMF in patients with relapses of AAV intolerant to cyclophosphamide therapy. Bone marrow suppression is an uncommon side effect of MMF treatment which can result in anemia, leucocytopenia or thrombocytopenia in a number of patients. Fortunately this side effect responds to temporary dose reduction in nearly all patients (Stassen et al., 2007).

**Methotrexate (MTX)**

For remission maintenance in patients who are intolerant of AZA or relapse while taking it other alternative agents such as MTX or MMF have been recommended (Ntatsaki et al., 2011). Methotrexate is an alternative drug for cyclophosphamide which is currently studied (De Groot et al., 2005; Sneller et al., 1995; Stone et al., 1999). According to one randomized (De Groot et al., 2005) and two uncontrolled studies (Sneller et al., 1995; Stone et al., 1999) MTX effectively induced remission in 90% and 71–74% of patients respectively, but relapses occurred more frequently than when cyclophosphamide was used (70% vs 47%) (De Groot et al., 2005). MTX is excreted by kidney, therefore should be used with caution in those with renal impairment (Ntatsaki et al., 2011; Metzler et al., 2007).

De Groot et al conducted an unblended, randomized, controlled trial study to determine whether MTX could replace treatment with cyclophosphamide and oral corticosteroids (De Groot et al., 2005). They found that MTX can replace cyclophosphamide for initial treatment of early AAV, but it was less effective for induction of remission in patients with extensive disease or those with pulmonary involvement. In addition the relapse was more common in patients who treated with MTX than those who received cyclophosphamide. Patients were randomized to receive either MTX 7.5 mg/week increasing to 20 mg/week at 8 weeks or LEF loading dose 100 mg/day for 3 days, followed by 20 mg/day until Week 4, then 30 mg/day for 2 years. Their study showed no differences in efficacy or safety between two treatments. Pagnoux et al (Pagnoux et al, 2008) compared AZA with MTX in maintaining remission in WG and MPA patients who had achieved remission with intravenous pulse of cyclophosphamide. They found no significant difference in adverse events and relapses rates either during the 12-month treatment phase or subsequent follow-up between two groups.
Intravenous immunoglobulins (IVIG)

ANCAs can induce cytokine-primed neutrophils undergo degranulation and respiratory bursts during which they release toxic oxygen species and lytic enzymes (Falk et al., 1990). Anti-idiotypic antibodies with inhibitory effects on ANCA in vitro have been found in a pooled human gamma globulin preparation (Pall et al., 1994). Evidence showed that IVIG acts in different phases of the immune response including neutralization of circulating pathogenic antibodies, Fc receptor modulation, blockade or suppression of antibody-dependent cell toxicity, natural killer cell function, auto antibody production and complement activation, and acceleration of neutrophil apoptosis (Kazatchkine & Kaveri, 2001; Tsujimoto et al., 2002).

Beneficial effect of high-dose intravenous immunoglobulin in ANCA-associated vasculitis has been approved by different studies (Ito-Ihara et al., 2006; Tuso et al., 1992; Jayne et al., 1993; Richter et al., 1995). Ito et al evaluated IVIG monotherapy (400 mg/kg/day for 5 days) in AAV patients. Their study showed that IVIG decreased the leukocyte count, C-reactive protein level, Birmingham Vasculitis Activity Score and improved the systemic symptoms (Ito-Ihara et al., 2006). Other studies suggested that IVIG induces remission in 40–82% of patients (Richter et al., 1995; Jayne et al., 1991; Levy et al., 1999; Jayne et al., 2000). IVIG appears to have an important place in the management of ANCA-RPGN, but it's indications have not been determined. Because IVIG is an expensive drug additional studies on its cost-effectiveness and rational introduction into clinical practice are needed (Hotta et al., 2005).

Anti-B-cell therapy (Rituximab)

Beneficial effects of Rituximab which is a monoclonal antibody against anti-CD20 have been reported in several case series (Gottenberg et al., 2005; Omdal et al., 2005; Speckset al., 2001), and uncontrolled studies (Eriksson, 2005; Keogh et al., 2005a, 2005b; Smith et al., 2006). The main problem of treatment with rituximab is that relapses commonly occurred after 6–9 months (Keogh et al., 2005a, 2005b; Smith et al., 2006).

Apheresis therapies

Results of clinical trial of apheresis therapies, either plasmapheresis or cytapheresis in AAV are disappointing. These studies showed no benefit (Glöckner et al., 1988; Cole et al., 1992; Zäuner et al., 2002), benefits just in dialysis dependent patients (Pusey et al., 1991) or benefits on preserving the renal function (Furuta et al., 1998; Hasegawa et al., 2005). Pusey et al. found that plasma exchange is of added benefit in dialysis-dependent patients because in patients who were initially dialysis-dependent, renal function was more likely to have recovered when treated with plasma exchange plus drugs rather than drugs alone (Pusey et al., 1991). Report from the European community group suggested that adding plasma exchange to immunosuppressive therapy was not beneficial if there was severe tubular atrophy and fewer than 33% of the glomeruli were normal (De Lind van Wijngaarden et al., 1998).

In contrast to AAV in anti-GBM RPGN, the beneficial effect of plasma exchange has been well established. It might be attributable to the direct role of anti-GBM antibody in the pathogenesis of anti-GBM antibody RPGN, while in AAV no direct role for ANCA have been established (Hotta et al., 2005). The main advantage of Apheresis therapies is that no severe infectious episodes have been noted (Nagase et al., 1998; Sawada et al., 2003). Japan nation wide survey of RPGN (Yamagata et al., 2004) recommends cytapheresis in patients with aggressive forms of RPGN (rapid deterioration of renal function like the PR3-ANCA-
associated RPGN, or pulmonary renal syndrome complicated by severe inflammation, or relapses with high MPO-ANCA titer).

**Anti-TNF-alpha**

Insights into the role of Th1 cytokines in the pathogenesis of WG have led to trial therapies with antagonists of tumor necrosis factor-alpha (TNFα) and inhibitors of monocyte function, such as interleukin-10 (Kamesh et al., 2002). Etanercept (Enbrel; soluble receptors), infliximab (Remicade; human-mouse chimeric antibody against TNF), and adalimumab (Humila; human anti-TNF antibody) are biological antagonists of TNF (Ozaki, 2007). Etanercept have been reported ineffective in maintaining remission and a higher rate of malignancy have been noted in patients who have received the drug (Wegener's granulomatosis etanercept trial [WGET] research group, 2005). In an open label study, infliximab was added to standard immuno suppressive therapy in 16 patients with acute AAV at first presentation or relapse and in 16 with persistent disease despite multiple immuno suppressive regimens; 88% of patients achieved remission within a mean of 6.4 weeks (Booth et al., 2004).

**Anti-T cell antibodies**: Different studies have been reported that active systemic vasculitis is mediated in part by T cell-induced injury. This finding has led to the evaluation of anti-T cell antibodies in patients with Wegener's granulomatosis who are resistant to cytotoxic therapy (Lockwood et al., 1993; Hagen et al., 1995; Schmitt et al., 2004). In one study, the administration of a combination of two humanized monoclonal antibodies led to long-lasting remission in four patients with different forms of refractory vasculitis (Lockwood et al., 1993). In another study 15 patients with refractory disease received antithymocyte globulin (ATG) which resulted in a partial or complete remission in 9 and 4 patients, respectively (Schmitt et al., 2004). The role of these experimental therapies remains to be determined.

**Intravenous azathioprine**

High dose intravenous azathioprine has been tested for treating a variety of immune-mediated diseases. In one report, four patients with WG who had not responded to oral cyclophosphamide were treated with monthly infusions of azathioprine (Benenson et al., 2005). Two reached remission of disease, one of whom developed renal involvement during relapse, which responded to retreatment.

**15-Deoxyspergualin**

15-deoxyspergualin (gusperimus), a drug with anti-proliferative effect on antigen-stimulated B cells, has been evaluated in a small number of patients who didn't respond to cyclophosphamide or had contraindications to the use of cyclophosphamide. The administration of 15-deoxyspergualin in 20 patients resulted in complete or partial remission in six and eight respectively (Bircket et al., 2003). All patient experienced transient leucopenia with each treatment cycle. In another study seven patients treated with 15-deoxyspergualin and glucocorticoids, all reached complete or partial remission. The main problem was that to maintain remission prolonged treatment up to 4 years was required (Schmitt et al., 2005). In addition serial monitoring of white blood count is required to avoid excessive leucopenia.

**Radiation therapy**

Radiation therapy has been evaluated in patients with WG and airway involvement (Eagleton et al., 1979; Neviani et al., 2002). The use of ionizing radiation for non-malignant
disease is controversial. Current data do not support its use in systemic disorder like Wegener's granulomatosis (Stone et al., 2010).

**Stem cell transplantation:** High-dose, myeloablative chemotherapy with stem cell transplantation has been used for the treatment of refractory severe vasculitis. There are few case reports of successful treatment in patients with WG (Kötter et al., 2005). More studies are required to determine the role of stem cell transplantation in the management of resistant AAV.

**Prophylaxis against Pneumocystis carinii pneumonia (PCP)**

Opportunistic infections especially Pneumocystis carinii pneumonia are potentially fatal complications of immuno suppressive therapy in RPGN and AAV. The estimated incidence of PCP is approximately 6 percent (Ognibene et al., 1995). Different approaches to prophylaxis against PCP infection during initial immunosuppressive therapy have been suggested: trimethoprim-sulfamethoxazole one single strength (80 mg/400 mg) tablet daily or one double strength tablet (160 mg/800 mg) three times per week or Atovaquone in patients who are allergic to sulfonamides or do not tolerate trimethoprim-sulfamethoxazole. During treatment with methotrexate and glucocorticoids, the addition of trimethoprim-sulfamethoxazole increases the risk of pancytopenia, therefore Atovaquone is suggested for prophylaxis in such patients. It has been recommended to continue PCP prophylaxis in maintenance immuno suppressive therapy phase until the CD4-positive T cell count exceeds 300/microL (Mansharamani et al., 2000). Patients who have received trimethoprim-sulfamethoxazole for prophylaxis during induction, should continue the prophylaxis when azathioprine is used and switch to atovaquone when methotrexate is replaced for maintenance therapy.

Some patients have low CD4-positive T cell counts for prolonged periods after the cessation of cyclophosphamide and require prolonged PCP prophylaxis; in addition glucocorticoids should be tapered to the lowest possible dose. If patients develop neutropenia when receive prophylaxis, the drug should be switched to atovaquone (Stone et al., 2010).

Management of RPGN and AAV in pregnancy: As with active disease in non-pregnant patients, prednisone alone is relatively ineffective, and to induce remission combined therapy with cyclophosphamide is needed. The major challenges of treatment during pregnancy are potentially serious adverse effects which can occur with both MMF and cyclophosphamide. In addition, insufficient data about the safety of rituximab is available. High risk of skeletal and palatal defects, as well as malformations of the limbs and eyes has been noted in case of fetal cyclophosphamide exposure during the first trimester. Although fetal risk is much smaller during the second and third trimesters, pancytopenia and impaired fetal growth can occur. MMF fetal exposure increases the risk of miscarriage and congenital malformation such as cleft lip and palate. As a result, some consider MMF to be contraindicated in pregnancy. The safer immuno suppressive drugs that have been effective in WG and MPA include glucocorticoids, azathioprine, cyclosporine and tacrolimus. Alternatives that could be considered include cyclophosphamide or rituximab in the second or third trimester once organogenesis is complete (Stone et al., 2010).

### 6.1.1 Prognosis of RPGN and ANCA associated vasculitis

RPGN If left untreated typically progresses to end-stage renal disease over a period of weeks to a few months. Patients with fewer crescents may present slowly progressive...
Despite various immuno suppressive therapy protocols, mortality of ANCA positive RPGN is still high, and the major cause of death is infectious complications (Booth et al., 2003; De Lind van Wijngaarden et al 1998). Outcome of AAV depends on patient's age, degree of renal impairment at presentation and presence of pulmonary involvement. During the first 6 months of treatment mortality is very high as a result of aggressive course of the disease and toxic effects of early immuno suppressive treatments (Sakai et al 2002; Booth et al., 2003). Although the introduction of steroids and cyclophosphamide pulse therapy have improved the overall mortality of AAV, the 2-year mortality rate is still high (20% in 2-year follow-up) (Booth et al., 2003; Hogan et al., 1996; Falk et al 2000; Franssen et al 2000). Serum creatinine, dialysis dependency, and percentage of non-crescentic glomeruli at diagnosis have been considered as the best predictors of disease outcome (Bajema et al., 1999; Levy et al., 2001; Slot et al., 2003). The prognosis for patients with anti-GBM antibody disease is poor (Hirayama, et al., 2008), and renal survival and mortality rates of 20.9% and 23.3% at 6 months after onset have been reported respectively. Early diagnosis and starting treatment without delay might improve the prognosis (Hirayama, et al., 2008). A large nationwide survey of RPGN in Japan showed that mortality correlates with age, severity of renal dysfunction, presence of pulmonary involvement, and high C-reactive protein level (Sakai et al., 2002). The mortality rate of Japanese patients was higher than in European or American patients because of the high incidence of lethal infection (Hotta et al., 2005).

Japan Nationwide Survey of RPGN noted that 6-month renal and patients' survival for PR3-ANCA-associated RPGN were 88.2% and 92.3% respectively, while for MPO-ANCA they were 69.9% and 74.2% respectively (Yamagata et al., 2005). Patients' survival is very low in MPA if the disease presents as pulmonary renal syndrome (Gallagher et al., 2002; Lauque et al., 2000; Niles et al., 1996). Introduction of immuno suppressive agents considerably has improved the outcome of AAV over the past 30 years. WG and MPA if left untreated have a rapidly progressive and usually fatal course (Ntatsaki et al., 2011). Walton reported a mean survival of 5 months in patients with WG, and mortality rate of 82% and 89% in 1 and 2 year follow-up respectively (Walton, 1958). Standard treatments significantly have improved prognosis in WG and MPA, and some studies have reported 5-year survival rates of 81% for MPA and 87% for WG (Eriksson et al., 2009). In European Vasculitis Study, there was 11.1% mortality at 1 year (Little et al., 2010). High age at presentation, severe renal involvement (high serum creatinine level at presentation), pulmonary involvement, high ESR and high scores of disease activity and damage were poor prognostic factors (Holle et al., 2011).

Suzuki et al.'s study confirmed that ANCA-associated vasculitis is the most serious etiologies of RPGN (Suzukiet al., 2010). In nationwide survey by Yamagata and Koyama (Koyama et al., 2009; Yamagata et al., 2005), and Suzuki's study main causes of patients' death were infectious complications, including DIC which was mainly linked to pneumonia by opportunistic pathogens (Pneumocystis carinii, Candida albicans, and cytomegalovirus). Researchers hope they will find new immuno suppressive drugs with highest efficacy, lowest side effects and high safety during pregnancy to improve patients' survival and quality of life. It's a dream that undoubetly will be achieved in next years.

7. Conclusion

RPGN is a nephrology emergency which needs special attention. If the disease left untreated typically progresses to end-stage renal disease over a period of weeks to a few...
months. When there is a strong clinical suspicion special immuno suppressive treatment should be started as soon as possible (preferably after kidney biopsy). Despite various immuno suppressive therapy protocols mortality of ANCA positive RPGN patients is still high, also prognosis of anti-GBM antibody disease is poor. Actually treatment of RPGN and AAV are serious challenges in nephrology medicine which needs more clinical trial studies in larger groups of patients.

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9. References


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An Update on Glomerulopathies - Clinical and Treatment Aspects
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An Update on Glomerulopathies - Clinical and Treatment Aspects is a systemic overview of recent advances in clinical aspects and therapeutic options in major syndromes of glomerular pathology. The book contains twenty four chapters divided conveniently into five sections. The first section deals with primary glomerulopathies, and the second section is devoted to glomerulopathies complicating infectious conditions. The third section deals with systemic autoimmune disorders and vasculitides which constitute major causes of glomerular disease and often renal failure. The fourth section includes chapters discussing the glomerular involvement in some major metabolic and systemic conditions. The final section has chapters which relate to some general aspects of glomerular diseases. This book will form an excellent reference tool for practicing and academic nephrology community.

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